

Disentangling the response of fishes to recreational fishing over 30 years within a fringing coral reef reserve network

Abstract

While there are numerous studies that have used no-take marine reserves (NTMRs) to understand the effects of commercial fishing, much less is known about the influence of NTMRs in regions subject to recreational fishing. We used meta-analysis to synthesise data from 4,444 samples from 30 years (1987-2017) of fish surveys, inside and outside a large network of NTMRs in the Ningaloo Marine Park, Western Australia, where the major fishing activities are recreational. The data were collected by different agencies, using varied survey designs and sampling methods. We contrasted the relative abundance and biomass of target and non-target fish groups between fished and NTMR locations. We considered the influence of, and possible interactions between, seven additional variables: age and size of NTMR, one of two reserve network configurations, reef habitat type, the level of recreational fishing activity, shore-based fishing regulations and survey method. Taxa responded differently: commonly targeted lethrinids had clear higher average abundance and biomass inside NTMRs, while the abundance/biomass of other targeted (and non-targeted groups) were indistinguishable inside to outside. Reef habitat was an important factor governing lethrinid response to protection, and we showed this variable can interact with reserve size, with larger NTMRs only demonstrably more effective than smaller ones in the back reef/lagoon habitats. There was little evidence of changes in relative abundance/biomass with reserve age, or before and after rezoning and expansion of the NTMR network. Our study demonstrates the complexity of fish responses to protection and fishing, and also highlights some of the key factors and interactions that likely underlie the varied results found to date in NTMR assessments, and that should thus be considered in future reserve design and assessment.

Keywords

Marine protected area; MPA; fisheries; coral reef; Ningaloo; adaptive management; recreational fishing; *Lethrinus*

1. Introduction

Anthropogenic activities continue to expand worldwide, particularly in the tropics, threatening natural systems and the ecosystem services they provide (Barlow et al. 2018). As a result, ‘protected areas’ that seek to balance extractive activities with other socio-ecological values are increasingly being used to manage terrestrial and marine systems (Jenkins & Joppa 2009; Sala et al. 2018). Many studies have assessed the conservation effects of no-take marine reserves (NTMR) (reviewed by Mosquera et al. 2000; Russ 2002), including quantitative syntheses of regional and global studies, with most finding higher abundance and size of targeted species within reserve boundaries (Lester et al. 2009). The large majority of these findings are from regions with commercial fisheries operating, and less is documented about the impacts of recreational fisheries, despite several studies flagging the potentially high impacts of these fisheries (McPhee et al. 2002; Coleman et al. 2004; Cowx & Cooke 2004; Lewin et al. 2006). NTMRs are a key tool for assessing the impacts of fishing (Ballantine 2014) and while there are a handful of empirical studies that have demonstrated the effects of fishing, using inside outside comparisons, on targeted invertebrates (Shears et al. 2006; Babcock et al. 2007) and finfish (Denny et al. 2004) a comprehensive assessment including NTMR with different characteristics over long time frames is lacking. The magnitude of differences inside to outside reserves has been correlated with their design, in particular size and age, with larger and older reserves typically resulting in greater abundance and/or size of targeted fishes than reserves that are smaller or newly established (Claudet et al. 2008; Edgar et al. 2014). The effects of NTMRs vary among biomes, locations and taxa of interest (Côté et al. 2005; Claudet et al. 2010; Mora & Sale 2011) and there are examples of NTMRs having negligible effects on targeted fish communities (McLaren et al. 2015). In addition to size and age of NTMRs, explanations for this variability include high levels of cross-boundary movement by fishes (Pillans et al. 2014) and minimal to no difference in fishing activity across no-take and

52 fished areas due to accessibility and/or non-compliance by fishers (Bergseth et al. 2017), all of which make
53 disentangling the true effects of fishing more complicated.

54
55 Ideally assessments of the influence of NTMRs are based on replicated studies across multiple comparable
56 reserves with long time series of biological data before and after reserve establishment (Underwood 1993;
57 Russ 2002; Osenberg et al. 2011). Yet such data are typically beyond the scope of single research programs,
58 necessitating the integration of multiple datasets. 'Adaptive management,' involving changes to the number,
59 size or boundaries of reserves in response to new scientific information, changes in fishing pressure or
60 changing social attitudes (McCook et al. 2010) further complicates long-term assessments. Ongoing
61 improvement of ecological sampling methods and technologies has resulted in new survey methods being
62 introduced to monitoring (Goetze et al. 2015): video based methods (baited remote underwater video
63 (BRUV) and diver operated video (DOV)) are now commonly used alongside or in place of the previously
64 more common underwater visual census (UVC) (Mallet & Pelletier 2014). Therefore, evaluations of
65 NTMRs must have the capacity to incorporate and evolve with changes in reserve design and survey
66 methods (Claudet & Guidetti 2010). Other factors, including differences in habitat and benthic structure,
67 have been shown to affect outcomes of NTMR evaluation (Miller & Russ 2014; Rees et al. 2018b) and
68 while these factors have been studied independently, few assessments consider multiple factors
69 simultaneously, including possible interactions (Edgar et al. 2014). Differences in fishing pressure outside of
70 reserves will also directly impact inside to outside comparisons, yet data that quantify localised variation in
71 fishing activity at the scale of marine parks and typical NTMR networks are rarely available (Lewin et al.
72 2006).

73
74 Here, we synthesise a unique 30 year dataset from within a multiple-use marine park at Ningaloo Reef,
75 Western Australia. The type of fishing activity at Ningaloo Reef (almost exclusively recreational) in
76 combination with a highly regulated network of NTMRs that have undergone significant expansion during
77 the study period, offers the opportunity to advance on previous studies and inform on the potential impacts
78 of recreational fisheries. We integrate data from numerous agencies with varied survey designs and methods,
79 and therefore use a meta-analytical approach to compare the abundance and biomass of select targeted and
80 non-targeted tropical reef fish inside NTMRs with adjacent fished areas. We tested two hypotheses: (1) the
81 relative abundance/biomass of targeted fish taxa will be greater inside NTMRs than outside due to
82 recreational fishing activity; and (2) the observed relative abundance/biomass will vary with survey method,
83 age and size of NTMR, spatial variability in fishing activity (including shore-based fishing) and/or habitat.
84 Our study offers four main novelties. First, the effect of recreational fishing on targeted species is assessed
85 in isolation from commercial fishing. Second, we explicitly consider potential interactions between
86 variables. Third, the influence of changes in the NTMR network are considered in the context of the
87 increasingly common adaptive management. Fourth, we consider the influence of shore-based fishing,
88 which has rarely been investigated. We therefore provide advances on previous work that are of importance
89 for future planning and assessment of protected areas.

90 **2. Material and methods**

91 *2.1 Study region*

92
93 Data for this study are from the Ningaloo Marine Park (NMP) on the western Australian coastline (22°S,
94 113°E). The Park covers the majority of Ningaloo Reef (a World Heritage site) which is a fringing coral reef
95 almost 300 km in length. The reef encompasses a sheltered lagoon that is highly accessible by shore-based
96 fishers and those operating recreational vessels (Smallwood & Beckley 2012). Despite a relatively small
97 permanent human population, this area is a popular tourism destination for recreational fishers (Sumner et
98 al. 2002; Smallwood & Beckley 2012; Mitchell et al. 2018). There have not been any major commercial
99 fishing activities within the marine park since the 1970s, (for summary see pg. 78, CALM (2005) and pg.
100 70, DPIRD (2017)). Recreational spearfishing has additional restrictions of varying degrees outside of the
101 NTMRs, with spearfishing prohibited along a 70km stretch of coast between Tantabiddi Well and
102 Winderabandi Point, and spear fishing for Labridae and Serranidae prohibited throughout the Park (DPIRD
103 2018) (Fig. 1).

105
106 A network of eight NTMRs was established as part of the Park in April 1987 to cover 10% of the total
107 marine park area ~22,400 ha, (Fig. 1a) (CALM 1989). In 2005, the majority of the existing eight reserves
108 were expanded in size and 10 new reserves were added (Fig. 1b), increasing the NTMR coverage to 88,365
109 ha (34% of the NMP). At the same time, three NTMRs, covering 1,929 ha, were established as a part of the
110 28,616 ha Muiron Islands Marine Management Area (MIMMA), immediately adjacent to the northern
111 boundary of the NMP (CALM 2005). Together, the NMP and MIMMA form a continuous network (CALM
112 2005). There is some variation in the regulations along the boundaries of the 21 current NTMRs,
113 complicating terminology and analysis, with eight NTMRs allowing shore-based fishing from their coastal
114 boundaries (Appendix A, CALM 2005). According to recent classifications of marine reserves, the two
115 forms of reserves in the present study, those with shore-based fishing prohibited and those where it is
116 allowed, would classify as Fully Protected Areas and Highly Protected Areas respectively (Horta e Costa et
117 al. 2016), both of which would be expected to provide protection for fished species (Zupan et al. 2018). In
118 the present study we refer to all reserves as NTMRs for comparisons ‘inside’ to ‘outside’, though we
119 explicitly include consideration of the effect of shore-based fishing in our analyses.
120

121 2.2 Survey data

122 Data from all major research and monitoring programs surveying fish in the NMP over the last 30 years
123 (1987 – 2017) were collated (Appendix B) to create a very large synthesis of information. Locations of
124 individual samples are given as Appendix C. Three different survey methods were used to census fish:
125 Baited Remote Underwater stereo-Video (BRUV), Diver Operated stereo-Video (DOV) and Underwater
126 Visual Census (UVC) (Langlois et al. 2010; Murphy & Jenkins 2010). The majority (90%) of surveys also
127 estimated the length of fish (an *in situ* estimate of total length for UVC, and fork length measured from
128 stereo-video for DOV and BRUV), which allowed estimates of biomass using formulas from FishBase
129 (Froese 2018) (Appendix B).
130

131 Data were organised hierarchically with a sample (individual transect or a BRUV deployment) being the
132 lowest level of replication. Samples were classified to the next hierarchical level and termed a ‘comparison
133 pair’, based on the criteria: (i) that there were at least two samples inside and two samples outside a given
134 NTMR, (ii) these samples were collected within 2 weeks of each other, (iii) samples were collected more
135 than 200 m from within or outside of the reserve boundaries (excluding one reserve, the small size of which
136 meant this was not a logical rule), (iv) samples were collected using the same survey method within one of
137 four *habitat* categories (see Table 1). Data satisfying these conditions consisted of 4,444 samples classified
138 into 305 relative abundance comparison pairs and 3,892 samples classified into 268 relative biomass
139 comparison pairs. These data covered seven of the initial eight NTMRs and 16 of the 21 current NTMRs
140 (Appendix B).
141

142 2.3 Fish groups

143 We consider three main fish groups common at Ningaloo Reef, at family/subfamily and species level
144 (Appendix D) which differ in terms of their behaviour and representation in fisheries catch reports. This
145 included: parrotfishes (Scarinae), which are not typically targeted by fishers in Australia, and two groups
146 which are highly targeted by recreational fishers in the region (Ryan et al. 2013; Ryan et al. 2015) that have
147 different behaviours; emperors (Lethrinidae; mobile roving predators) and groupers (Epinephelinae; mostly
148 site-attached ambush predators). Previous work has indicated both Epinephelinae and Lethrinidae are
149 vulnerable to fishing and many species in both subfamilies are targeted across the Indo-Pacific (Abesamis et
150 al. 2014). Species level analyses included two species from Lethrinidae: the spangled emperor, *Lethrinus*
151 *nebulosus*, which is recognised as the most highly targeted species in the region, consistently featuring at the
152 top of the estimated catch for the bioregion over the 30-year study period, and the yellow-tailed emperor, *L.*
153 *atkinsoni*, a species that is anecdotally retained by fishers and featured as the 6th most common species
154 recorded in the 1998/9 catch survey, but was a minor component in subsequent surveys (Sumner et al. 2002;
155 Ryan et al. 2013; Ryan et al. 2015). The Chinaman Rockcod, *Epinephelus rivulatus* (Epinephelinae) was
156 also considered, with catches comparable to those of *L. nebulosus* across the catch reports (Ryan et al. 2013;

157 Ryan et al. 2015). Individual species were not considered from the Scarinae subfamily due to inconsistencies
158 in the accuracy of identification of species from this family.
159

160 161 2.4 Meta-analysis

162 We used a mixed-effects meta-analytical approach to assess the effect of NTMRs on fish abundance and
163 biomass. We calculated effect sizes as log-ratios for each of the comparison pairs inside to outside the
164 NTMRs (Claudet et al. 2008) (see Appendix E for formulas). A constant was added to the mean abundance
165 ($c = 0.5$) and mean biomass ($c = 100$ g) to allow calculation of the log ratio in cases where fish were absent
166 either inside or outside (i.e. zero values). We ran a sensitivity analysis on the value of the constant
167 (Appendix F) to determine an appropriate value. The size of the constant impacted the magnitude of the
168 effect size, but in general did not influence the significance. Nonetheless, the exact magnitude of the overall
169 effect size should be interpreted with caution. In cases where both the inside and outside mean count were
170 zero, the samples were excluded from the analysis. Effect sizes were weighted by the inverse of the sum of
171 the within- and among-study variances (Appendix E). Weighted effect sizes and variances were calculated
172 using the *metafor* package (Viechtbauer 2010) in the statistical program R (R Core Team 2017) with the
173 variance estimator set to “REML,” restricted maximum likelihood estimator. Overall effect sizes were
174 comparable for both abundance and biomass and for simplicity we presented the abundance results as these
175 were available for a larger dataset, providing biomass results in Appendix H.
176

177 2.5 Sources of variability

178 We considered seven variables that might mediate the response of fish abundance/biomass to the presence of
179 the NTMRs (Table 1): (i) the number of years between when a sample was collected and when the zoning
180 went into place; (ii) initial or current zoning scheme (see Figure A1); (iii) survey method; (iv) four coarse
181 *habitats* with distinct coral/algae assemblages, exposed reef slope, reef flat, back reef/lagoon coral, and
182 lagoon algae; (v) spatial area of a reserve; (vi) an estimate of fishing pressure outside of individual reserves;
183 (vii) regulations on shore-based fishing. Data were explored following the protocol of Zuur et al. (2010) and
184 transformed to normalise their distribution where appropriate (see Table 1).

185 As all effect sizes were heterogeneous (Appendix G), we first explored the influence of the seven variables
186 using weighted mixed-effects categorical meta-analyses and meta-regression, considering each variable as a
187 moderator in isolation to determine which variables explained significant heterogeneity in the overall effect
188 size (see Appendix E for formulas). We also investigated *reserve identity* to allow comparison between
189 individual NTMRs. Given there were correlations among the variables and potential interactions and non-
190 linear effects, we used weighted full-subsets generalised additive mixed modelling (*FSSgam*) (Fisher et al.
191 2018 A simple function for full-subsets multiple regression in ecology with R. *Ecology and Evolution*) to
192 investigate the relative importance of each variable in explaining variability in the overall effect size for
193 each fish group. The response variable, effect size e , was modelled with a Gaussian distribution using *gam()*
194 in the *mgcv* package in R (Wood 2011). *Years protection* and *boat fishing* were included as continuous
195 smoothers in the *FSSgam* to allow for non-linear relationships. The distribution of *reserve size* was not much
196 improved by transformation and $\sqrt{\text{reserve size}}$ was therefore included in the model set as a linear
197 predictor. *Reserve identity* was highly collinear with other variables (in particular *reserve size*), and
198 therefore, rather than including this as a random effect, a smoother of the mean *latitude* of comparison pairs
199 was included in all models (and as part of the null model). This yielded comparable results to including
200 *reserve identity* as a random effect. Interactions between factor variables *habitat* and *shore fishing* and the
201 continuous variables *reserve size* and *years protection* were tested. In all models the smoothing parameter
202 was limited to a simple spline, allowing only monotonic relationships ($k=3$) for all continuous variables
203 except for *latitude*, which was unlimited. Summed AICc weights were used as a metric of variable
204 importance to investigate the relative importance of each predictor variable across the full set of models
205 (Anderson & Burnham 2002). Variables included in the most parsimonious model (fewest variables and
206 lowest estimated degrees of freedom within two units of the AICc) were plotted to visualise the shape and
207 direction of relationships between the variables and the effect size. AICc can overfit models (REF) so we

interpret results of variable importance and the top models with caution and consider the results of the mixed-effects meta-analyses and meta-regression alongside the results of the *FSSgam*.

Lastly, given the importance of temporal patterns in investigations of protected areas, we explicitly investigate data from the Osprey NTMR (see Fig. 1), the best temporally replicated NTMR in the dataset. Using available and relatively consistently collected UVC and DOV data we estimated mean fish density as count per transect area. We tested for significant linear and quadratic relationships between the density of *L. nebulosus* and survey year and fitted generalised additive mixed models.

3. Results

When compared to areas open to fishing, Lethrinidae were on average 57% more abundant (78% more biomass) inside NTMRs ($e = 0.45 \pm 0.12$, 95%CI, Fig. 2a), however the effect was heterogeneous ($Q_T = 2002.6$, $df = 301$, $p < 0.001$, Table G1). The most parsimonious model for Lethrinidae abundance consisted of an interaction between *habitat* and *reserve size* (Table 2), with the same true for biomass (Appendix H). The categorical meta-analysis supported the importance of *habitat* for relative abundance; showing it explained significant heterogeneity among effect sizes ($Q_M = 39.5$, $df = 3$, $p < 0.001$, Table G2) with the most positive effect identified in *back reef & lagoon coral* sites with an average of 93% more Lethrinidae inside the NTMRs ($e = 0.66 \pm 0.14$, 95%CI) (Fig. 2a, Fig. G1) in this habitat. On the *reef flat* Lethrinidae were 53% more abundant inside the NTMRs ($e = 0.42 \pm 0.32$, 95%CI) while there was no significant effect on the *exposed reef slope* and a negative effect in the *lagoon algae* habitat (Fig. G1). The interaction of *reserve size* and *habitat* was evident as an increase in effect size with increasing *reserve size* in the *back reef & lagoon coral* habitat versus no clear trends in the other habitats.

Lethrinus nebulosus were on average 42% more abundant (86% more biomass) inside NTMRs than outside ($e = 0.35 \pm 0.15$, 95%CI, Fig 2a). The effect was heterogeneous ($Q_T = 1971.1$, $df = 256$, $p < 0.001$, Table G1). The most parsimonious model included the interaction between *habitat* and *reserve size* with these two variables also having the highest variable importance across the full-subsets model set (Table G3, Fig. 2b). The same was true in the biomass analysis (Appendix H). *Habitat* explained significant heterogeneity for relative fish abundance ($Q_M = 32.5$, $df = 3$, $p < 0.001$, Table G2) and *L. nebulosus* were on average 84% more abundant within *back reef & lagoon coral* sites inside the NTMRs ($e = 0.61 \pm 0.17$, 95%CI), whereas no differences were observed for the *reef flat* or *exposed reef slope* sites and a negative effect was observed for *lagoon algae* sites (Fig. G1). As for Lethrinidae, the interaction of *reserve size* and *habitat* was evident by an increase in the effect size with increasing *reserve size* in the *back reef & lagoon coral* habitat and no clear effects in the other habitats.

On average, the abundance of *L. atkinsoni* was 40% more abundant (60% more biomass) inside NTMRs than outside ($e = 0.34 \pm 0.09$, 95%CI). The effect was heterogeneous ($Q_T = 1739.7$, $df = 279$, $p < 0.001$, Table G1). The most parsimonious model included *zoning scheme* and *method*, which also had the highest importance according to weighted AICc. These two variables explained significant heterogeneity according to the categorical mixed-effects meta-analyses. Predictions indicated that the BRUV *method* contributed the most to the positive effect size of *L. atkinsoni* (Fig. 3c), though this was not significant, nor were the differences between initial and current zoning, showing a slightly higher effect size from the older zoning scheme. Multiple variables explained significant heterogeneity for *L. atkinsoni* according to the categorical meta-analysis and the meta-regression (Table G2), including *habitat* ($Q_M = 14.6$, $df = 3$, $p < 0.001$, Table G2). *Reef flat* sites had 94% higher abundance, ($e = 0.66 \pm 0.26$, 95%CI) and *back reef & lagoon coral* sites 43% higher abundance ($e = 0.36 \pm 0.12$, 95%CI) inside the NTMRs. There were no significant effects for the other habitats (Fig. G1). The biomass analysis for *L. atkinsoni* indicated that *years protection* may interact with *habitat*, and that on the reef flat the effect size was higher and showed a parabolic pattern with *years protection* (Fig. H2).

The effect size for Epinephelinae abundance was significantly negative with 9% fewer fishes inside than outside the NTMRs ($e = -0.09 \pm 0.08$, 95%CI), although this result was heterogeneous ($Q_T = 1125.7$, $df =$

276, $p < 0.001$, Table G1). Variable importance scores showed no variables with high importance relative to the Lethrinidae and *L. nebulosus* model sets. *Reserve size* and *years protection* were present in the most parsimonious model, while for the biomass it was *method* and *boat fishing* (Appendix H). There were weak increasing trends for both *reserve size* and *years protection*, however the lack of strongly important or consistent variables in these model sets means the results should be interpreted cautiously.

On average there was no significant difference inside to outside the reserves for *E. rivulatus* abundance ($e = -0.06 \pm 0.09$, 95%CI), though the effect was heterogeneous ($Q_T = 477.3$, $df = 166$, $p < 0.001$, Table G1). *Zoning scheme* and *boat fishing* had the highest variable importance across the model set and featured in the most parsimonious model. The effect size transitioned from no effect for low boat fishing activity, to a positive effect when there was high boat fishing activity, but the confidence intervals did not show this trend to be significant. The initial reserve network (in place longer) had a more positive effect than the newer reserves, but again this was not significant (Fig. 3e).

The control fish group, Scarinae, showed no significant difference inside to outside NTMRs ($e = -0.01 \pm 0.11$, 95%CI) and this effect was heterogeneous ($Q_T = 1701.1$, $df = 260$, $p < 0.001$, Table G1). All variables had low importance according to AICc (Fig 2b, Table 2) and while *boat fishing* and *shore fishing* appear in the most parsimonious model we interpret this with caution. In the biomass analysis *habitat* made up the most parsimonious model (Appendix H).

The temporal investigation of effect sizes for the most highly targeted fish, *L. nebulosus*, at Osprey NTMR did not show strong or significant patterns with time, except for the abundance density outside of the NTMR which had a significantly negative linear trend (Fig 4). These results generally confirm what was found for the full meta-analysis for *L. nebulosus*, showing effect sizes that are mostly positive across time, with higher abundance inside than outside. Trends, particularly in the latter half of the study period indicate that both abundance and biomass may have declined both inside and outside the NTMR, while there is some indication that abundance initially increased inside of the NTMR following establishment, however confidence in these trends is low.

4. Discussion

Across the 30 year synthesis higher abundance and biomass of certain targeted fish taxa inside NTMRs suggests that recreational fishing can have significant effects in isolation from commercial harvest, as also shown in some previous studies (Denny et al. 2004; Shears et al. 2006; Babcock et al. 2007). We found the extent of this effect was variable among targeted taxa and influenced by a range of other factors. While our analyses revealed higher relative abundance and biomass of lethrinids (*Lethrinus nebulosus* and *L. atkinsoni*) inside NTMRs, no significant effect was found for the abundance of *Epinephelus rivulatus*, and a small negative effect was detected for the epinephelids as a group. All effects were heterogeneous, which was not surprising given the size and complexity of the synthesised dataset (including differences in size and age of reserves) and given that fish responses to NTMRs are known to vary with taxon-specific, ecological and zoning factors (Barrett et al. 2007; Claudet et al. 2010; Edgar et al. 2014). Here we advance previous findings with the largest meta-analysis on recreational fishing in isolation from commercial fishing, illustrating the new information that can be gained from synthesising existing data, though we do not discount the advantages of strategic and consistent monitoring data. We show that it is important for assessments of NTMRs to take into account habitat effects, and potential interactions with factors such as reserve size or age, as well as variability in fishing activity and subtle elements of reserve design (e.g. partial fishing restrictions) in order to avoid oversimplified conclusions on how fish abundance/ biomass respond to management.

Recreational fishing, specifically angling, is the only major fishing activity within the marine park. Some previous studies in the Park have linked higher abundance/biomass of targeted species inside NTMRs to protection from fishing (Westera 2003a; Babcock et al. 2008; Fitzpatrick et al. 2015); though results of other

313 studies are more equivocal (Wilson et al. 2012; Wilson et al. 2018b). The reasons behind the disparate
314 conclusions are unclear, but may be due to limited and/or varied spatial and temporal scales of the individual
315 studies, different survey methodologies, the confounding influence of habitat, or high variability in target
316 species abundance distributions. High correlation between fish recruitment and larger natural cycles (El
317 Niño Southern Oscillation) has also been suggested as a reason for these inconsistencies (Wilson et al.
318 2018a). In the present study we found high variability in the relative fish abundances of lethrinids among the
319 different NTMRs, which can at least partly account for the varied conclusions of previous studies of smaller
320 spatial scales (Fig. G2). Nonetheless, when all data were pooled the average effect was clearly positive for
321 abundance and biomass of the three lethrinid groups. The magnitudes of the positive effects were small
322 (max 57% higher inside) relative to studies in other parts of the world (Watson & Ormond 1994; Russ et al.
323 2015). A significant positive response for *L. atkinsoni* (40% higher), similar to that of *L. nebulosus* (42%)
324 was not expected, given *L. atkinsoni* does not feature highly in catch reports (Ryan et al. 2017), suggesting it
325 may be more susceptible to recreational angling that previously recognised.

326
327 Known differences in behaviour between lethrinid and serranid taxa did not correlate with their response to
328 NTMR as expected. Lethrinids are known to have large home ranges relative to many epinephelids,
329 including *E. rivulatus*, and are therefore more likely to move across NTMR boundaries (Mackie & Black
330 1999; Pillans et al. 2014; Babcock et al. 2017), with the expectation that they may experience lower levels of
331 protection than epinephelids. However, we only observed positive responses for the lethrinids. It is possible
332 that higher counts of lethrinids than epinephelids in the dataset may have reduced the power to detect an
333 effect in the latter group, or there are other factors that have not been captured in our analyses.

334
335 The age of no-take reserves has been shown to be a significant positive correlate of relative fish abundance
336 for targeted species (Claudet et al. 2008; Edgar et al. 2014; Zupan et al. 2018) and demonstrated increases in
337 effect size with time help attribute positive effect sizes to the presence of a protected area, rather than other
338 factors (Russ et al. 2015). In the present study there was negligible evidence of changes in effect sizes with
339 age of reserve. Where relationships were present, the shape of the trend was generally parabolic, showing an
340 increase initially, before subsequent decrease around 2005, though no relationships were significant. This
341 was supported by examining data for *L. nebulosus*, from the best temporally replicated NTMR, Osprey,
342 where again no clear temporal patterns were found. Potentially of concern for managers was the
343 significantly negative decline in *L. nebulosus* density outside of the Osprey NTMR, and a slight increase
344 followed by a decrease inside this NTMR. However the confidence intervals on all temporal patterns were
345 large. These findings are in contrast with previous studies, for example Russ et al. (2015) showed lethrinids
346 continued to increase in density inside NTMRs in the Philippines on time scales of 8-30 years. In the present
347 study rezoning in 2005 made temporal analyses more complex, though by including *zoning scheme* as a
348 variable we partly addressed this. Effect sizes were not strongly influenced by this variable, implying that
349 the effect sizes were broadly consistent across the initial and current NTMR networks. Where *zoning scheme*
350 did feature for *L. atkinsoni*, the older reserves had a more positive effect, as expected.

351
352 The absence of a strong temporal link with effect size must be considered when interpreting the positive
353 effect sizes, however there are various factors which may have contributed to the absence of a strong
354 relationship. First, while there is limited evidence of a reduction in fishing activity within the Park (Ryan et
355 al. 2015, 2017) a shift in fishing activity to areas offshore (>100m depth) (West et al. 2015; Mitchell et al.
356 2018), which are not part of the current survey data, is likely. Second, the mobile behaviour of lethrinid
357 fishes may be capping the levels of the observed effect size, if a proportion of their population is travelling
358 further than the NTMR boundaries. Pillans et al. (2014) found that approximately 60% of lethrinid
359 individuals move at scales greater than the average NTMR size over a year period. Third, illegal fishing
360 within the reserves may also limit a temporal increase in effect size, as Smallwood and Beckley (2012)
361 found 8-12% of observed vessels were fishing inside NTMRs in the Park in 2007. Third, we do not discount
362 that the unevenness of sampling through time, with some years being more highly sampled than others (Fig.
363 B2) potentially influenced our capacity to detect a trend if it were present. The analysis of *L. nebulosus*
364 density at Osprey showed that the temporal patterns inside and outside NTMRs can be complex and not
365 always captured by the overall effect size. Parallel declines or increases in density occurring both inside and

366 outside are masked from the effect size, and such declines have been observed in other fisheries closures on
367 the western Australian coast (Bornt et al. 2015).

368
369 Though our study only had a very coarse level of habitat classification available, our results support
370 previous studies (Miller & Russ 2014; Rees et al. 2018a; Rees et al. 2018b), showing the importance of
371 habitat when assessing the ability of NTMRs to support target species abundance. We further demonstrate
372 interactions between habitat and reserve size, showing that conclusions on both the magnitude and direction
373 (positive or negative) of observed effects for the relative abundance of Lethrinidae and *L. nebulosus* are
374 influenced by this interaction. In the case of *L. atkinsoni* biomass we also found an interaction between
375 habitat and reserve age, though the models were not as strong. Previous studies have demonstrated the
376 positive influence of larger and older reserves (Halpern & Warner 2002; Claudet et al. 2008; Edgar et al.
377 2014; Zupan et al. 2018), however the interaction with habitat has not previously been explored.
378 Furthermore, it is noteworthy that effect sizes were greatest in the *back reef & lagoon coral* habitat for *L.*
379 *nebulosus*, while for *L. atkinsoni*, the effect was greatest on the *reef flat*, a result that may be attributed to
380 these habitats being preferred by the adults of each species respectively (Babcock et al. 2008; Wilson et al.
381 2017). This is important when considering potential changes to habitat inside or outside of reserves, as Russ
382 et al. (2015), showed that changes in benthic habitat due to disturbance could markedly influence the effect
383 of NTMRs for lethrinids. We advise that NTMRs must incorporate adequate amounts of the essential
384 habitats of the species or communities they are designed to protect, and assessment of NTMR effectiveness
385 must account for possible interactions between habitat and reserve size and age.

386
387 While *habitat* was particularly important for the lethrinid groups, it was not found to be an important
388 predictor for Epinephelinae or *E. rivulatus*. Again, this was contrary to expectations given the often high site
389 fidelity of Epinephelinae (Mackie & Black 1999). However, the relatively coarse habitat classification
390 available for our analyses likely did not adequately capture the habitat requirements for this group. Previous
391 work has shown *E. rivulatus* is strongly associated with macroalgal habitats at Ningaloo Reef (Wilson et al.
392 2012) but that variability in the quality of macroalgal habitats can be substantial and have major implications
393 for fish abundance (Fulton et al. 2014; Wilson et al. 2014; Lim et al. 2016). Furthermore, Beckley and
394 Lombard (2012) found that deeper habitats seaward of the reef have relatively lower spatial protection from
395 recreational fishing, despite these habitats potentially supporting a high biomass of epinephelids (Babcock et
396 al. 2008). It is thus plausible that habitats outside of the NTMRs were more appropriate for Epinephelinae,
397 particularly prior to re-zoning in 2005, which could explain the overall negative and null effects for these
398 groups. A much better understanding of the habitat requirements, electivity and movement across seascapes
399 by targeted taxa and appropriate ‘micro-habitat’ classifications are needed to more fully understand these
400 results.

401 Where the boat fishing variable appeared in models for *E. rivulatus*, there were subtle positive trends in
402 effect size as fishing activity increased, i.e. where boat fishing was most prevalent the effect size was
403 greater. Our metric for fishing activity is unlikely to be representative across the 30 years of data, as it was
404 an estimate from 2007 (Smallwood & Beckley 2012), yet still showed some importance. We think this is a
405 particularly important factor when assessing NTMRs, as variability in fishing activity (spatially and
406 temporally) makes it very difficult to disentangle the true effect of the NTMRs if this variability is not
407 quantified. We suggest that finer-scale spatiotemporal data on the pressures outside, and indeed inside, of
408 reserves would clarify NTMR assessments, both in the case of the present study but also more generally in
409 any assessment of spatial protection. In the case of marine reserves, quantitative standardised data on fishing
410 activity at the scale of individual NTMRs should be prioritised alongside the collection of ecological data.

411
412 Synthesizing data from multiple survey methods leads to larger datasets, and the advent of video-based
413 methods in the last decades (e.g. BRUV and DOV) has increased the diversity of methods used to monitor
414 fish. Contrary to expectations, in general, survey method did not strongly influence the effect size. The
415 strongest effect sizes (Lethrinidae and *L. nebulosus*) were consistently detected regardless of the survey
416 method. *L. atkinsoni* exhibited a more positive effect when surveyed by remote video as compared to diver-
417 based methods, which may be partly explained by fish behaviours associated with both the attraction to bait
418 and avoidance of divers (Watson et al. 2005; Goetze et al. 2015), particularly on SCUBA (Radford et al.

2005). On balance, we did not distinguish a single survey method as optimal, and in most cases it was appropriate to compare data from the three methods for the effect size calculation. This is likely possible because of the nature of our effect size, which, as a ratio, is more robust to different units of measurement. However, this cannot provide the same level of information as standardised temporal data on fish density inside to outside, as shown by density patterns inside and outside at Osprey, underlying the overall effect size for this NTMR. We therefore suggest that monitoring programs should prioritise resurveying existing monitoring sites with comparable methodology to build more robust time-series data, else adopt the method(s) that are best suited to surveying the taxa of interest.

5. Conclusions

We have presented a comprehensive meta-analysis of NTMR assessments using a variety of survey methods over 30 years as a case study to show what can be learnt from collaboration and the synthesis of disparate data spanning adaptive management changes. Adaptive management is likely to become more common, indeed at Ningaloo, a new Australian Marine Park, in commonwealth waters directly seaward of the Ningaloo Marine Park of the present study has recently been implemented (1/1/18). Our findings suggest that consistent monitoring producing data that can be compared to that of the present study should be implemented for this new Park, across different habitats. More broadly, we suggest that comparisons inside to outside NTMRs, and indeed protected areas in general, can be improved by accounting for (i) habitat; (ii) potential interactions between habitat and reserve size and age; and (iv) variability in fishing activity outside of reserves and compliance inside reserves. While there is much to learn by synthesising existing data, there are important limitations in the ability of meta-analytical techniques to provide quantification of temporal changes to overall fish populations, and here we found no strong evidence of temporal in effect size. Using two diver-based methods we showed that the patterns in estimated fish density inside and outside and reserves can be complex and not fully captured by effect sizes. Therefore, there is a strong need to make monitoring more meaningful by ensuring data are being consistently recorded across time and space. Further to this, integration of the collection of fishing activity data with the collection of ecological data is likely to help interpret the true effects of NTMRs. The two are clearly intertwined and having data on both the pressure and the response is essential for holistic assessments of the efficacy of spatial management interventions. A key message for managers designing or adapting NTMRs is the need to account for the preferred habitat of the species of interest and how this may interact with NTMR size or age. Targeting relevant habitats, refining measures of fishing activity and standardising methodologies will vastly improve our ability to assess the effects of NTMRs.

Appendices

Appendix A – Additional information on individual reserves and rezoning

Appendix B – Data summary

Appendix C – Geographic locations of individual surveys

Appendix D – Information on fish groups

Appendix E – Formulas used for calculating effect sizes

Appendix F – Sensitivity analysis for cases of one-armed zero events

Appendix G – Meta-analysis statistics

Appendix H – Biomass results

References

- Abesamis RA, Green AL, Russ GR, Jadloc CRL. 2014. The intrinsic vulnerability to fishing of coral reef fishes and their differential recovery in fishery closures. *Reviews in Fish Biology and Fisheries* **24**:1033-1063.
- Anderson DR, Burnham KP. 2002. Avoiding pitfalls when using information-theoretic methods. *The Journal of Wildlife Management*:912-918.
- Babcock R, Haywood M, Vanderklift M, Clapin G, Kleczkowski M, Dennis D, Skewes T, Milton D, Murphy N, Pillans R. 2008. Ecosystem Impacts of Human Usage and the Effectiveness of Zoning for

472 Biodiversity conservation: Broad-scale Fish Census. Final Analysis and Recommendations 2007.,
473 Hobart.

474 Babcock R, Phillips J, Lourey M, Clapin G. 2007. Increased density, biomass and egg production in an
475 unfished population of Western Rock Lobster (*Panulirus cygnus*) at Rottne Island, Western
476 Australia. *Marine and Freshwater Research* **58**:286-292.

477 Babcock R, Pillans R, Rochester W. 2017. Environmental and individual effects on the behaviour and
478 spawning movements of *Lethrinus nebulosus* on a coral reef. *Marine and Freshwater Research*
479 **68**:1422-1437.

480 Barlow J, França F, Gardner TA, Hicks CC, Lennox GD, Berenguer E, Castello L, Economo EP, Ferreira J,
481 Guenard B. 2018. The future of hyperdiverse tropical ecosystems. *Nature* **559**:517.

482 Barrett NS, Edgar GJ, Buxton CD, Haddon M. 2007. Changes in fish assemblages following 10 years of
483 protection in Tasmanian marine protected areas. *Journal of Experimental Marine Biology and*
484 *Ecology* **345**:141-157.

485 Beckley LE, Lombard AT. 2012. A systematic evaluation of the incremental protection of broad-scale
486 habitats at Ningaloo Reef, Western Australia. *Marine and Freshwater Research* **63**:17-22.

487 Bergseth BJ, Williamson DH, Russ GR, Sutton SG, Cinner JE. 2017. A social–ecological approach to
488 assessing and managing poaching by recreational fishers. *Frontiers in Ecology and the Environment*
489 **15**:67-73.

490 Bornt KR, McLean DL, Langlois TJ, Harvey ES, Bellchambers LM, Evans SN, Newman SJ. 2015. Targeted
491 demersal fish species exhibit variable responses to long-term protection from fishing at the Houtman
492 Abrolhos Islands. *Coral Reefs* **34**:1297-1312.

493 CALM. 1989. Ningaloo Marine Park Management Plan 1989 - 1999. Perth, WA, Australia.

494 CALM. 2005. Management Plan for the Ningaloo Marine Park and Muiron Islands Marine Management
495 Area 2005-2015: Management Plan No 52. Perth, WA, Australia.

496 Cinner JE, Maire E, Huchery C, MacNeil MA, Graham NA, Mora C, McClanahan TR, Barnes ML,
497 Kittinger JN, Hicks CC. 2018. Gravity of human impacts mediates coral reef conservation gains.
498 *Proceedings of the National Academy of Sciences*:201708001.

499 Claudet J, Guidetti P. 2010. Improving assessments of marine protected areas. *Aquatic conservation: marine*
500 *and freshwater ecosystems* **20**:239-242.

501 Claudet J, Osenberg C, Domenici P, Badalamenti F, Milazzo M, Falcón JM, Bertocci I, Benedetti-Cecchi L,
502 García-Charton JA, Goñi R. 2010. Marine reserves: fish life history and ecological traits matter.
503 *Ecological applications* **20**:830-839.

504 Claudet J, Osenberg CW, Benedetti-Cecchi L, Domenici P, García-Charton JA, Pérez-Ruzafa Á,
505 Badalamenti F, Bayle-Sempere J, Brito A, Bulleri F. 2008. Marine reserves: size and age do matter.
506 *Ecology letters* **11**:481-489.

507 Collins LB, Zhu ZR, Wyrwoll K-H, Eisenhauer A. 2003. Late Quaternary structure and development of the
508 northern Ningaloo Reef, Australia. *Sedimentary Geology* **159**:81-94.

509 Côté IM, Mosqueira I, Reynolds JD. 2005. Effects of marine reserve characteristics on the protection of fish
510 populations: a meta-analysis. *Journal of Fish Biology* **59**:178-189.

511 Cowx IG, Cooke SJ. 2004. The Role of Recreational Fishing in Global Fish Crises. *BioScience* **54**:857-859.

512 Denny CM, Willis TJ, Babcock RC. 2004. Rapid recolonisation of snapper *Pagrus auratus*: Sparidae within
513 an offshore island marine reserve after implementation of no-take status. *Marine Ecology Progress*
514 *Series* **272**:183-190.

515 DPIRD DoPIaRD. 2018. Ningaloo Marine Park, Available from
516 [https://parks.dpaw.wa.gov.au/sites/default/files/downloads/parks/2462-](https://parks.dpaw.wa.gov.au/sites/default/files/downloads/parks/2462-13%20Spearfishing%20in%20Ningaloo%20WEB.pdf)
517 [13%20Spearfishing%20in%20Ningaloo%20WEB.pdf](https://parks.dpaw.wa.gov.au/sites/default/files/downloads/parks/2462-13%20Spearfishing%20in%20Ningaloo%20WEB.pdf) (accessed 4/11/2018 2018).

518 DPIRD DoPIaRDWA. 2017. Gascoyne Coast Bioregion, Status reports of the fisheries and aquatic resources
519 of Western Australia 2016/17.

520 Edgar GJ, Stuart-Smith RD, Willis TJ, Kininmonth S, Baker SC, Banks S, Barrett NS, Becerro MA, Bernard
521 AT, Berkhout J. 2014. Global conservation outcomes depend on marine protected areas with five key
522 features. *Nature* **506**:216-220.

523 Fisher R, Wilson Shaun K, Sin Tsai M, Lee Ai C, Langlois Tim J. 2018. A simple function for full-subsets
524 multiple regression in ecology with R. *Ecology and Evolution* **0**.

- 525 Fitzpatrick B, Harvey E, Langlois T, Babcock R, Twiggs E. 2015. Effects of fishing on fish assemblages at
526 the reefscape scale. *Marine Ecology Progress Series* **524**:241-253.
- 527 Froese RaDP. 2018. FishBase, Available from www.fishbase.org (accessed 15/1/2018 2018).
- 528 Fulton CJ, Depczynski M, Holmes TH, Noble MM, Radford B, Wernberg T, Wilson SK. 2014. Sea
529 temperature shapes seasonal fluctuations in seaweed biomass within the Ningaloo coral reef
530 ecosystem. *Limnology and Oceanography* **59**:156-166.
- 531 Goetze J, Jupiter S, Langlois T, Wilson S, Harvey E, Bond T, Naisilisili W. 2015. Diver operated video most
532 accurately detects the impacts of fishing within periodically harvested closures. *Journal of*
533 *Experimental Marine Biology and Ecology* **462**:74-82.
- 534 Halpern BS, Warner RR. 2002. Marine reserves have rapid and lasting effects. *Ecology Letters* **5**:361-366.
- 535 Horta e Costa B, Claudet J, Franco G, Erzini K, Caro A, Gonçalves EJ. 2016. A regulation-based
536 classification system for Marine Protected Areas (MPAs). *Marine Policy* **72**:192-198.
- 537 Jenkins CN, Joppa L. 2009. Expansion of the global terrestrial protected area system. *Biological*
538 *conservation* **142**:2166-2174.
- 539 Langlois TJ, Harvey ES, Fitzpatrick B, Meeuwig JJ, Shedrawi G, Watson DL. 2010. Cost-efficient sampling
540 of fish assemblages: comparison of baited video stations and diver video transects. *Aquatic biology*
541 **9**:155-168.
- 542 Lester SE, Halpern BS, Grorud-Colvert K, Lubchenco J, Ruttenberg BI, Gaines SD, Airamé S, Warner RR.
543 2009. Biological effects within no-take marine reserves: a global synthesis. *Marine Ecology Progress*
544 *Series* **384**:33-46.
- 545 Lewin W-C, Arlinghaus R, Mehner T. 2006. Documented and Potential Biological Impacts of Recreational
546 Fishing: Insights for Management and Conservation. *Reviews in Fisheries Science* **14**:305-367.
- 547 Lim IE, Wilson SK, Holmes TH, Noble MM, Fulton CJ. 2016. Specialization within a shifting habitat
548 mosaic underpins the seasonal abundance of a tropical fish. *Ecosphere* **7**:e01212.
- 549 Mackie M, Black R 1999. Research on Two Serranid Species (Serranidae: Epinephelinae) in Western
550 Australian Waters. University of Western Australia/Fisheries Research & Development Corporation.
- 551 Mallet D, Pelletier D. 2014. Underwater video techniques for observing coastal marine biodiversity: a
552 review of sixty years of publications (1952–2012). *Fisheries Research* **154**:44-62.
- 553 McCook LJ, et al. 2010. Adaptive management of the Great Barrier Reef: A globally significant
554 demonstration of the benefits of networks of marine reserves. *Proceedings of the National Academy*
555 *of Sciences* **107**:18278.
- 556 McLaren BW, Langlois TJ, Harvey ES, Shortland-Jones H, Stevens R. 2015. A small no-take marine
557 sanctuary provides consistent protection for small-bodied by-catch species, but not for large-bodied,
558 high-risk species. *Journal of Experimental Marine Biology and Ecology* **471**:153-163.
- 559 McPhee DP, Leadbitter D, Skilleter GA. 2002. Swallowing the bait: is recreational fishing in Australia
560 ecologically sustainable? *Pacific Conservation Biology* **8**:40-51.
- 561 Miller KI, Russ GR. 2014. Studies of no-take marine reserves: Methods for differentiating reserve and
562 habitat effects. *Ocean & Coastal Management* **96**:51-60.
- 563 Mitchell J, McLean D, Collin S, Taylor S, Jackson G, Fisher R, Langlois T. 2018. Quantifying shark
564 depredation in a recreational fishery in the Ningaloo Marine Park and Exmouth Gulf, Western
565 Australia. *Marine Ecology Progress Series* **587**:141-157.
- 566 Mora C, Sale PF. 2011. Ongoing global biodiversity loss and the need to move beyond protected areas: a
567 review of the technical and practical shortcomings of protected areas on land and sea. *Marine*
568 *ecology progress series* **434**:251-266.
- 569 Mosquera I, Côté IM, Jennings S, Reynolds JD. 2000. Conservation benefits of marine reserves for fish
570 populations. Pages 321-332. *Animal Conservation forum*. Cambridge University Press.
- 571 Murphy HM, Jenkins GP. 2010. Observational methods used in marine spatial monitoring of fishes and
572 associated habitats: a review. *Marine and Freshwater Research* **61**:236-252.
- 573 Osenberg CW, Shima JS, Miller SL, Stier AC. 2011. Ecology: assessing effects of marine protected areas:
574 confounding in space and possible solutions. *Marine protected areas: a multidisciplinary*
575 *approach*:143-167.

576 Pillans RD, Bearham D, Boomer A, Downie R, Patterson TA, Thomson DP, Babcock RC. 2014. Multi year
577 observations reveal variability in residence of a tropical demersal fish, *Lethrinus nebulosus*:
578 implications for spatial management. *PLoS One* **9**:e105507.

579 R Core Team. 2017. R: A language and environment for statistical computing. R Foundation for Statistical
580 Computing, Vienna, Austria.

581 Radford CA, Jeffs AG, Tindle CT, Cole RG, Montgomery JC. 2005. Bubbled waters: The noise generated
582 by underwater breathing apparatus. *Marine and freshwater behaviour and physiology* **38**:259-267.

583 Rees MJ, Knott NA, Davis AR. 2018a. Habitat and seascape patterns drive spatial variability in temperate
584 fish assemblages: implications for marine protected areas. *Marine Ecology Progress Series* **607**:171-
585 186.

586 Rees MJ, Knott NA, Neilson J, Linklater M, Osterloh I, Jordan A, Davis AR. 2018b. Accounting for habitat
587 structural complexity improves the assessment of performance in no-take marine reserves. *Biological
588 Conservation* **224**:100-110.

589 Russ GR. 2002. Yet another review of marine reserves as reef fishery management tools. *Coral reef fishes:
590 dynamics and diversity in a complex ecosystem* **24**:421.

591 Russ GR, Miller KI, Rizzari JR, Alcala AC. 2015. Long-term no-take marine reserve and benthic habitat
592 effects on coral reef fishes. *Marine Ecology Progress Series* **529**:233-248.

593 Ryan K, Hall N, Lai E, Smallwood C, Taylor S, Wise B 2015. State-wide survey of boat-based recreational
594 fishing in Western Australia 2013/14. Fisheries Research Division.

595 Ryan K, Hall N, Lai E, Smallwood C, Taylor S, Wise B. 2017. State-wide survey of boat-based recreational
596 fishing in Western Australia 2015/16.

597 Ryan K, Wise B, Hall N, Pollock K, Sulin E, Gaughan DJ 2013. An integrated system to survey boat-based
598 recreational fishing in Western Australia 2011/12. Fisheries Research Division, Western Australian
599 Fisheries and Marine Research Laboratories.

600 Sala E, Lubchenco J, Grorud-Colvert K, Novelli C, Roberts C, Sumaila UR. 2018. Assessing real progress
601 towards effective ocean protection. *Marine Policy* **91**:11-13.

602 Shears NT, Grace RV, Usmar NR, Kerr V, Babcock RC. 2006. Long-term trends in lobster populations in a
603 partially protected vs. no-take Marine Park. *Biological conservation* **132**:222-231.

604 Smallwood CB, Beckley LE. 2012. Spatial distribution and zoning compliance of recreational fishing in
605 Ningaloo Marine Park, north-western Australia. *Fisheries Research* **125**:40-50.

606 Sumner NR, Williamson PC, Malseed BE 2002. A 12-month survey of recreational fishing in the Gascoyne
607 bioregion of Western Australia during 1998-99. Department of Fisheries, Western Australia.

608 Underwood A. 1993. The mechanics of spatially replicated sampling programmes to detect environmental
609 impacts in a variable world. *Australian Journal of ecology* **18**:99-116.

610 Viechtbauer W. 2010. Conducting meta-analyses in R with the metafor package. *J Stat Softw* **36**:1-48.

611 Watson DL, Harvey ES, Anderson MJ, Kendrick GA. 2005. A comparison of temperate reef fish
612 assemblages recorded by three underwater stereo-video techniques. *Marine Biology* **148**:415-425.

613 Watson M, Ormond R. 1994. Effect of an artisanal fishery on the fish and urchin populations of a Kenyan
614 coral reef. *Marine Ecology Progress Series*:115-129.

615 West L, Stark K, Murphy J, Lyle J, Ochwada-Doyle F. 2015. Survey of recreational fishing in New South
616 Wales and the ACT, 2013/14.

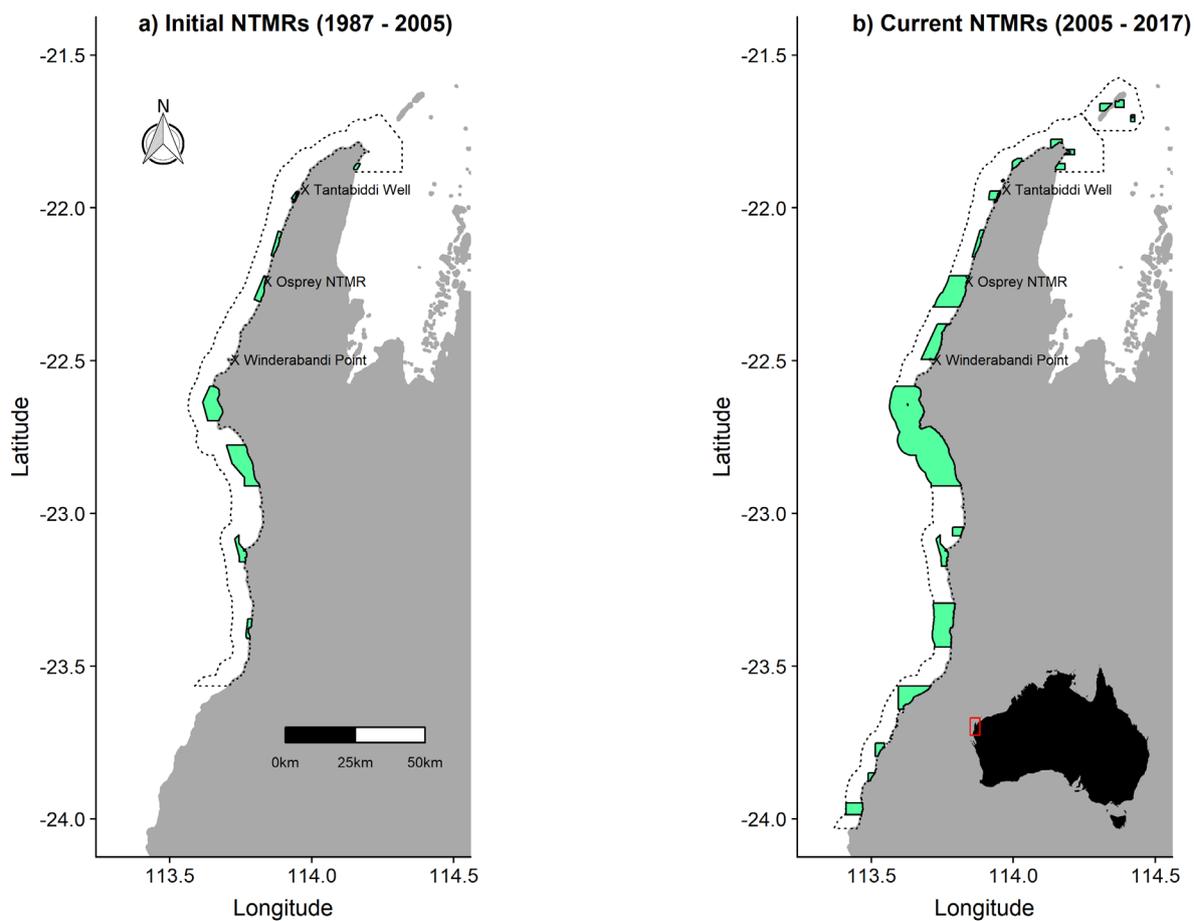
617 Westera MB. 2003. The effect of recreational fishing on targeted fishes and trophic structure, in a coral reef
618 marine park.

619 Wilson S, Fulton C, Depczynski M, Holmes T, Noble M, Radford B, Tinkler P. 2014. Seasonal changes in
620 habitat structure underpin shifts in macroalgae-associated tropical fish communities. *Marine biology*
621 **161**:2597-2607.

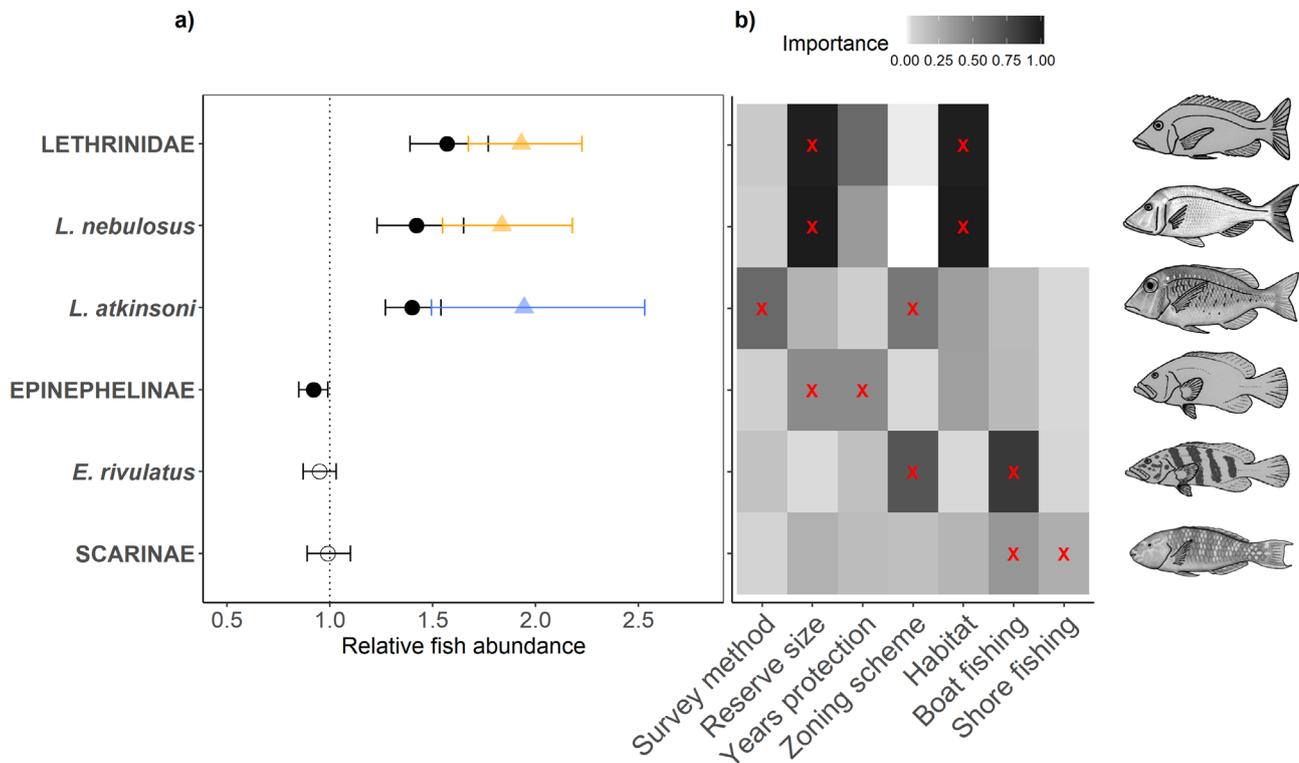
622 Wilson SK, Babcock RC, Fisher R, Holmes TH, Moore JAY, Thomson DP. 2012. Relative and combined
623 effects of habitat and fishing on reef fish communities across a limited fishing gradient at Ningaloo.
624 *Marine Environmental Research* **81**:1-11.

625 Wilson SK, Depczynski M, Fisher R, Holmes TH, Noble MM, Radford BT, Rule M, Shedrawi G, Tinkler P,
626 Fulton CJ. 2018a. Climatic forcing and larval dispersal capabilities shape the replenishment of fishes
627 and their habitat-forming biota on a tropical coral reef. *Ecology and Evolution* **8**:1918-1928.

- 628 Wilson SK, Depczynski M, Holmes TH, Noble MM, Radford BT, Tinkler P, Fulton CJ. 2017. Climatic
629 conditions and nursery habitat quality provide indicators of reef fish recruitment strength. *Limnology
630 and Oceanography* **62**:1868-1880.
- 631 Wilson SK, Graham NAJ, Holmes T, MacNeil MA, Ryan N. 2018b. Visual versus video methods for
632 estimating reef fish biomass. *Ecological Indicators* **85**:146-152.
- 633 Wood SN. 2011. Fast stable restricted maximum likelihood and marginal likelihood estimation of
634 semiparametric generalized linear models. *Journal of the Royal Statistical Society: Series B
635 (Statistical Methodology)* **73**:3-36.
- 636 Zupan M, Fragkopoulou E, Claudet J, Erzini K, Horta e Costa B, Gonçalves EJ. 2018. Marine partially
637 protected areas: drivers of ecological effectiveness. *Frontiers in Ecology and the Environment*
638 **16**:381-387.
- 639 Zuur AF, Ieno EN, Elphick CS. 2010. A protocol for data exploration to avoid common statistical problems.
640 *Methods in ecology and evolution* **1**:3-14.
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649 **Figure 1** The Ningaloo Marine Park (NMP) and Muiron Islands Marine Management Area boundaries
650 (dotted lines) with the location of sanctuary zones (referred to as NTMRs in the present study) shown in
651 green along the Ningaloo coast of Western Australia under the a) initial (1987 – 2005) and b) current (2005 -
652 2017) zoning schemes. Tantabiddi Well and Winderabandi Point are shown as spearfishing is prohibited
653 between these locations. The Osprey NTMR is also indicated.
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655 **Figure 2.** a) Relative fish abundance inside to outside NTMRs (back-transformed weighted mean effect
656 sizes) with 95% confidence intervals), for the six fish groups: Lethrinidae, *Lethrinus nebulosus*, *L. atkinsoni*,
657 Epinephelinae, *Epinephelus rivulatus* and Scarinae. Effect sizes are significant when the confidence
658 intervals do not overlap 1.0 Open dots correspond to non-significant effects (i.e. no effect). Sample sizes are
659 given in Table F1. Triangular points show the predicted effect size when *habitat* was included as a
660 moderator variable in the meta-analysis, for the habitat with the largest mean effect (orange represents the
661 back reef & lagoon coral, and blue represents the reef flat). b) Importance scores (based on summed Akaike
662 weights corrected for finite samples (AICc)) from full-subsets analyses exploring the influence of seven
663 variables on the overall effect size for each fish taxa: 1 is highly important while 0 is not important. Red X
664 symbols mark the variables that were included in the most parsimonious models for each fish taxa (also see
665 Table 2 and Fig. 3).
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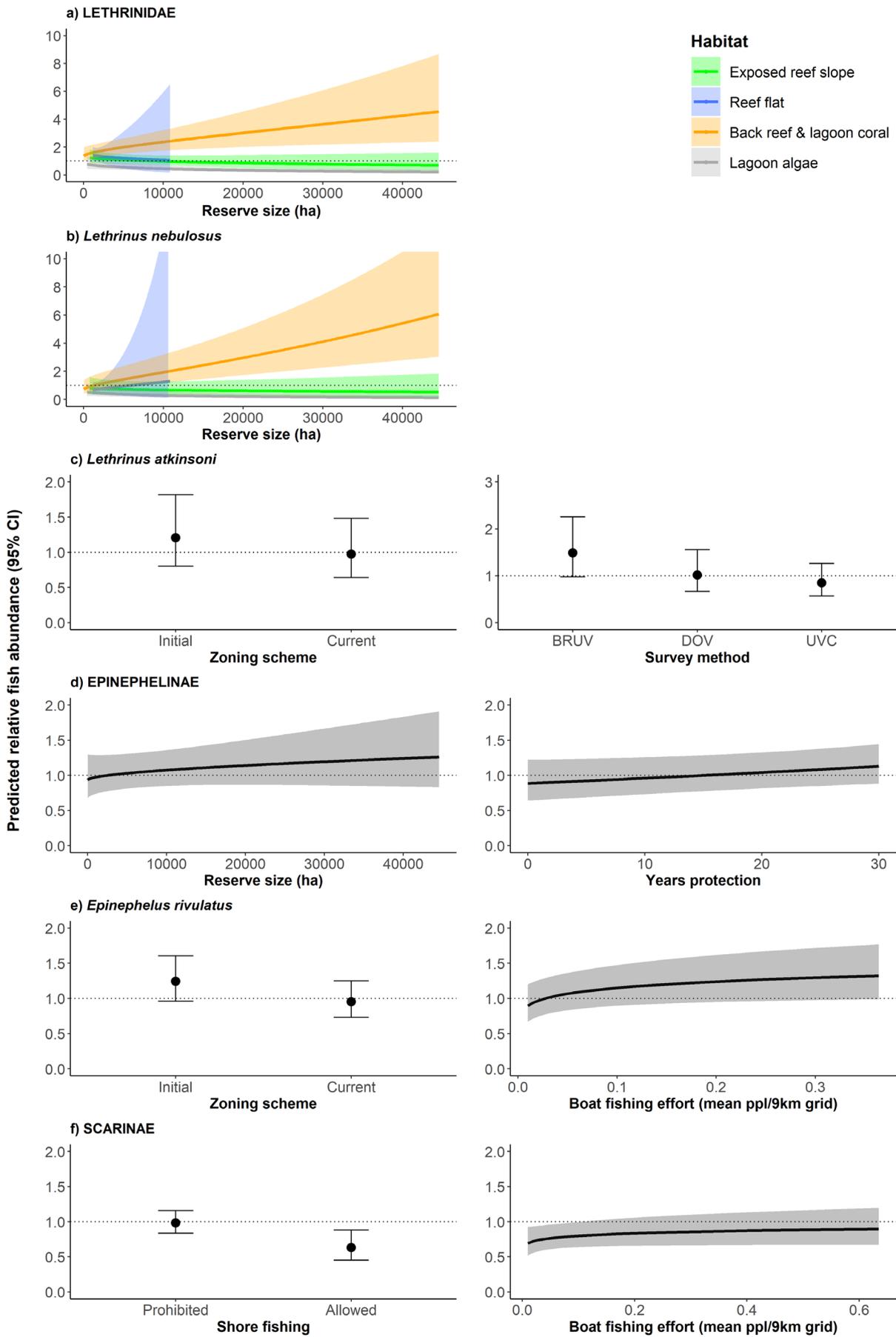
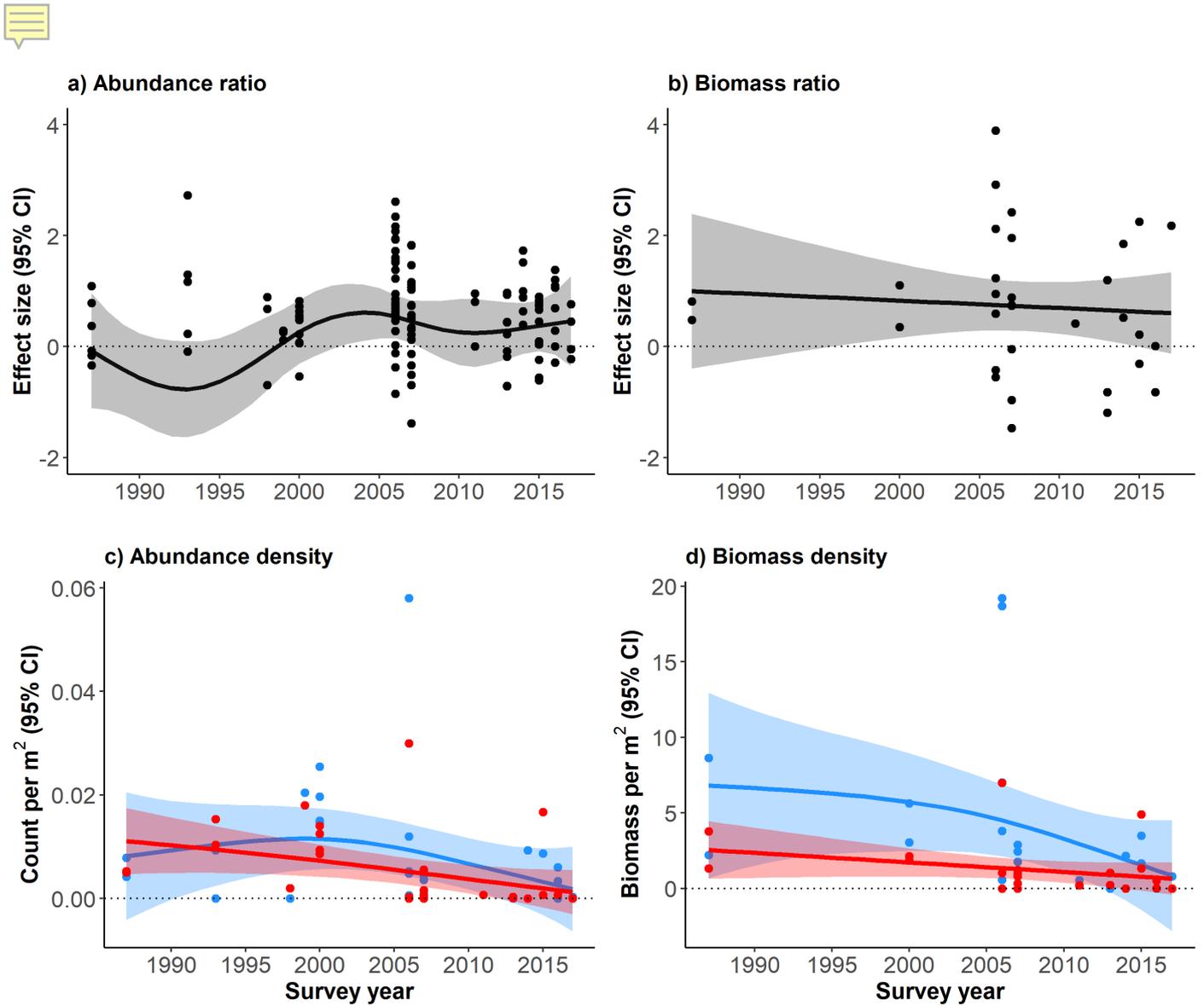


Figure 3. Predicted relative fish abundance inside to outside NTMRs (back-transformed predicted weighted effect sizes) with 95% confidence intervals) for the six fish groups – a) Lethrinidae; b) *Lethrinus nebulosus*; c) *Lethrinus atkinsoni*; d) Epinephelinae; e) *Epinephelus rivulatus*; f) Scarinae for abundance– as a function

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of variables present in the most parsimonious models (Table 2) from full-subsets GAMM analysis. Ribbons represent 95% confidence intervals



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Figure 4. Effect sizes for a) abundance and b) biomass from comparison pairs for the Osprey NTMR through time and estimated density of c) abundance and d) biomass inside and outside the NTMR through time. Ribbons indicate 95% confidence intervals on generalised additive models.

Table 1. Description and summary of the seven variables used in analysis

Variable (transformation used in analyses)	Description	Summary/ description of variable levels	Source
Years protection	Years between zoning and survey data collection	A survey is classified to a single reserve based on its location. In cases where rezoning means that a reserves size was increased, a survey falling inside the old area is classified as the pre-zoning reserve, while if the survey falls in the extended area it is classified as the post-zoning reserve. Years since protection is calculated on the same principal relating to the initial zoning or the rezoning dependent on survey location. 0 - 30 years from time of survey to reserve implementation.	(CALM 2005) (CALM 1989)
Zoning scheme	Factor describing the two major spatial	<i>Initial</i>	(CALM 2005) (CALM 1989)

	zonings implemented in the Ningaloo Marine Park	<i>Current</i>	2005-present, 18 no-take (excepting shore fishing) and 3 no-take zones in the Muiron Islands Marine Management Area, see Fig. 1b	(Appendix A)
Survey method	Factor describing major survey methods used to collect the fish count and size data	<i>UVC</i>	Underwater visual census, collected along transect lines of set length and width (25 x 5 m, 50 x 5 m or 100 x 10 m, 250 x 10 m). Most on SCUBA, some via snorkel. Fish counted and length estimated <i>in situ</i> .	For more information on methods see Appendix B
		<i>BRUV</i>	Baited remote underwater stereo-video deployments, (30-60 minutes) point location, fish counted and length estimated <i>post hoc</i> from video	
		<i>DOV</i>	Diver operated stereo-video, collected along a transect line of set length and width (5 m in width and varying between 25 and 50 m length), fish counted and length estimated <i>post hoc</i> from video	
Habitat	Factor describing four major habitat types which have differences both in the dominant benthic community and wave exposure	<i>Exposed reef slope</i>	The ocean side of the fringing reef, where the reef slopes to deeper water and the majority of wave energy is received	Classified by authors, (see Collins et al. 2003)
		<i>Reef flat</i>	Shallow (~2-3m deep), shoreward from the reef crest for tens to hundreds of meters, typically dominated by the plate coral <i>Acropora spicifera</i> on limestone bedrock	
		<i>Back reef & lagoon coral</i>	From where the reef flat breaks into more patchy reef and sand environments, sheltered from wave energy and including some large coral bommies	
		<i>Lagoon algae</i>	Sheltered shallow water lagoon, dominated by fleshy canopy forming seaweed of the genera <i>Sargassum</i> and <i>Sargassopsis</i> .	
Reserve size (square-root)	Area (ha) of each no-take reserve at time of survey	50 – 44752 hectares Mean: 6031 ha; Median: 1756 ha		(CALM 2005) (CALM 1989)
Boat fishing * (log-transformation)	A mean estimate of the number of vessels recreationally fishing at the outside reserve survey sites	Mean density of vessels observed fishing during aerial surveys in peak season in 2007. Each survey sample latitude and longitude was assigned the value of the underlying spatial data in Fig. 4 of Smallwood and Beckley (2012). For surveys inside the NTMRs it was assumed that fishing activity was 0. 0 - 0.625 vessels fishing per 9 km ² Mean: 0.12; Median: 0.11 * Not available for the Muiron Islands Other estimates of fishing activity exist (Sumner et al. 2002) but the metric chosen was deemed the most detailed		Smallwood and Beckley (2012)
Shore fishing	Factor describing whether or not shore fishing permitted in a reserve	<i>Allowed</i>	Shore fishing is allowed along the entire, or part of the coastline side of the reserve (26% of data)	(CALM 2005)
		<i>Prohibited</i>	No shore fishing is permitted anywhere in the reserve (74% of data)	

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Table 2. Top Generalised Additive Mixed Models (GAMMs) for predicting the response ratio inside to outside NTMRs, \bar{E} , for abundance from full subset analyses for the abundance of the six fish groups. Difference between the lowest reported corrected Akaike Information Criterion ($\Delta AICc$), $AICc$ weights ($\omega AICc$), variance explained (R^2) and estimated degrees of freedom (EDF) are reported for model comparison. Model selection was based on the most parsimonious model (fewest variables and lowest EDF) within two units of the lowest $AICc$. This model is shown in bold text.

Fish group	Model	$\Delta AICc$	$\omega AICc$	R^2	EDF
LETHRINIDAE	Habitat + Years protection by Habitat + Size by Habitat	0.00	0.31	0.14	14.6
	Years protection + Habitat + Size by Habitat	0.17	0.28	0.13	10.7
	Habitat + Size by Habitat	0.19	0.28	0.12	9.0
<i>L. nebulosus</i>	Habitat + Size by Habitat	0.00	0.57	0.17	9.0
<i>L. atkinsoni</i>	Method + Zoning scheme	0.00	0.19	0.08	6.3
	Habitat + Method + Zoning scheme	0.09	0.18	0.09	9.0
	boat.log + Method + Zoning scheme	0.71	0.14	0.08	7.4
	Habitat + Method + Size	1.41	0.10	0.08	8.0
	Habitat + Size + Years protection by Habitat	0.00	0.18	0.11	14.4
	Years protection + Boat fishing + Size	0.92	0.11	0.08	8.3
EPINEPHELINAE	Years protection + Size	1.51	0.08	0.08	7.3

<i>E. rivulatus</i>	Boat fishing + Zoning scheme	0.00	0.60	0.17	8.9
SCARINAE	Boat fishing + Shore fishing	0.00	0.16	0.03	4.0
	Boat fishing + Zoning scheme	0.80	0.11	0.03	4.2
	Years protection + Habitat + Size by Habitat	1.25	0.08	0.05	10.5
	Habitat + Size by Habitat	1.84	0.06	0.04	9.0

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713 *Appendix A – Additional information on individual reserves and rezoning*

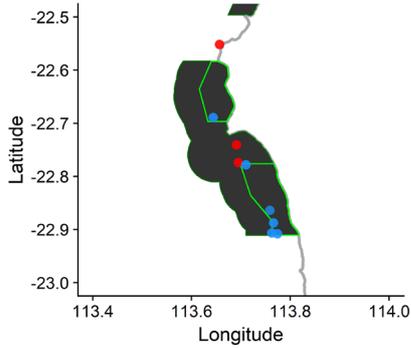
714 Maps of individual reserves can be found in CALM (1989), pgs. 55, for the 1987-2005 zoning and (CALM
 715 2005), pgs. 89-96, for the 2005-current zoning. Table A1 details individual reserves, their year of
 716 establishment, size, and the regulations on shore-based fishing on the coastal edges of the reserves. As well,
 717 the number of inside outside comparisons and the total number of surveys are given for each reserve.
 718

719 **Table A1.** Features of historical and current reserves in the Ningaloo Marine Park

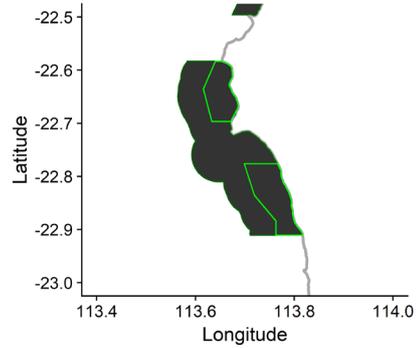
	Reserve	Management area	Year of Establishment	Size (ha)	Shore fishing	Total number comparison pairs	Total number of samples	
2005 – PRESENT MANAGEMENT (CURRENT)	3 Mile	Ningaloo Marine Park	2005	395	N	0	0	
	Bateman	Ningaloo Marine Park	2005	1111	N	1	12	
	Bundegi rezoned	Ningaloo Marine Park	1987/2005	696	N	3	24	
	Cape Farquhar	Ningaloo Marine Park	2005	5326	N	3	31	
	Cloates/ Dugong rezoned	Ningaloo Marine Park	1987/2005	44752	P	17	268	
	Gnaraloo Bay	Ningaloo Marine Park	2005	1021	N	2	39	
	Jurabi	Ningaloo Marine Park	2005	754	Y	9	78	
	Lakeside	Ningaloo Marine Park	2005	8	N	0	0	
	Lighthouse Bay	Ningaloo Marine Park	2005	763	P	9	136	
	Mandu rezoned	Ningaloo Marine Park	1987/2004	1349	N	12	110	
	Mangrove rezoned	Ningaloo Marine Park	1987/2005	1135	N	27	429	
	Maud rezoned	Ningaloo Marine Park	1987/2005	2151	P	10	133	
	Murat	Ningaloo Marine Park	2005	490	Y	0	0	
	North Muiron	Muiron Islands Marine Management Size	2005	828	N	4	66	
	Osprey rezoned	Ningaloo Marine Park	1987/2005	9513	P	7	54	
	Pelican rezoned	Ningaloo Marine Park	1987/2006	10864	P	28	454	
	South Muiron	Muiron Islands Marine Management Size	2005	784	N	4	54	
	Sunday Island	Muiron Islands Marine Management Size	2005	317	N	0	0	
	1987 – 2005 MANAGEMENT (INITIAL)	Tantabiddi	Ningaloo Marine Park	2005	50	N	8	98
		Turtles	Ningaloo Marine Park	2005	2461	N	0	0
Winderabandi		Ningaloo Marine Park	2005	5526	Y	7	52	
Bundegi		Ningaloo Marine Park	1987	297	N	20	333	
Cloates		Ningaloo Marine Park	1987	6257	N	16	186	
Dugong		Ningaloo Marine Park	1987	8852	N	0	0	
Mandu		Ningaloo Marine Park	1987	1163	N	50	1001	
Mangrove		Ningaloo Marine Park	1987	403	N	2	14	
Maud		Ningaloo Marine Park	1987	1806	N	36	652	
Osprey		Ningaloo Marine Park	1987	1756	N	44	701	
Pelican		Ningaloo Marine Park	1987	908	N	7	66	
3 Mile	Ningaloo Marine Park	2005	395	N	0	0		

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 721 The rezoning of the eight NTMRs in 2005, with the addition of new reserves and the expansion of the
 722 existing reserves required that data be clearly classified to account for this. Figure A1 shows an example of
 723 how samples were referenced based on their spatial location to the initial or current reserves and the time
 724 period of sampling.

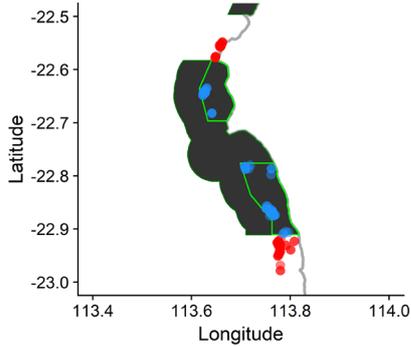
a) Surveys referenced to 'Initial zoning', date pre 2005



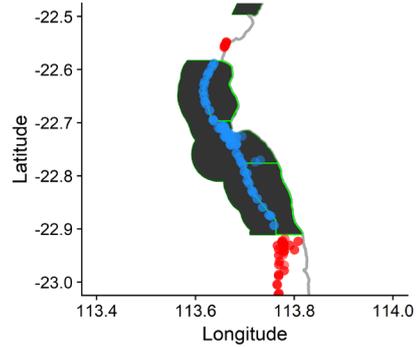
b) Surveys referenced to 'Current zoning', date pre 2005



c) Surveys referenced to 'Initial zoning', date post 2005



d) Surveys referenced to 'Current zoning', date post 2005



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Figure A1. Example classification of 'initial' and 'current' zoning. The boundaries of the initial 1987 – 2005 reserves, Cloates and Dugong, are indicated by a green outline, while the current reserve (2005 – 2017) is shown in black which combined the two initial reserves into one larger reserve, 'Cloates/Dugong rezoned' (also see Table 1). Pre 2005, surveys were classified as inside/outside based on their spatial relation to initial reserves (green outline). Post 2005, surveys were classified as inside/outside based on their spatial relation to the current reserves (black). If an inside reserve is within the initial boundaries, but surveyed post 2005, it was referenced as initial as shown in c). If a sample was collected at a site within the new area of the rezoned reserve post the rezoning, it was referenced as current as seen in d).

Appendix B– Data summary

Suitable data were identified through searches on Google Scholar and research databases compiled by the Department of Biodiversity, Conservation and Attractions. In addition, relevant researchers from universities, research institutions, industry and citizen science programs operating in Western Australia were contacted to source unpublished data. Nine major custodians contributed data to this study. Table B1 summarises the survey methods used by each custodian, the number of samples collected, the temporal span of data, the reserves surveyed and directs to further reading for more information. Figure B2 graphically illustrates the important data constraints.

Table B1. Summary of data contributions

Data custodian	Total number of surveys	Reserves surveyed	Maximum temporal span of surveys	Main survey methods	More information available:
Australian Institute of Marine Science – Woodside Energy (AIMS-Woodside)	183	Bundegi, Bundegi rezoned, Cloates, Cloates/Dugong rezoned, Mandu, Mangrove rezoned, Maud, North Muiron, Osprey, Pelican rezoned, Tantabiddi	1993 -2014	Underwater Visual Census : 50 x 5 m	(Depczynski et al. 2015)
Ben Fitzpatrick	345	Mandu, Osprey	2006 - 2007	Baited Remote Underwater stereo-Video	(Fitzpatrick et al. 2015)
Commonwealth Industrial and Scientific Research Organisation (CSIRO)	18	Bundegi, Bundegi rezoned, Cape Farquhar, Cloates, Cloates/Dugong rezoned, Gnaraloo Bay, Jurabi, Lighthouse Bay, Mandu, Mandu rezoned, Mangrove, Mangrove rezoned, Maud, North Muiron, Osprey, Osprey rezoned, Pelican, Pelican rezoned, South Muiron	2006 - 2017	Underwater Visual Census, 4 main sizes: 1) Either a singular, or three, 25 × 5 m transects per site. 2) 30 × 5 m transects 3) 50 × 5 m transects 4) 100 × 10 m transects at a site	(Babcock et al. 2008)
Western Australian Department of Biodiversity, Conservation and Attractions (DBCA)	1237	Bundegi , Cape Farquhar, Cloates, Cloates/Dugong rezoned, Jurabi, Lighthouse Bay, Mandu, Mangrove rezoned, Maud, Maud rezoned, North Muiron, Osprey, Pelican, Pelican rezoned, Tantabiddi, Winderabandi	2010 - 2016	Diver Operated stereo-Video: six replicate 50 x 5 m belt transects per site; or nine replicate 30 x 5 m belt transects per site	(Wilson et al. 2012; Holmes et al. 2013; Wilson et al. 2018b)
Mark Westera	257	Mandu, Maud, Osprey	1999 - 2000	Baited Remote Underwater stereo-Video, 30 minute deployments at 12 replicate locations in each zone, Underwater visual census using snorkel, 250 x 10 m transects	(Westera 2003a, b)
Reef Life Survey (RLS)	291	Bateman, Bundegi, Cloates, Maud, Maud rezoned, Pelican rezoned	2010 - 2017	Underwater Visual Census, 50 x 5 m transects	(see http://reeflifesurvey.com/files/2008/09/rils-reef-monitoring-procedures.pdf).
Tony Ayling	60	Osprey	1987		(Ayling & Ayling 1987)
The University of Western Australia	325	Cloates, Cloates/Dugong rezoned, Mandu rezoned, Mangrove rezoned, Osprey, Osprey rezoned, Pelican rezoned, Winderabandi	2014 - 2015	Baited Remote Underwater stereo-Video, generally 60 minute deployments	(McLean et al. 2016)
Joint data custodians WA Department of Biodiversity, Conservation and Attractions, Australian Institute of Marine Science, Australian National University	587	Cloates/Dugong rezoned, Mandu, Mangrove rezoned, Maud, Maud rezoned, Pelican, Pelican rezoned,	2013 - 2015	Underwater Visual Census, 4 main sizes: 1) Either a singular, or three, 25 × 5 m transects per site. 2) 30 × 5 m transects 3) 50 × 5 m transects 4) 100 × 10 m transects at a site	(Fulton et al. 2014)

In most surveys biomass had been calculated by the respective data custodians from the estimated or measured fish length. In these cases the provided values were used (with the reasoning that different survey methods may warrant slightly different biomass calculations). In cases where length data was available but biomass had not been calculated the fish counts and length estimates were converted to biomass (kg) using constants and formulas from Fishbase (www.fishbase.org).

$$Biomass = e^{\ln(a)+b \times \ln(L)}$$

where L is the estimated total length of the fish and a and b are constants for the species or family in question. In cases where data provided fork length measurements, these were converted to total length using the formula from FishBase:

$$TL = c + d \times FL$$

where TL is estimated total fish length, FL is the measured fork length from BRUV or DOV video and c and d are parameters specific to the fish species in question.

More data was available for some NTMRs and years, with increased sampling after 2005 (Appendix B).

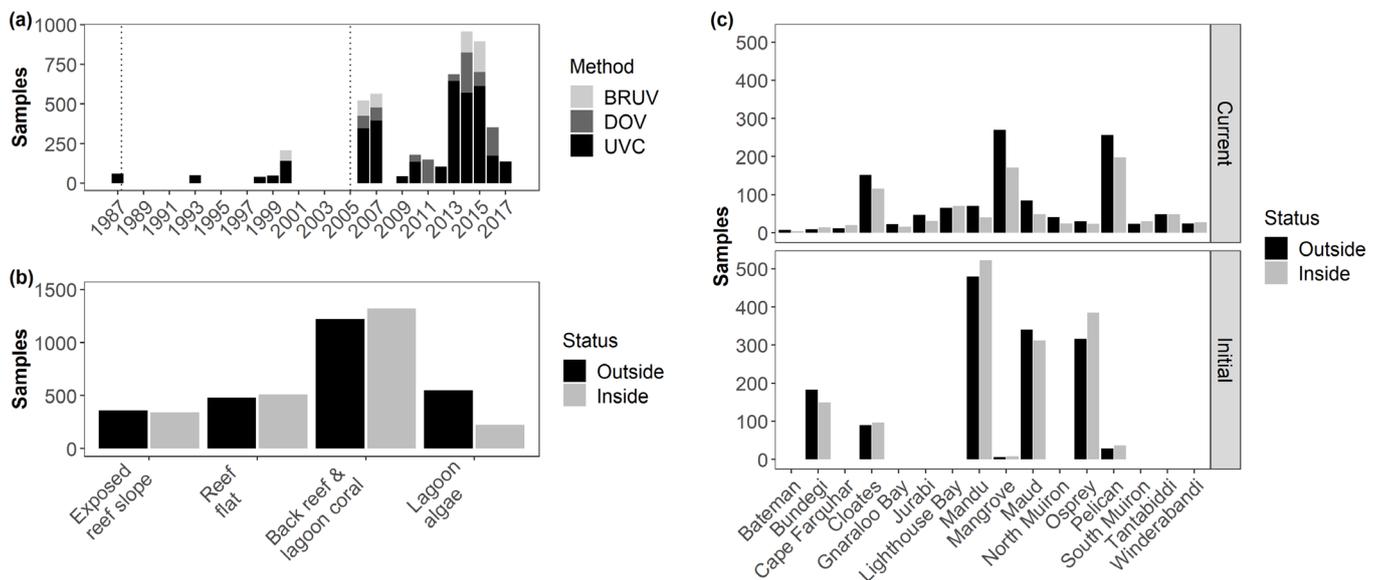


Figure B2. Temporal and spatial distribution of samples: a) number of samples by year, with shade indicating the type of survey method (BRUV, Baited Remote Underwater stereo-Video, DOV, Diver Operated stereo-Video and UVC, Underwater Visual Census); b) distribution of samples inside and outside NTMRs for each of the four major habitats; c) distribution of samples inside and outside for each NTMR under both the initial and current zoning.

References

- 776 Ayling T, Ayling AL 1987. Ningaloo Marine Park: preliminary fish density assessment and habitat survey:
777 with information on coral damage due to *Drupella cornus* grazing: a report prepared for the
778 Department of Conservation and Land Management, Western Australia. Department of Conservation
779 and Land Management.
- 780 Babcock R, Haywood M, Vanderklift M, Clapin G, Kleczkowski M, Dennis D, Skewes T, Milton D,
781 Murphy N, Pillans R. 2008. Ecosystem Impacts of Human Usage and the Effectiveness of Zoning for
782 Biodiversity conservation: Broad-scale Fish Census. Final Analysis and Recommendations 2007.,
783 Hobart.
- 784 Babcock RC, Shears NT, Alcala AC, Barrett NS, Edgar GJ, Lafferty KD, McClanahan TR, Russ GR. 2010.
785 Decadal trends in marine reserves reveal differential rates of change in direct and indirect effects.
786 *Proceedings of the National Academy of Sciences* **107**:18256.
- 787 Depczynski MM, Tinkler PP, Cheal AA, Speed CC. 2015. Ningaloo fish communities. Page 121 in Miller
788 KK, Depczynski MM, Cappo MM, Wakeford MM, Speed CC, Stowar MM, Colquhoun JJ, Tinkler
789 PP, Cheal AA, and Fisher RR, editors. Ningaloo and Outer Shark Bay Environmental Baseline
790 Survey 2014. Report prepared by the Australian Institute of Marine Science for Woodside Energy
791 Ltd. Australian Institute of Marine Science, Townsville, Australia.
- 792 Fitzpatrick B, Harvey E, Langlois T, Babcock R, Twiggs E. 2015. Effects of fishing on fish assemblages at
793 the reefscape scale. *Marine Ecology Progress Series* **524**:241-253.
- 794 Holmes TH, Wilson SK, Travers MJ, Langlois TJ, Evans RD, Moore GI, Douglas RA, Shedrawi G, Harvey
795 ES, Hickey K. 2013. A comparison of visual-and stereo-video based fish community assessment
796 methods in tropical and temperate marine waters of Western Australia. *Limnology and*
797 *Oceanography, Methods* **11**:337-350.
- 798 Westera MB. 2003a. The effect of recreational fishing on targeted fishes and trophic structure, in a coral reef
799 marine park.
- 800 Westera MB. 2003b. The effect of recreational fishing on targeted fishes and trophic structure, in a coral reef
801 marine park. Edith Cowan University, Perth, Western Australia.
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805 *Appendix C – Geographic locations of individual samples*

806 Kml file for viewing samples in GoogleEarth

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809 **Appendix D: Fish groups**

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811 Analyses were conducted at the family/subfamily level and species level. Investigation of patterns at the
812 family/subfamily level allowed the inclusion of data from targeted but rare species.

813

814 Table D1. Total count of fish groups across all data

Taxa	Total count across synthesised data
LETHRINIDAE	10307
<i>L. nebulosus</i>	4183
<i>L. atkinsoni</i>	4765
EPINEPHELINAE	4012
<i>E. rivulatus</i>	1119
SCARINAE	44931

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818 Genera included in the family/ sub-family analysis that were sampled in the synthesised data are
819 summarised below.

820

821 **Lethrinidae**

- | | | | |
|-----|-------------------------------------|-----|-------------------------------------|
| 822 | • <i>Gnathodentex aureolineatus</i> | 833 | • <i>Lethrinus microdon</i> |
| 823 | • <i>Gymnocranius euanus</i> | 834 | • <i>Lethrinus miniatus</i> |
| 824 | • <i>Gymnocranius grandoculis</i> | 835 | • <i>Lethrinus nebulosus</i> |
| 825 | • <i>Gymnocranius griseus</i> | 836 | • <i>Lethrinus obsoletus</i> |
| 826 | • <i>Gymnocranius spp.</i> | 837 | • <i>Lethrinus olivaceus</i> |
| 827 | • <i>Lethrinus amboinensis</i> | 838 | • <i>Lethrinus ravus</i> |
| 828 | • <i>Lethrinus atkinsoni</i> | 839 | • <i>Lethrinus rubrioperculatus</i> |
| 829 | • <i>Lethrinus genivittatus</i> | 840 | • <i>Lethrinus semicinctus</i> |
| 830 | • <i>Lethrinus harak</i> | 841 | • <i>Lethrinus variegatus</i> |
| 831 | • <i>Lethrinus laticaudis</i> | 842 | • <i>Lethrinus spp.</i> |
| 832 | • <i>Lethrinus lentjan</i> | 843 | • <i>Monotaxis grandoculis</i> |

844

845 **Epinephelinae ***

- | | | | |
|-----|------------------------------------|-----|--|
| 846 | • <i>Aethaloperca rogae</i> | 855 | • <i>Cephalopholis sonnerati</i> |
| 847 | • <i>Anyperodon leucogrammicus</i> | 856 | • <i>Cromileptes altivelis</i> |
| 848 | • <i>Cephalopholis spp.</i> | 857 | • <i>Epinephelus spp.</i> |
| 849 | • <i>Cephalopholis argus</i> | 858 | • <i>Epinephelus areolatus</i> |
| 850 | • <i>Cephalopholis boenak</i> | 859 | • <i>Epinephelus bilobatus</i> |
| 851 | • <i>Cephalopholis cyanostigma</i> | 860 | • <i>Epinephelus coeruleopunctatus</i> |
| 852 | • <i>Cephalopholis formosa</i> | 861 | • <i>Epinephelus coioides</i> |
| 853 | • <i>Cephalopholis miniata</i> | 862 | • <i>Epinephelus corallicola</i> |
| 854 | • <i>Cephalopholis sexmaculata</i> | 863 | • <i>Epinephelus fasciatus</i> |

864	• <i>Epinephelus fuscoguttatus</i>	873	• <i>Epinephelus quoyanus</i>
865	• <i>Epinephelus hexagonatus</i>	874	• <i>Epinephelus rivulatus</i>
866	• <i>Epinephelus lanceolatus</i>	875	• <i>Epinephelus tauvina</i>
867	• <i>Epinephelus macrospilos</i>	876	• <i>Epinephelus tukula</i>
868	• <i>Epinephelus maculatus</i>	877	• <i>Plectropomus leopardus</i>
869	• <i>Epinephelus malabaricus</i>	878	• <i>Plectropomus maculatus</i>
870	• <i>Epinephelus melanostigma</i>	879	• <i>Plectropomus</i> spp.
871	• <i>Epinephelus merra</i>	880	• <i>Variola albimarginata</i>
872	• <i>Epinephelus polyphkadion</i>	881	• <i>Variola louti</i>
882	•		
883			

885 * Note that there is current discussion of whether Epinephelinae may be better classified as its own family,
886 Epinephelidae (Ma & Craig 2018), however we have chosen to name it as its subfamily here.

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889 **Scarinae**

- 890 • *Calotomus carolinus*
- 891 • *Calotomus spinidens*
- 892 • *Cetoscarus ocellatus*
- 893 • *Chlorurus bleekeri*
- 894 • *Chlorurus microrhinos*
- 895 • *Chlorurus sordidus*
- 896 • *Hipposcarus longiceps*
- 897 • *Leptoscarus vaigiensis*
- 898 • *Scarus chameleon*
- 899 • *Scarus dimidiatus*
- 900 • *Scarus flavipectoralis*
- 901 • *Scarus frenatus*
- 902 • *Scarus ghobban*
- 903 • *Scarus globiceps*
- 904 • *Scarus niger*
- 905 • *Scarus oviceps*
- 906 • *Scarus prasiognathos*
- 907 • *Scarus psittacus*
- 908 • *Scarus rivulatus*
- 909 • *Scarus rubroviolaceus*
- 910 • *Scarus schlegeli*
- 911 • *Scarus* spp.

912

913 **References**

914 Ma KY, Craig MT. 2018. An inconvenient monophyly: an update on the taxonomy of the groupers
915 (Epinephelidae). *Copeia* **106**:443-456.

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Appendix E: Formulas used for calculating effect sizes

For each inside/ outside comparison pair a log response ratio (e) (Hedges et al. 1999) was calculated as the ratio of the mean abundance/ biomass inside to outside a NTMR for comparison pair i and fish group j as

$$e_{ij} = \ln\left(\frac{\bar{X}_{ij,I}}{\bar{X}_{ij,O}}\right)$$

where $\bar{X}_{ij,I}$ and $\bar{X}_{ij,O}$ are the mean abundance or biomass inside (I) and outside (O) a NTMR. Therefore, a positive e_{ij} implies a greater fish abundance/ biomass inside the NTMRs than outside. A log response ratio was appropriate because it is independent of the actual unit of measurement across the different survey methods.

The variance of e_{ij} was also quantified (i.e. the within-study variance, where a study is a comparison pair), given that sampling error plays an important role in introducing variability in the overall outcome of a meta-analysis:

$$v_{ij} = \frac{\sigma_{ij,I}^2}{(n_{ij,I}\bar{X}_{ij,I}^2)} + \frac{\sigma_{ij,O}^2}{(n_{ij,O}\bar{X}_{ij,O}^2)}$$

Here, v_{ij} is calculated from the standard deviation, σ_{ij} , sample size, n_{ij} , and mean \bar{X}_{ij} for inside (I) and outside (O) for comparison pair i and fish group j .

Effect sizes were weighted by the inverse of the sum of the within- and among-study variances, w_{ij} . The within-study variance (where a study is a comparison pair) was the sum of the variances associated with each mean in the log-ratio. The among-study variance was calculated using the *metafor* package (Viechtbauer 2010) in the statistical program R (R Core Team 2017).

$$w_{ij} = \frac{1}{v_{ij}}$$

This weighting minimized the influence of studies with low statistical power, and increased the influence of studies with high statistical power, meaning that each inside/ outside replicate did not contribute equally to the final pooled outcome. Weighted meta-analyses of this sort are considered to increase the precision and power of meta-analyses (Osenberg et al. 1999) and was an appropriate approach in the present study as there was a large distribution of sample sizes and variance associated with the inside/ outside comparison pairs.

The weighted cumulative effect size for fish group j , \bar{E}_j , was obtained as

$$\bar{E}_j = \frac{\sum_{i=1}^{n_{ij}} w_{ij} e_{ij}}{\sum_{i=1}^{n_{ij}} w_{ij}} \text{ with associated variance, } \bar{v}_j = \frac{1}{\sum_{i=1}^{n_{ij}} w_{ij}}.$$

where w_{ij} and e_{ij} are defined above. The overall heterogeneity (Q_t) for fish group j was calculated as

$$Q_{t,j} = \sum_{i=1}^{n_{ij}} w_{ij} (e_{ij} - \bar{E}_j)^2$$

and its significance was tested against the χ^2 distribution with $n_{ij} - 1$ degrees of freedom.

We used a random-effects model to calculate \bar{E}_j and $Q_{t,j}$ using the *metafor* package in R through the call `rma(eij, vij, method = "REML", data =)` following the suggestions in Viechtbauer (2005) that the REML, restricted maximum likelihood estimator for variance, strikes a good balance between unbiasedness and efficiency.

We ran mixed-effects categorical analyses and meta-regression to examine how the seven additional variables (Table 1, main text) mediated the overall effect size \bar{E}_j . For a given level (L) of the factor, a weighted cumulative effect size was calculated as

$$\bar{E}_{L,j} = \frac{\sum_{i=1}^{n_{Lj}} w_{ij} e_{ij}}{\sum_{i=1}^{n_{Lj}} w_{ij}}$$

Where n_{Lj} is the number of comparison pairs in level, L , of the factor, and e_{ij} and w_{ij} are defined above.

The heterogeneity of the model explained by the factor (Q_m) was calculated as

$$Q_{m,j} = \sum_{i=1}^{n_{Lj}} w_{ij} (\bar{E}_{L,j} - \bar{E}_j)^2$$

The significance of Q_m was tested against the χ^2 distribution with $n_{Lj} - 1$ degrees of freedom

References

- Hedges LV, Gurevitch J, Curtis PS. 1999. The meta-analysis of response ratios in experimental ecology. *Ecology* **80**:1150-1156.
- Osenberg CW, Sarnelle O, Cooper SD, Holt RD. 1999. Resolving ecological questions through meta-analysis: goals, metrics, and models. *Ecology* **80**:1105-1117.

Appendix F - Sensitivity analysis for case of one-armed zero events

Cheng et al. (2016) define two scenarios relating to zero events: zero-event and double-zero-event. In the present study there were both double-zero-events (a zero mean abundance/biomass for both inside and outside) and zero-events (zero mean abundance/biomass for either the inside or the outside), summarised in Table 1. In both cases, $\ln R$ cannot be computed due to division by zero and/or logarithm of zero. The former case are often removed from meta-analyses, while the latter is often handled via the addition of a small constant (Bradburn et al. 2007; Spittal et al. 2015). The total number of comparisons (n) used in the final calculated of the weighted average effect size is given.

In the present study the species examined can be encountered infrequently on a survey. As well, one of the key species, *Lethrinus nebulosus*, is a schooling species which, when it occurs, can be present in larger numbers. Consequently there are many zero counts in the data which needed to be explicitly considered.

Table F1. Summary of total number of comparisons and zero events

	Fish Group	Total number of possible comparisons	Double-zero events	Zero-events	Final sample size (n) in calculation of mean weighted effect size
ABUNDANCE	LETHRINIDAE	305	4	151	301
	<i>L. nebulosus</i>	305	48	162	257
	<i>L. atkinsoni</i>	305	25	101	280
	EPINEPHELINAE	305	28	110	277
	<i>E. rivulatus</i>	305	138	210	167
	SCARINAE	305	44	46	261
BIOMASS	LETHRINIDAE	268	4	49	264
	<i>L. nebulosus</i>	268	45	143	223
	<i>L. atkinsoni</i>	268	21	86	247
	EPINEPHELINAE	268	15	87	253
	<i>E. rivulatus</i>	268	112	180	156
	SCARINAE	268	45	48	223

Double-zero-events were removed from analysis. For the remaining data a constant of 0.5 fish (half the smallest unit in the abundance analysis) was added to both the inside and outside mean counts, while for biomass a constant of 100 grams was added to both the inside and outside mean biomass.

Given there is no clear protocol for the size of the constant used to deal with zeros in a $\ln R$ meta-analysis, we conducted a sensitivity analysis on the value of the constant. We ran the sensitivity analysis for the abundance data for the six taxa examined. We tested constant values, c , ranging from $c = 0.01$ to $c = 1$ in 0.01 increments.

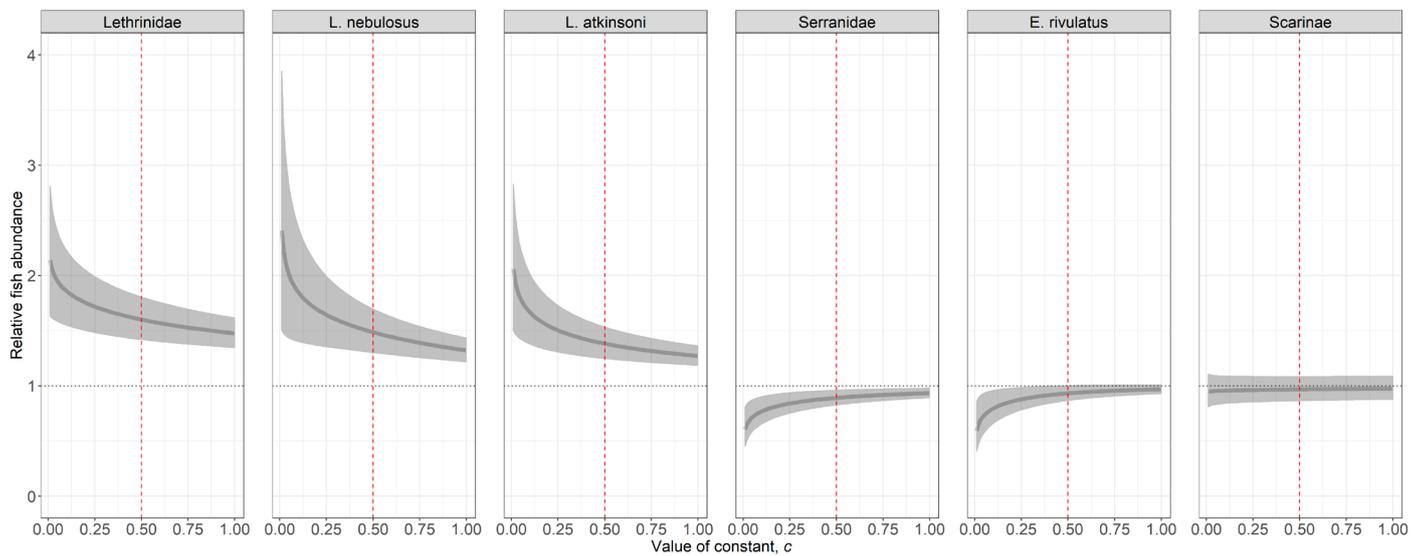


Figure F1. Sensitivity analysis for the size of the constant added to inside/outside mean abundance showing the resulting transformed mean weighted effect size, exponential of \bar{E} , and 95% confidence interval (CI) for values of constant c ranging from 0.01 to 1 in 0.01 increments. a) Lethrinidae, b) *L. nebulosus*, c) *L. atkinsoni*, d) Epinephelinae, e) *E. rivulatus*, f) Scarinae. The mean effect sizes are considered significant when the confidence intervals do not include one.

It is clear that the size of the constant used influenced the mean overall effect size. For example, the magnitude of \bar{E} for *L. nebulosus* total abundance varied from $\bar{E} = 0.73$ (0.54) when $c = 0.01$ to $\bar{E} = 0.28$ (0.08) when $c = 1$. However, while the constant size impacted the magnitude of the effect size, it did not influence the significance, except for *E. rivulatus*, which had the highest count of zero-events, and transitioned from marginally negative to not significantly different from one. From this analysis we decided a constant of 0.5 would be an adequate, and conservative addition for the calculation of $\ln R$ in this analysis. Given the high levels of inherent variability expected in fish count data (Samoilys et al. 1995; Cappo & Brown 1996) and additional variation from uncontrolled variables, even with addition of a constant, overall differences in abundance would have to be consistent in order to observe statistical significance. Nonetheless, we urge caution in the interpretation of the magnitude of the overall effect.

References

- Abesamis RA, Green AL, Russ GR, Jadloc CRL. 2014. The intrinsic vulnerability to fishing of coral reef fishes and their differential recovery in fishery closures. *Reviews in Fish Biology and Fisheries* **24**:1033-1063.
- Anderson DR, Burnham KP. 2002. Avoiding pitfalls when using information-theoretic methods. *The Journal of Wildlife Management*:912-918.

- 026 Ayling T, Ayling AL 1987. Ningaloo Marine Park: preliminary fish density assessment and habitat survey:
027 with information on coral damage due to *Drupella cornus* grazing: a report prepared for the
028 Department of Conservation and Land Management, Western Australia. Department of Conservation
029 and Land Management.
- 030 Babcock R, Haywood M, Vanderklift M, Clapin G, Kleczkowski M, Dennis D, Skewes T, Milton D,
031 Murphy N, Pillans R. 2008. Ecosystem Impacts of Human Usage and the Effectiveness of Zoning for
032 Biodiversity conservation: Broad-scale Fish Census. Final Analysis and Recommendations 2007.,
033 Hobart.
- 034 Babcock R, Phillips J, Lourey M, Clapin G. 2007. Increased density, biomass and egg production in an
035 unfished population of Western Rock Lobster (*Panulirus cygnus*) at Rottne Island, Western
036 Australia. *Marine and Freshwater Research* **58**:286-292.
- 037 Babcock R, Pillans R, Rochester W. 2017. Environmental and individual effects on the behaviour and
038 spawning movements of *Lethrinus nebulosus* on a coral reef. *Marine and Freshwater Research*
039 **68**:1422-1437.
- 040 Ballantine B. 2014. Fifty years on: lessons from marine reserves in New Zealand and principles for a
041 worldwide network. *Biological Conservation* **176**:297-307.
- 042 Barlow J, França F, Gardner TA, Hicks CC, Lennox GD, Berenguer E, Castello L, Economo EP, Ferreira J,
043 Guenard B. 2018. The future of hyperdiverse tropical ecosystems. *Nature* **559**:517.
- 044 Barrett NS, Edgar GJ, Buxton CD, Haddon M. 2007. Changes in fish assemblages following 10 years of
045 protection in Tasmanian marine protected areas. *Journal of Experimental Marine Biology and*
046 *Ecology* **345**:141-157.
- 047 Beckley LE, Lombard AT. 2012. A systematic evaluation of the incremental protection of broad-scale
048 habitats at Ningaloo Reef, Western Australia. *Marine and Freshwater Research* **63**:17-22.
- 049 Bergseth BJ, Williamson DH, Russ GR, Sutton SG, Cinner JE. 2017. A social–ecological approach to
050 assessing and managing poaching by recreational fishers. *Frontiers in Ecology and the Environment*
051 **15**:67-73.
- 052 Bornt KR, McLean DL, Langlois TJ, Harvey ES, Bellchambers LM, Evans SN, Newman SJ. 2015. Targeted
053 demersal fish species exhibit variable responses to long-term protection from fishing at the Houtman
054 Abrolhos Islands. *Coral Reefs* **34**:1297-1312.
- 055 Bradburn MJ, Deeks JJ, Berlin JA, Russell Localio A. 2007. Much ado about nothing: a comparison of the
056 performance of meta-analytical methods with rare events. *Statistics in medicine* **26**:53-77.
- 057 CALM. 1989. Ningaloo Marine Park Management Plan 1989 - 1999. Perth, WA, Australia.
- 058 CALM. 2005. Management Plan for the Ningaloo Marine Park and Muiron Islands Marine Management
059 Area 2005-2015: Management Plan No 52. Perth, WA, Australia.
- 060 Cappo MM, Brown II. 1996. Evaluation of sampling methods for reef fish populations of commercial,
061 recreational interest. CRC Reef Research Technical report No. 6.
- 062 Cheng J, Pullenayegum E, Marshall JK, Iorio A, Thabane L. 2016. Impact of including or excluding both-
063 armed zero-event studies on using standard meta-analysis methods for rare event outcome: a
064 simulation study. *BMJ Open* **6**.
- 065 Claudet J, Guidetti P. 2010. Improving assessments of marine protected areas. *Aquatic conservation: marine*
066 *and freshwater ecosystems* **20**:239-242.
- 067 Claudet J, Osenberg C, Domenici P, Badalamenti F, Milazzo M, Falcón JM, Bertocci I, Benedetti-Cecchi L,
068 García-Charton JA, Goñi R. 2010. Marine reserves: fish life history and ecological traits matter.
069 *Ecological applications* **20**:830-839.
- 070 Claudet J, Osenberg CW, Benedetti-Cecchi L, Domenici P, García-Charton JA, Pérez-Ruzafa Á,
071 Badalamenti F, Bayle-Sempere J, Brito A, Bulleri F. 2008. Marine reserves: size and age do matter.
072 *Ecology letters* **11**:481-489.
- 073 Coleman FC, Figueira WF, Ueland JS, Crowder LB. 2004. The Impact of United States Recreational
074 Fisheries on Marine Fish Populations. *Science* **305**:1958.
- 075 Collins LB, Zhu ZR, Wyrwoll K-H, Eisenhauer A. 2003. Late Quaternary structure and development of the
076 northern Ningaloo Reef, Australia. *Sedimentary Geology* **159**:81-94.
- 077 Côté IM, Mosqueira I, Reynolds JD. 2005. Effects of marine reserve characteristics on the protection of fish
078 populations: a meta-analysis. *Journal of Fish Biology* **59**:178-189.
- 079 Cowx IG, Cooke SJ. 2004. The Role of Recreational Fishing in Global Fish Crises. *BioScience* **54**:857-859.

- 080 Denny CM, Willis TJ, Babcock RC. 2004. Rapid recolonisation of snapper *Pagrus auratus*: Sparidae within
081 an offshore island marine reserve after implementation of no-take status. *Marine Ecology Progress*
082 *Series* **272**:183-190.
- 083 Depczynski MM, Tinkler PP, Cheal AA, Speed CC. 2015. Ningaloo fish communities. Page 121 in Miller
084 KK, Depczynski MM, Cappo MM, Wakeford MM, Speed CC, Stowar MM, Colquhoun JJ, Tinkler
085 PP, Cheal AA, and Fisher RR, editors. *Ningaloo and Outer Shark Bay Environmental Baseline*
086 *Survey 2014*. Report prepared by the Australian Institute of Marine Science for Woodside Energy
087 Ltd. Australian Institute of Marine Science, Townsville, Australia.
- 088 DPIRD DoPIaRD. 2018. Ningaloo Marine Park, Available from
089 [https://parks.dpaw.wa.gov.au/sites/default/files/downloads/parks/2462-](https://parks.dpaw.wa.gov.au/sites/default/files/downloads/parks/2462-13%20Spearfishing%20in%20Ningaloo%20WEB.pdf)
090 [13%20Spearfishing%20in%20Ningaloo%20WEB.pdf](https://parks.dpaw.wa.gov.au/sites/default/files/downloads/parks/2462-13%20Spearfishing%20in%20Ningaloo%20WEB.pdf) (accessed 4/11/2018 2018).
- 091 DPIRD DoPIaRDWA. 2017. Gascoyne Coast Bioregion, Status reports of the fisheries and aquatic resources
092 of Western Australia 2016/17.
- 093 Edgar GJ, Stuart-Smith RD, Willis TJ, Kininmonth S, Baker SC, Banks S, Barrett NS, Becerro MA, Bernard
094 AT, Berkhout J. 2014. Global conservation outcomes depend on marine protected areas with five key
095 features. *Nature* **506**:216-220.
- 096 Fisher R, Wilson Shaun K, Sin Tsai M, Lee Ai C, Langlois Tim J. 2018. A simple function for full-subsets
097 multiple regression in ecology with R. *Ecology and Evolution* **0**.
- 098 Fitzpatrick B, Harvey E, Langlois T, Babcock R, Twigg E. 2015. Effects of fishing on fish assemblages at
099 the reefscape scale. *Marine Ecology Progress Series* **524**:241-253.
- 100 Froese RaDP. 2018. FishBase, Available from www.fishbase.org (accessed 15/1/2018 2018).
- 101 Fulton CJ, Depczynski M, Holmes TH, Noble MM, Radford B, Wernberg T, Wilson SK. 2014. Sea
102 temperature shapes seasonal fluctuations in seaweed biomass within the Ningaloo coral reef
103 ecosystem. *Limnology and Oceanography* **59**:156-166.
- 104 Goetze J, Jupiter S, Langlois T, Wilson S, Harvey E, Bond T, Naisilisili W. 2015. Diver operated video most
105 accurately detects the impacts of fishing within periodically harvested closures. *Journal of*
106 *Experimental Marine Biology and Ecology* **462**:74-82.
- 107 Halpern BS, Warner RR. 2002. Marine reserves have rapid and lasting effects. *Ecology Letters* **5**:361-366.
- 108 Hedges LV, Gurevitch J, Curtis PS. 1999. The meta-analysis of response ratios in experimental ecology.
109 *Ecology* **80**:1150-1156.
- 110 Holmes TH, Wilson SK, Travers MJ, Langlois TJ, Evans RD, Moore GI, Douglas RA, Shedrawi G, Harvey
111 ES, Hickey K. 2013. A comparison of visual-and stereo-video based fish community assessment
112 methods in tropical and temperate marine waters of Western Australia. *Limnology and*
113 *Oceanography, Methods* **11**:337-350.
- 114 Horta e Costa B, Claudet J, Franco G, Erzini K, Caro A, Gonçalves EJ. 2016. A regulation-based
115 classification system for Marine Protected Areas (MPAs). *Marine Policy* **72**:192-198.
- 116 Jenkins CN, Joppa L. 2009. Expansion of the global terrestrial protected area system. *Biological*
117 *conservation* **142**:2166-2174.
- 118 Langlois TJ, Harvey ES, Fitzpatrick B, Meeuwig JJ, Shedrawi G, Watson DL. 2010. Cost-efficient sampling
119 of fish assemblages: comparison of baited video stations and diver video transects. *Aquatic biology*
120 **9**:155-168.
- 121 Lester SE, Halpern BS, Grorud-Colvert K, Lubchenco J, Ruttenberg BI, Gaines SD, Airamé S, Warner RR.
122 2009. Biological effects within no-take marine reserves: a global synthesis. *Marine Ecology Progress*
123 *Series* **384**:33-46.
- 124 Lewin W-C, Arlinghaus R, Mehner T. 2006. Documented and Potential Biological Impacts of Recreational
125 Fishing: Insights for Management and Conservation. *Reviews in Fisheries Science* **14**:305-367.
- 126 Lim IE, Wilson SK, Holmes TH, Noble MM, Fulton CJ. 2016. Specialization within a shifting habitat
127 mosaic underpins the seasonal abundance of a tropical fish. *Ecosphere* **7**:e01212.
- 128 Ma KY, Craig MT. 2018. An inconvenient monophyly: an update on the taxonomy of the groupers
129 (Epinephelidae). *Copeia* **106**:443-456.
- 130 Mackie M, Black R 1999. Research on Two Serranid Species (Serranidae: Epinephelinae) in Western
131 Australian Waters. University of Western Australia/Fisheries Research & Development Corporation.
- 132 Mallet D, Pelletier D. 2014. Underwater video techniques for observing coastal marine biodiversity: a
133 review of sixty years of publications (1952–2012). *Fisheries Research* **154**:44-62.

- 134 McCook LJ, et al. 2010. Adaptive management of the Great Barrier Reef: A globally significant
135 demonstration of the benefits of networks of marine reserves. *Proceedings of the National Academy*
136 *of Sciences* **107**:18278.
- 137 McLaren BW, Langlois TJ, Harvey ES, Shortland-Jones H, Stevens R. 2015. A small no-take marine
138 sanctuary provides consistent protection for small-bodied by-catch species, but not for large-bodied,
139 high-risk species. *Journal of Experimental Marine Biology and Ecology* **471**:153-163.
- 140 McLean DL, et al. 2016. Distribution, abundance, diversity and habitat associations of fishes across a
141 bioregion experiencing rapid coastal development. *Estuarine, Coastal and Shelf Science* **178**:36-47.
- 142 McPhee DP, Leadbitter D, Skilleter GA. 2002. Swallowing the bait: is recreational fishing in Australia
143 ecologically sustainable? *Pacific Conservation Biology* **8**:40-51.
- 144 Miller KI, Russ GR. 2014. Studies of no-take marine reserves: Methods for differentiating reserve and
145 habitat effects. *Ocean & Coastal Management* **96**:51-60.
- 146 Mitchell J, McLean D, Collin S, Taylor S, Jackson G, Fisher R, Langlois T. 2018. Quantifying shark
147 depredation in a recreational fishery in the Ningaloo Marine Park and Exmouth Gulf, Western
148 Australia. *Marine Ecology Progress Series* **587**:141-157.
- 149 Mora C, Sale PF. 2011. Ongoing global biodiversity loss and the need to move beyond protected areas: a
150 review of the technical and practical shortcomings of protected areas on land and sea. *Marine*
151 *ecology progress series* **434**:251-266.
- 152 Mosquera I, Côté IM, Jennings S, Reynolds JD. 2000. Conservation benefits of marine reserves for fish
153 populations. Pages 321-332. *Animal Conservation forum*. Cambridge University Press.
- 154 Murphy HM, Jenkins GP. 2010. Observational methods used in marine spatial monitoring of fishes and
155 associated habitats: a review. *Marine and Freshwater Research* **61**:236-252.
- 156 Osenberg CW, Sarnelle O, Cooper SD, Holt RD. 1999. Resolving ecological questions through meta-
157 analysis: goals, metrics, and models. *Ecology* **80**:1105-1117.
- 158 Osenberg CW, Shima JS, Miller SL, Stier AC. 2011. Ecology: assessing effects of marine protected areas:
159 confounding in space and possible solutions. *Marine protected areas: a multidisciplinary*
160 *approach*:143-167.
- 161 Pillans RD, Bearham D, Boomer A, Downie R, Patterson TA, Thomson DP, Babcock RC. 2014. Multi year
162 observations reveal variability in residence of a tropical demersal fish, *Lethrinus nebulosus*:
163 implications for spatial management. *PLoS One* **9**:e105507.
- 164 R Core Team. 2017. R: A language and environment for statistical computing. R Foundation for Statistical
165 Computing, Vienna, Austria.
- 166 Radford CA, Jeffs AG, Tindle CT, Cole RG, Montgomery JC. 2005. Bubbled waters: The noise generated
167 by underwater breathing apparatus. *Marine and freshwater behaviour and physiology* **38**:259-267.
- 168 Rees MJ, Knott NA, Davis AR. 2018a. Habitat and seascape patterns drive spatial variability in temperate
169 fish assemblages: implications for marine protected areas. *Marine Ecology Progress Series* **607**:171-
170 186.
- 171 Rees MJ, Knott NA, Neilson J, Linklater M, Osterloh I, Jordan A, Davis AR. 2018b. Accounting for habitat
172 structural complexity improves the assessment of performance in no-take marine reserves. *Biological*
173 *Conservation* **224**:100-110.
- 174 Russ GR. 2002. Yet another review of marine reserves as reef fishery management tools. *Coral reef fishes:*
175 *dynamics and diversity in a complex ecosystem* **24**:421.
- 176 Russ GR, Miller KI, Rizzari JR, Alcala AC. 2015. Long-term no-take marine reserve and benthic habitat
177 effects on coral reef fishes. *Marine Ecology Progress Series* **529**:233-248.
- 178 Ryan K, Hall N, Lai E, Smallwood C, Taylor S, Wise B 2015. State-wide survey of boat-based recreational
179 fishing in Western Australia 2013/14. Fisheries Research Division.
- 180 Ryan K, Hall N, Lai E, Smallwood C, Taylor S, Wise B. 2017. State-wide survey of boat-based recreational
181 fishing in Western Australia 2015/16.
- 182 Ryan K, Wise B, Hall N, Pollock K, Sulin E, Gaughan DJ 2013. An integrated system to survey boat-based
183 recreational fishing in Western Australia 2011/12. Fisheries Research Division, Western Australian
184 Fisheries and Marine Research Laboratories.
- 185 Sala E, Lubchenco J, Grorud-Colvert K, Novelli C, Roberts C, Sumaila UR. 2018. Assessing real progress
186 towards effective ocean protection. *Marine Policy* **91**:11-13.

- 187 Samoilys M, Fuentes H, Tuwai I, Tikomainiusiladi B, Leqata J, Oreihaka E, Mobiha A, Potuku T, Die D,
188 Connell S. 1995. Application of underwater visual census to assessing coral reef fish stocks in the
189 tropical Pacific. Report prepared for the Australian Centre for International Agricultural Research
190 (ACIAR). ACIAR Project.
- 191 Shears NT, Grace RV, Usmar NR, Kerr V, Babcock RC. 2006. Long-term trends in lobster populations in a
192 partially protected vs. no-take Marine Park. *Biological conservation* **132**:222-231.
- 193 Smallwood CB, Beckley LE. 2012. Spatial distribution and zoning compliance of recreational fishing in
194 Ningaloo Marine Park, north-western Australia. *Fisheries Research* **125**:40-50.
- 195 Spittal MJ, Pirkis J, Gurrin LC. 2015. Meta-analysis of incidence rate data in the presence of zero events.
196 *BMC medical research methodology* **15**:42.
- 197 Sumner NR, Williamson PC, Malseed BE 2002. A 12-month survey of recreational fishing in the Gascoyne
198 bioregion of Western Australia during 1998-99. Department of Fisheries, Western Australia.
- 199 Underwood A. 1993. The mechanics of spatially replicated sampling programmes to detect environmental
200 impacts in a variable world. *Australian Journal of ecology* **18**:99-116.
- 201 Viechtbauer W. 2005. Bias and efficiency of meta-analytic variance estimators in the random-effects model.
202 *Journal of Educational and Behavioral Statistics* **30**:261-293.
- 203 Viechtbauer W. 2010. Conducting meta-analyses in R with the metafor package. *J Stat Softw* **36**:1-48.
- 204 Watson DL, Harvey ES, Anderson MJ, Kendrick GA. 2005. A comparison of temperate reef fish
205 assemblages recorded by three underwater stereo-video techniques. *Marine Biology* **148**:415-425.
- 206 Watson M, Ormond R. 1994. Effect of an artisanal fishery on the fish and urchin populations of a Kenyan
207 coral reef. *Marine Ecology Progress Series*:115-129.
- 208 West L, Stark K, Murphy J, Lyle J, Ochwada-Doyle F. 2015. Survey of recreational fishing in New South
209 Wales and the ACT, 2013/14.
- 210 Westera MB. 2003a. The effect of recreational fishing on targeted fishes and trophic structure, in a coral reef
211 marine park.
- 212 Westera MB. 2003b. The effect of recreational fishing on targeted fishes and trophic structure, in a coral reef
213 marine park. Edith Cowan University, Perth, Western Australia.
- 214 Wilson S, Fulton C, Depczynski M, Holmes T, Noble M, Radford B, Tinkler P. 2014. Seasonal changes in
215 habitat structure underpin shifts in macroalgae-associated tropical fish communities. *Marine biology*
216 **161**:2597-2607.
- 217 Wilson SK, Babcock RC, Fisher R, Holmes TH, Moore JAY, Thomson DP. 2012. Relative and combined
218 effects of habitat and fishing on reef fish communities across a limited fishing gradient at Ningaloo.
219 *Marine Environmental Research* **81**:1-11.
- 220 Wilson SK, Depczynski M, Fisher R, Holmes TH, Noble MM, Radford BT, Rule M, Shedrawi G, Tinkler P,
221 Fulton CJ. 2018a. Climatic forcing and larval dispersal capabilities shape the replenishment of fishes
222 and their habitat-forming biota on a tropical coral reef. *Ecology and Evolution* **8**:1918-1928.
- 223 Wilson SK, Depczynski M, Holmes TH, Noble MM, Radford BT, Tinkler P, Fulton CJ. 2017. Climatic
224 conditions and nursery habitat quality provide indicators of reef fish recruitment strength. *Limnology*
225 *and Oceanography* **62**:1868-1880.
- 226 Wilson SK, Graham NAJ, Holmes T, MacNeil MA, Ryan N. 2018b. Visual versus video methods for
227 estimating reef fish biomass. *Ecological Indicators* **85**:146-152.
- 228 Wood SN. 2011. Fast stable restricted maximum likelihood and marginal likelihood estimation of
229 semiparametric generalized linear models. *Journal of the Royal Statistical Society: Series B*
230 *(Statistical Methodology)* **73**:3-36.
- 231 Zupan M, Fragkopoulou E, Claudet J, Erzini K, Horta e Costa B, Gonçalves EJ. 2018. Marine partially
232 protected areas: drivers of ecological effectiveness. *Frontiers in Ecology and the Environment*
233 **16**:381-387.
- 234 Zuur AF, Ieno EN, Elphick CS. 2010. A protocol for data exploration to avoid common statistical problems.
235 *Methods in ecology and evolution* **1**:3-14.
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Appendix G - Meta-analysis statistics

Table G1. Total heterogeneity statistics

Fish group	ABUNDANCE			BIOMASS		
	Q _T	df	P	Q _T	df	P
LETHRINIDAE	2002.57	300	<0.001	2318.26	263	<0.001
<i>L. nebulosus</i>	1971.07	256	<0.001	2886.99	222	<0.001
<i>L. atkinsoni</i>	1739.71	279	<0.001	1928.65	246	<0.001
EPINEPHELINAE	1125.65	276	<0.001	1307.49	252	<0.001
<i>E. rivulatus</i>	477.33	166	<0.001	590.76	155	<0.001
SCARINAE	1701.09	260	<0.001	1224.57	222	<0.001

Table G2 and G3 summarise the results of weighted mixed-effects meta-analyses for all seven variables and for *reserve identity* modelled individually. Figure 1 shows the predicted effect size for the cases where the moderator reserve identity explained a significant amount of heterogeneity for *habitat* so that it can be directly compared to the overall effect sizes in Figure 2 (main text).

Table G2. Mixed-effects models heterogeneity statistics for abundance data

	Fish group	ABUNDANCE					
		Model heterogeneity*			Residual heterogeneity		
		Q _m	df	P	Q _e	df	P
HABITAT	LETHRINIDAE	39.46	3	<0.001	1622.38	297	<0.001
	<i>L. nebulosus</i>	32.51	3	<0.001	1574.07	253	<0.001
	<i>L. atkinsoni</i>	14.55	3	<0.001	1614.93	276	<0.001
	EPINEPHELINAE	0.31	3	0.96	1117.33	273	<0.001
	<i>E. rivulatus</i>	5.39	3	0.15	467.33	163	<0.001
	SCARINAE	6	3	0.11	1589.43	257	<0.001
METHOD	LETHRINIDAE	4.09	2	0.13	1975.79	298	<0.001
	<i>L. nebulosus</i>	0.71	2	0.7	1946.56	254	<0.001
	<i>L. atkinsoni</i>	7.85	2	0.02	1676.16	277	<0.001
	EPINEPHELINAE	4.35	2	0.11	1094.35	274	<0.001
	<i>E. rivulatus</i>	0.98	2	0.61	474.68	164	<0.001
	SCARINAE	0.01	2	0.99	1633.03	258	<0.001
RESERVE SIZE	LETHRINIDAE	3.8	1	0.05	1959.06	299	<0.001
	<i>L. nebulosus</i>	0.01	1	0.94	1970.56	255	<0.001
	<i>L. atkinsoni</i>	8.11	1	<0.001	1688.35	278	<0.001
	EPINEPHELINAE	1.58	1	0.21	1112.23	275	<0.001
	<i>E. rivulatus</i>	0.47	1	0.49	477.08	165	<0.001
	SCARINAE	0.02	1	0.88	1701.09	259	<0.001
YEARS PROTECTION	LETHRINIDAE	0	1	0.95	2001.07	299	<0.001
	<i>L. nebulosus</i>	0.92	1	0.34	1949.89	255	<0.001
	<i>L. atkinsoni</i>	1.86	1	0.17	1736.82	278	<0.001
	EPINEPHELINAE	0.42	1	0.52	1115.08	275	<0.001
	<i>E. rivulatus</i>	5.43	1	0.02	462.46	165	<0.001
	SCARINAE	0.83	1	0.36	1689.03	259	<0.001
ZONING SCHEME	LETHRINIDAE	0.83	1	0.36	2002.31	299	<0.001
	<i>L. nebulosus</i>	0.87	1	0.35	1961.63	255	<0.001
	<i>L. atkinsoni</i>	8.43	1	<0.001	1696.16	278	<0.001
	EPINEPHELINAE	1.1	1	0.29	1100.82	275	<0.001
	<i>E. rivulatus</i>	13.43	1	<0.001	440.94	165	<0.001
	SCARINAE	3.82	1	0.05	1680.66	259	<0.001
BOAT FISHING	LETHRINIDAE	1.15	1	0.28	1987.99	295	<0.001
	<i>L. nebulosus</i>	0.7	1	0.4	1960.92	251	<0.001
	<i>L. atkinsoni</i>	0.55	1	0.46	1698.56	275	<0.001
	EPINEPHELINAE	0.01	1	0.92	1099.86	271	<0.001
	<i>E. rivulatus</i>	4.79	1	0.03	457.62	162	<0.001
	SCARINAE	5.9	1	0.02	1670.41	255	<0.001
SH OR FIS	LETHRINIDAE	7.09	1	0.01	1953.98	299	<0.001
	<i>L. nebulosus</i>	0.16	1	0.69	1966.75	255	<0.001
	<i>L. atkinsoni</i>	9.03	1	<0.001	1689.1	278	<0.001

	EPINEPHELINAE	3.9	1	0.05	1060.95	275	<0.001
	<i>E. rivulatus</i>	6.16	1	0.01	461.9	165	<0.001
	SCARINAE	5.59	1	0.02	1674.58	259	<0.001
RESERVE IDENTITY	LETHRINIDAE	43.22	21	<0.001	1746.07	279	<0.001
	<i>L. nebulosus</i>	48.82	21	<0.001	1665.43	235	<0.001
	<i>L. atkinsoni</i>	35.29	20	0.02	1494.98	259	<0.001
	EPINEPHELINAE	54.33	21	<0.001	880.04	255	<0.001
	<i>E. rivulatus</i>	68.95	20	<0.001	344.61	146	<0.001
	SCARINAE	27.81	21	0.15	1484.39	239	<0.001
SURVEY YEAR	LETHRINIDAE	2.39	1	0.12	1994.3	299	<0.001
	<i>L. nebulosus</i>	0	1	0.99	1962.36	255	<0.001
	<i>L. atkinsoni</i>	5.75	1	0.02	1674.53	278	<0.001
	EPINEPHELINAE	0.5	1	0.48	1115.22	275	<0.001
	<i>E. rivulatus</i>	3.59	1	0.06	468.95	165	<0.001
	SCARINAE	21.29	1	<0.001	1579.86	259	<0.001

* Total heterogeneity is provided in Table G1

Table G3. Mixed-effects models heterogeneity statistics for abundance data

		BIOMASS					
		Model heterogeneity*			Residual heterogeneity		
Fish group		Q _m	df	P	Q _e	df	P
HABITAT	LETHRINIDAE	41.75	3	<0.001	2082.5	260	<0.001
	<i>L. nebulosus</i>	29.38	3	<0.001	2743.83	219	<0.001
	<i>L. atkinsoni</i>	17.13	3	<0.001	1714.19	243	<0.001
	EPINEPHELINAE	1.21	3	0.75	1296.97	249	<0.001
	<i>E. rivulatus</i>	2.53	3	0.47	517.38	152	<0.001
	SCARINAE	22.86	3	<0.001	1038.84	219	<0.001
METHOD	LETHRINIDAE	2.46	2	0.29	2140.53	261	<0.001
	<i>L. nebulosus</i>	0.63	2	0.73	2829.86	220	<0.001
	<i>L. atkinsoni</i>	7.79	2	0.02	1720.61	244	<0.001
	EPINEPHELINAE	12.37	2	<0.001	1258.74	250	<0.001
	<i>E. rivulatus</i>	5.9	2	0.05	586.26	153	<0.001
	SCARINAE	1.16	2	0.56	1193.63	220	<0.001
RESERVE SIZE	LETHRINIDAE	1.3	1	0.25	2318.18	262	<0.001
	<i>L. nebulosus</i>	0.1	1	0.75	2886.72	221	<0.001
	<i>L. atkinsoni</i>	1.97	1	0.16	1925.76	245	<0.001
	EPINEPHELINAE	3.78	1	0.05	1307.41	251	<0.001
	<i>E. rivulatus</i>	1.32	1	0.25	571.52	154	<0.001
	SCARINAE	0.59	1	0.44	1220.28	221	<0.001
YEARS PROTECTION	LETHRINIDAE	0.01	1	0.94	2312.18	262	<0.001
	<i>L. nebulosus</i>	0.71	1	0.4	2882.59	221	<0.001
	<i>L. atkinsoni</i>	3.13	1	0.08	1924.77	245	<0.001
	EPINEPHELINAE	0.12	1	0.73	1307.3	251	<0.001
	<i>E. rivulatus</i>	5.01	1	0.03	587.67	154	<0.001
	SCARINAE	0.02	1	0.89	1218.81	221	<0.001
ZONING SCHEME	LETHRINIDAE	0.44	1	0.51	2314.92	262	<0.001
	<i>L. nebulosus</i>	0.54	1	0.46	2883.99	221	<0.001
	<i>L. atkinsoni</i>	9.59	1	<0.001	1925.41	245	<0.001
	EPINEPHELINAE	0.04	1	0.83	1307.27	251	<0.001
	<i>E. rivulatus</i>	9.96	1	<0.001	587.78	154	<0.001
	SCARINAE	3.09	1	0.08	1222.86	221	<0.001
BOAT FISHING	LETHRINIDAE	0.18	1	0.67	2313.23	258	<0.001
	<i>L. nebulosus</i>	0.04	1	0.84	2879.67	217	<0.001
	<i>L. atkinsoni</i>	0.14	1	0.7	1923.96	243	<0.001
	EPINEPHELINAE	0.82	1	0.37	1287.94	247	<0.001
	<i>E. rivulatus</i>	3.75	1	0.05	578.7	151	<0.001
	SCARINAE	0.57	1	0.45	1193.33	217	<0.001
SHORE FISHING	LETHRINIDAE	2.19	1	0.14	2295.06	262	<0.001
	<i>L. nebulosus</i>	0	1	0.96	2606.71	221	<0.001
	<i>L. atkinsoni</i>	5.25	1	0.02	1927.59	245	<0.001
	EPINEPHELINAE	0.38	1	0.54	1268.07	251	<0.001
	<i>E. rivulatus</i>	3.05	1	0.08	580.64	154	<0.001
	SCARINAE	5.78	1	0.02	1168.18	221	<0.001
RESERVE IDENTITY	LETHRINIDAE	29.09	21	0.11	2019.62	242	<0.001
	<i>L. nebulosus</i>	27.02	21	0.17	1941.89	201	<0.001
	<i>L. atkinsoni</i>	33.93	20	0.03	1846.92	226	<0.001
	EPINEPHELINAE	58.1	21	<0.001	987.83	231	<0.001
	<i>E. rivulatus</i>	49.47	20	<0.001	503.04	135	<0.001

SURVEY YEAR	SCARINAE	33.11	21	0.04	1046.15	201	<0.001
	LETHRINIDAE	2.39	1	0.12	1994.3	299	<0.001
	<i>L. nebulosus</i>	0	1	0.99	1962.36	255	<0.001
	<i>L. atkinsoni</i>	5.75	1	0.02	1674.53	278	<0.001
	EPINEPHELINAE	0.5	1	0.48	1115.22	275	<0.001
	<i>E. rivulatus</i>	3.59	1	0.06	468.95	165	<0.001
	SCARINAE	21.29	1	<0.001	1579.86	259	<0.001

* Total heterogeneity is provided in Table G1

Figure G1 shows the transformed predicted effect sizes as relative fish abundance by *habitat* while Figure G2 shows this for *reserve identity*.

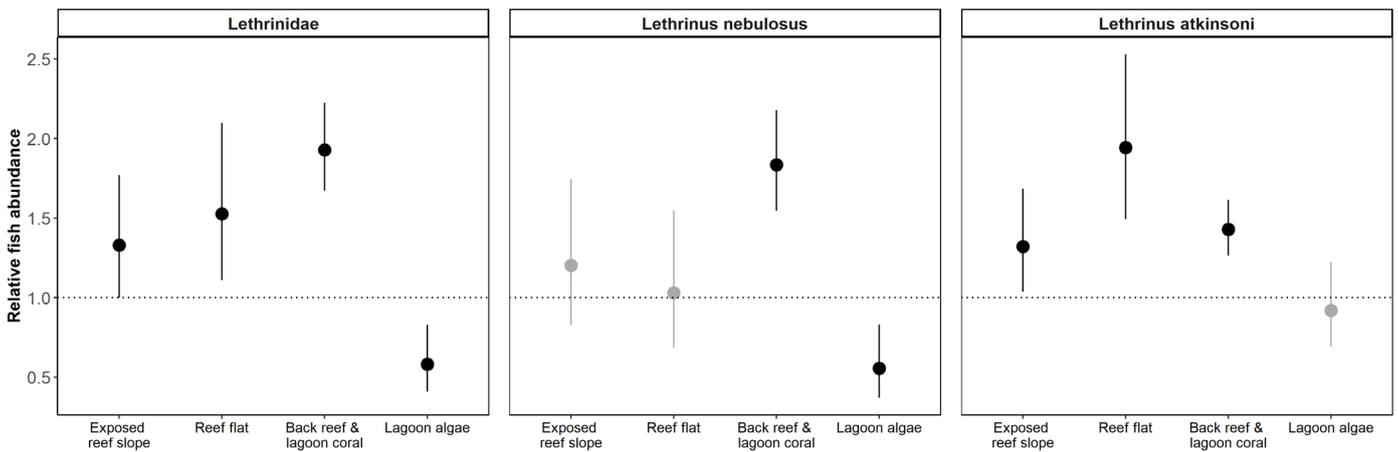
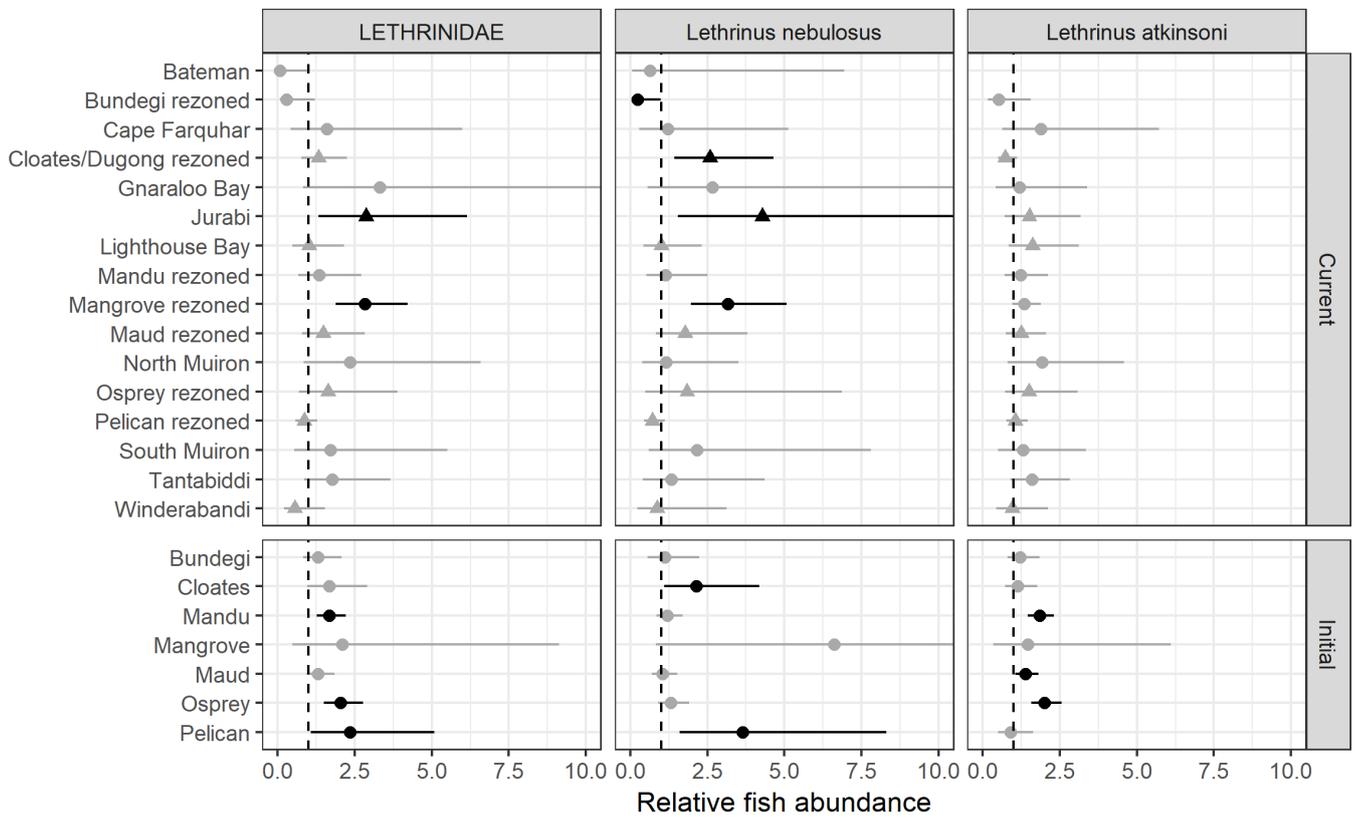
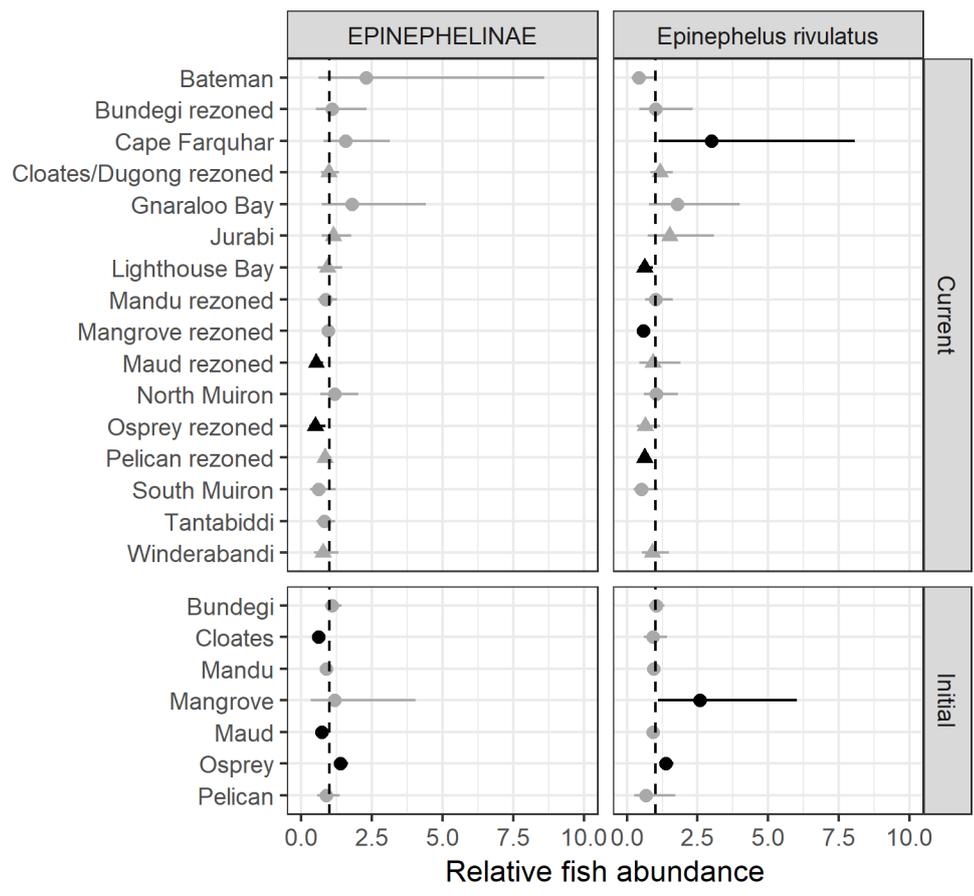


Figure G1. Relative fish abundance inside to outside NTMRs (back-transformed weighted mean effect sizes) with 95% confidence intervals) for the Lethrinidae, *Lethrinus nebulosus*, and *L. atkinsoni*, the fish groups for which *habitat* explained a significant amount of variation. Effect sizes are considered significant when the confidence intervals do not include one. Black dots correspond to significant effects while grey dots correspond to non-significant effects.

Figure G2 is included to show that there is heterogeneity between individual NTMRs and because it may be useful to managers of the Ningaloo Marine Park.



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282 Figure G2. Relative fish abundance (Transformed weighted mean effect sizes ($\exp(e)$) with 95%
 283 confidence intervals (CI)), for the Lethrinidae, *Lethrinus nebulosus*, *Lethrinus atkinsoni*, Epinephelinae and
 284 *Epinephelus rivulatus*, the fish groups for which *reserve identity* explained a significant amount of variation.
 285 Results are categorised into panels based on the initial or current zoning. Point shape indicates whether

286 shore fishing is allowed or prohibited along the coastal edge of the reserve (circle = prohibited; triangle =
287 allowed). Effect sizes are considered significant when the confidence intervals do not include one. Black
288 dots correspond to significant effects while grey dots correspond to non-significant effects.
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293 *Appendix H – Biomass results*

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296 **Table H1.** Top Generalised Additive Mixed Models (GAMMs) for predicting the response ratio inside to
297 outside NTMRs, \bar{E} , for biomass from full subset analyses for the abundance of the six fish groups.
298 Difference between the lowest reported corrected Akaike Information Criterion (ΔAICc), AICc weights
299 (ωAICc), variance explained (R^2) and estimated degrees of freedom (EDF) are reported for model
300 comparison. Model selection was based on the most parsimonious model (fewest variables and lowest EDF)
301 within two units of the lowest AICc. This model is shown in bold text.

Fish group	Model	ΔAICc	ωAICc	R^2	EDF
LETHRINIDAE	Habitat + Size by Habitat	0	0.855	0.17399	9
<i>L. nebulosus</i>	Habitat + Years protection by Habitat + Size by Habitat	0	0.443	0.19863	14.51
	Habitat + Size by Habitat	0.857	0.289	0.17016	9
	Years protection + Habitat + Size by Habitat	1.301	0.231	0.17594	10.66
<i>L. atkinsoni</i>	Habitat + Method + Years protection by Habitat	0	0.29	0.10828	12.98
	Habitat + Years protection by Habitat	0.39	0.238	0.09662	10.67
	Habitat + Method + Zoning scheme	1.273	0.153	0.08162	8
EPINEPHELINAE	Years protection + Boat fishing + Method	0	0.196	0.09835	7.81
	Boat fishing + Method	0.516	0.152	0.09171	6.68
	Years protection + Method	1.092	0.114	0.09192	7.24
	Method	1.198	0.108	0.08585	5.88
	Boat fishing + Method + Shore fishing	1.751	0.082	0.09098	7.61
	Boat fishing + Method + Zoning scheme	1.955	0.074	0.09064	7.71
<i>E. rivulatus</i>	Boat fishing + Method + Zoning scheme	0	0.546	0.17358	12.31
	Years protection + Boat fishing + Method	1.457	0.264	0.16292	11.96
SCARIDAE	Years protection + Habitat + Size by Habitat	0	0.271	0.12181	16.44
	Habitat	1.354	0.138	0.05738	5
	Years protection + Habitat + Size	1.406	0.134	0.09791	13.04
	Habitat + Zoning scheme	1.698	0.116	0.06077	6
	Habitat + Size by Habitat	1.889	0.106	0.07496	9
	Habitat + Size	1.952	0.102	0.06103	6.28

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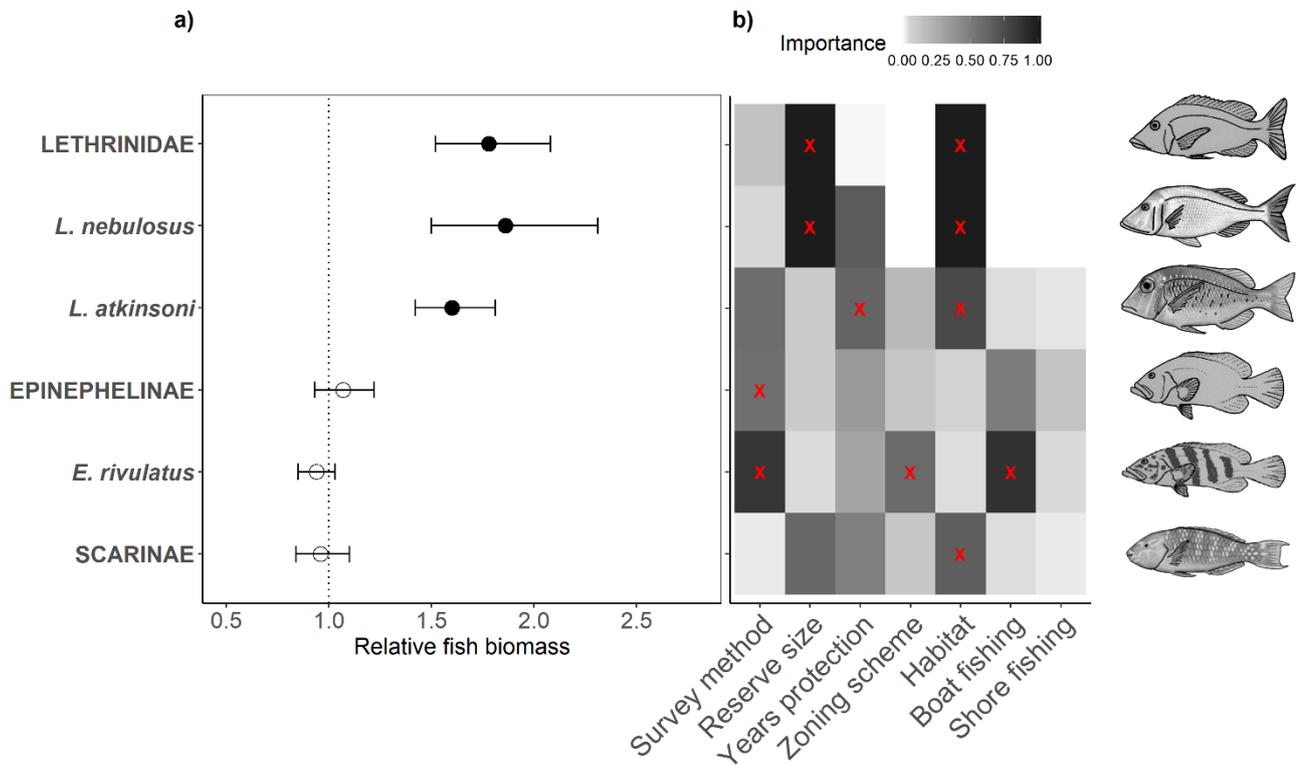
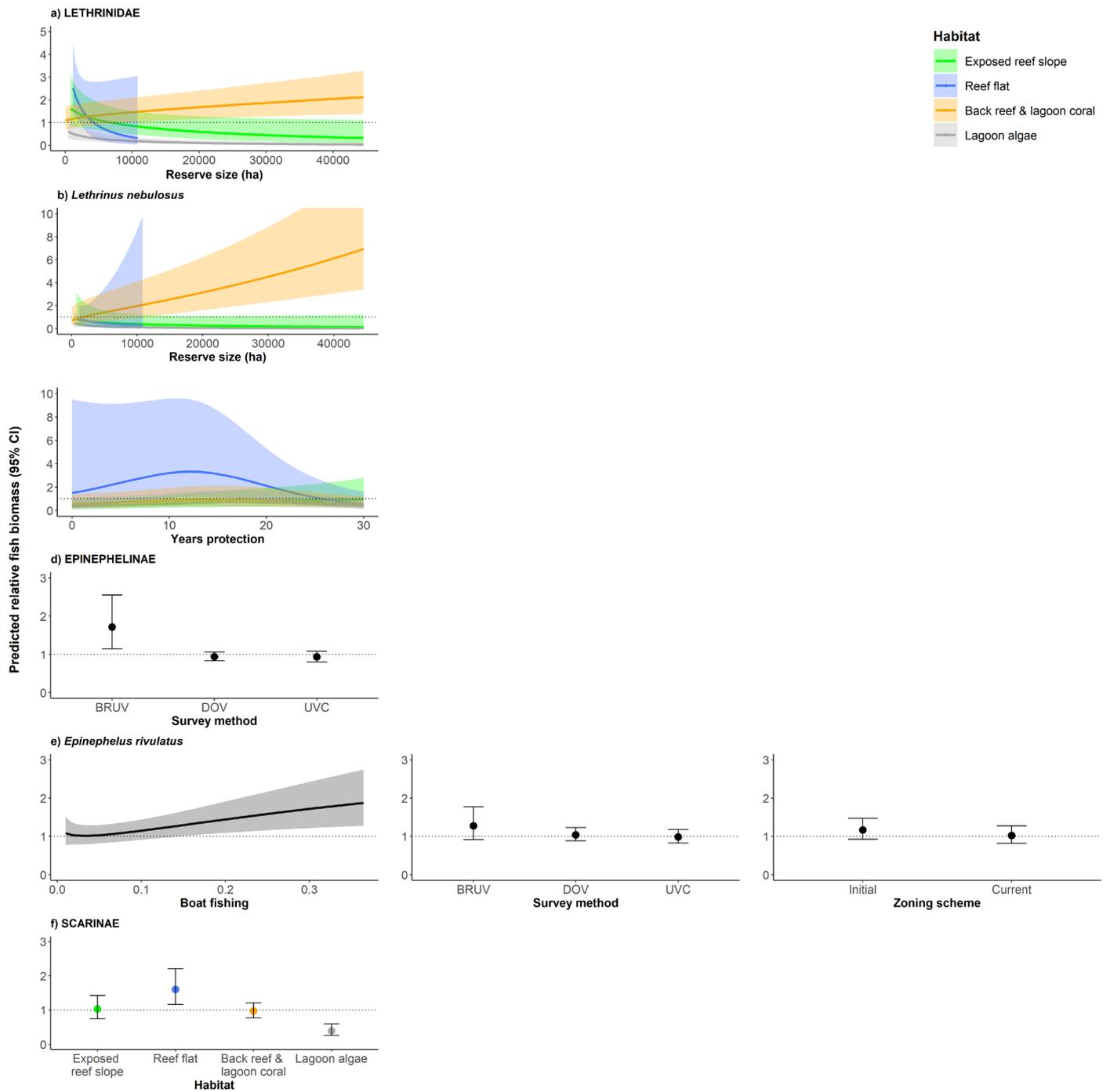


Figure H1. a) Relative fish biomass (Transformed weighted mean effect sizes ($\exp(\bar{E})$) with 95% confidence intervals (CI)), for the relative abundance of the six fish taxa: Lethrinidae, *Lethrinus nebulosus*, *L. atkinsoni*, Epinephelinae, *Epinephelus rivulatus* and Scarinae. Effect sizes are considered significant when the confidence intervals do not include one. Open dots correspond to non-significant effects. Sample sizes are given in Appendix G. b) Importance scores (based on summed Akaike weights corrected for finite samples (AICc)) from full-subsets analyses exploring the influence of seven variables on the overall effect size for each fish taxa. *Reserve size* was square-root transformed and *boat fishing* was log-transformed in all models. Red X symbols mark the variables that were included in the most parsimonious models for each fish taxa (also see Table H1 and Fig. H2).

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Figure H2. Predicted relative fish biomass inside to outside NTMRs (back-transformed predicted weighted effect sizes) with 95% confidence intervals) for the six fish groups – a) Lethrinidae; b) *Lethrinus nebulosus*; c) *Lethrinus atkinsoni*; d) Epinephelinae; e) *Epinephelus rivulatus*; f) Scarinae for abundance– as a function of variables present in the most parsimonious models (Table G4) from full-subsets GAMM analysis.

Ribbons represent 95% confidence intervals