

1 Measuring heterogeneity in soil networks: a network analysis and  
2 simulation-based approach

3

4 Natalie Davis <sup>a, b, \*</sup>

5 J. Gareth Polhill <sup>b</sup>

6 M. J. Aitkenhead <sup>b</sup>

7 <sup>a</sup> Lancaster Environment Centre, Lancaster, UK.

8 <sup>b</sup> The James Hutton Institute, Aberdeen, UK.

9 \* Corresponding author: ndavis.research@gmail.com

10

11 **Abstract**

12

13 Quantifying soil structural and ecological heterogeneity is crucial for understanding their interactions  
14 and their relationships to the resilience and health of the wider ecosystem. However, a clear  
15 understanding of how structural heterogeneity affects soil biodiversity is still emerging. Previous  
16 work has primarily used expensive, often laboratory-based methods to quantify soil pore network  
17 structure, and typically separated study of structural and biological dimensions. Here, we test  
18 whether standard network metrics can be used to quantify structural heterogeneity in soil pore  
19 networks, and how this network structure, along with characteristics of the consumer and resource  
20 populations, affects the heterogeneity of a population of consumers. Specifically, we extract  
21 simplified soil pore networks from digital photographs of soil profiles and apply established metrics  
22 from network science and transport geography to quantify and compare the networks. The  
23 networks are also used as the medium for an agent-based model of generalised consumers, to  
24 analyse the effects of consumer and resource parameterisations and network structure. Combining  
25 network analysis and simulation modelling in this way can provide insights on the structure,  
26 function, and diversity possible in the soil, as well as avenues for exploring the impact of future  
27 structural or environmental changes.

28 **Keywords:** soil networks; soil structure; digital soil morphometrics; agent-based model; network  
29 analysis; ecological heterogeneity.

30

# 31 1. Introduction

32 The distribution of energetic resources in an ecosystem plays a key role in determining the  
33 complexity, quantity, and behaviour of organisms that it can support (e.g. Giller, 1996; Tews *et al.*,  
34 2004; Roshier, Doerr and Doerr, 2008; Stevens & Tello, 2011). To understand these systems more  
35 fully, and inform actions to protect those relying on them, we must understand how resource  
36 distribution networks develop and function. For example, resource location and movement can  
37 create heterogeneity that allows species to specialise and differentiate (e.g. Bardgett, Yeates and  
38 Anderson, 2009; Tews *et al.*, 2004; Stevens & Tello, 2011), as well as cause inequality among  
39 individuals of the same species, topics that are relevant for both biologists and ecologists.

40 The soil provides a unique and diverse ecosystem in which to study resource distribution, and its  
41 effect on organisms. Soil structure can be defined as the collection of soil particles and pore space  
42 among them (Oades, 1993). This pore space provides access to nutrients stored on the surface of soil  
43 particles, allows for preferential flow of water through the soil matrix, and serves as the resource  
44 distribution network through which micro-, meso-, and macrofauna (soil biota) forage. As this  
45 structure determines how air, water, and soil biota move through the soil, it allows or impedes the  
46 foraging of organisms, regulates the air and water balance in the soil matrix, and affects chemical  
47 signals used in foraging, such as those of bacterial decomposition (Young and Ritz, 2009).  
48 Furthermore, crevices and niches along soil pores provide habitats for smaller microbes to avoid  
49 predation, and the overall spatial and temporal heterogeneity of the soil environment allows for  
50 resource partitioning and habitat specialisation that limits the effect of competitive exclusion  
51 (Bardgett, Yeates and Anderson, 2009). This is similar to the hypothesised effect of heterogeneity in  
52 aboveground habitats (e.g. Tews *et al.*, 2004; Stevens and Tello, 2011).

53 Soil biota in turn can increase the porosity of soil, through burrowing and consuming organic matter,  
54 and releasing gases during decomposition, which create or expand soil pores (Kravchenko and  
55 Guber, 2017). Additionally, there is evidence of feedbacks between the soil biota and aboveground  
56 plant communities (e.g. Baer *et al.*, 2005; Wijesinghe, John and Hutchings, 2005; García-Palacios *et al.*,  
57 2012), which alter soil structure as their roots burrow in pore networks, and roots and hyphae  
58 bind and stabilise soil particles (Vezzani *et al.*, 2018). Through regulating movement and diffusion of  
59 water and energy resources, gases, and fauna in the soil matrix; providing habitat; and mediating  
60 biological feedbacks; soil structure is the foundation of all earth systems.

61 Past efforts to quantify and model soil structure have primarily focussed on measuring the stability  
62 of soil, by utilising soil aggregate size distribution as a measure of structure. While this does  
63 represent the spatial distribution in the soil, it is not a complete representation of physical  
64 properties (see e.g. Young, Crawford and Rappoldt, 2001). Methods for visualising the pore network  
65 within a soil sample include CT scans and X-ray tomography, NMR, and SPECT scanning, mostly for  
66 the purposes of measuring solute flow and transport processes (see review in Young, Crawford and  
67 Rappoldt, 2001). Gas diffusion and solute flow have also been examined with modelling approaches,  
68 including neural networks, Boolean models, and cellular automata. Additionally, fractal modelling  
69 has also been used successfully to quantify the degree of connectivity, tortuosity, and heterogeneity  
70 of the soil pore network (Crawford, Ritz and Young, 1993), three characteristics that have also been  
71 associated with a higher level of heterogeneity of resource distribution in generalised networks  
72 (Davis *et al.*, 2020).

73 Overall, past work has highlighted the important connections between soil function and structure,  
74 especially of the pore network. Much of this work has been done from a geometric or hydrological  
75 perspective, however, rather than an energetic one, leading to criticisms of unrealistic separation of

76 soil physics and biology, and emphasis on the importance of integrating these spatially explicit  
77 approaches in future soil ecology research (Bardgett, Yeates and Anderson, 2009). Additionally,  
78 much of the imaging equipment required for the techniques above is large and expensive, requiring  
79 soil samples to be brought back to the laboratory. Even if disruption to the soil structure during  
80 extraction and transport is minimised, these methods are more suitable for intensive analyses of  
81 individual samples and smaller areas.

82 In contrast, some previous work has focussed on quantifying the structure of soil networks through  
83 image morphology techniques applied to a photograph of a sample, in order to extract the relevant  
84 network (e.g. Velde, Moreau and Terribile, 1996; Gargiulo, Mele and Terribile, 2013; Hartemink and  
85 Minasny, 2014). This method will not reveal the network at the same level of detail as CT scans or X-  
86 ray tomography, and may require use of resins and dyes to highlight the underlying structure  
87 (Hartemink and Minasny, 2014). Good arguments have also been raised regarding the importance of  
88 analysing soil structure from a three-dimensional perspective, as it reveals considerably more about  
89 the habitat of the soil (Young and Ritz, 2009). However, if rotational invariance is assumed,  
90 connectivity and structure of a two-dimensional sample can be assumed representative of any  
91 random two-dimensional plane taken through the system. This inference does not consider lateral  
92 flow, which would undoubtedly play an important influence in sloping areas by transporting  
93 nutrients laterally through the soil. In areas where the surface is flat and lateral flow effects are  
94 negligible, standard network metrics could usefully approximate soil structure and provide insights  
95 into its effect on biotic and abiotic processes within an environment.

96 Moreover, the two-dimensional techniques are considerably more portable and feasible than the  
97 three-dimensional techniques, and processing time can be significantly faster. Image analysis  
98 methods, particularly those that can be performed entirely in the field, could potentially be  
99 incorporated into software for use by farmers and researchers who may otherwise not have access  
100 to the equipment necessary for the more costly and lab-intensive methods of quantifying structure  
101 (e.g. Aitkenhead *et al.*, 2016). These methods could also act as preliminary investigations to highlight  
102 potential areas of future exploration using more intensive analyses.

103 In this paper, we test whether standard network metrics can be used to quantify structural  
104 heterogeneity in soil pore networks, and how this network structure, along with characteristics of  
105 the consumer and resource populations, affects the heterogeneity of a population of consumers.  
106 Specifically, we develop a method for extracting approximate soil networks from digital photographs  
107 using image morphology techniques, then apply metrics from network science and transport  
108 geography to quantify and compare the networks. The networks are also used as the medium for an  
109 agent-based model (ABM, which in ecology is more typically known as an individual-based model,  
110 e.g. Grimm *et al.*, 2006), where the agents represent generalised consumers who explore the  
111 network and consume food resources. The variation in population size and resource consumption is  
112 compared across simulations, to evaluate how both the network structure and simulation  
113 parameters affect outcomes of the biotic community. This methodology is applied to a case study  
114 using soil images from two test sites in Aberdeenshire, United Kingdom.

115

## 116 2. Methods

### 117 2.1 Soil image collection

118 Images were taken at two field sites in Aberdeenshire, United Kingdom. The first site had a brown  
119 forest soil, or Cambisol (Fig A1a); photographs were taken from seven locations in both forested and  
120 converted agricultural areas. The second site had a sandy beach soil, or Arenosol (Fig A1b);  
121 photographs were taken at five locations across a dune area, with sparse grass and shrub cover.  
122 Neither Cambisols nor Arenosols are highly developed, but Cambisols have some diagnostic features,  
123 while Arenosols are lacking diagnostic features and are defined only on the basis of being coarse  
124 (sandy) textured (FAO, 2015). The known difference between the two soils therefore provides a  
125 basis for preliminarily evaluating the methodology. Additionally, both soil types can be assumed to  
126 show limited profile variation with depth on the scale of the observed soil profile sections under  
127 study (FAO, 2015), such that a uniform network extraction method and analysis can be applied  
128 across the image. The specific sampling sites were also chosen as they provided easy access to  
129 multiple sampling locations for both soil types. As this work is an exploratory proof-of-concept, an  
130 exhaustive sampling regime across different soil, land use, and geographic regions was not  
131 undertaken.

132 The methodology for taking pictures was replicated from Aitkenhead *et al.* (2016). In summary, the  
133 photographs were taken of the soil profile of shallow (30 cm) pits in flat areas, using an angle that  
134 provided maximum natural light and minimum shadow (Fig A1a, b). No artificial lighting was  
135 required during photography. Additionally, each photograph included a 10 cm x 6 cm colour  
136 correction card within the frame. Colour correction has been used in past work (e.g. Aitkenhead *et*  
137 *al.*, 2016) to correct colour variation in ambient lighting. However, in this work we were only  
138 interested in overall intensity, rather than light balance, so the cards were inserted into the image to  
139 provide a spatial scale reference for future work.

140 In Fig A1a and A1b, the white area is an excised section of the image that is larger than the  
141 correction card. The imaging was taken with the card viewed straight on, without distortion, so the  
142 image distortion and impact on length of edges is not an issue. Extracting an area larger than the  
143 correction card also attempted to eliminate shading effects around the card. This may not have been  
144 done sufficiently to eliminate all the shading, possibly introducing some additional dark pixels and  
145 error into the network metric calculations. However, taking multiple pictures within the same profile  
146 can provide some robustness against this. Future work should attempt to remove this effect from  
147 near the correction card.

148 In total, seven Cambisol profiles and five Arenosol pits were used for each soil type, with several  
149 images taken of the profile of each pit. In taking multiple images from each soil pit, we moved the  
150 camera slightly to present different viewing angles and thus generate different images. This was  
151 done to compare the robustness of extracted networks from each pit (see Section 2.2), and  
152 replication within pit was considered in all statistical analyses.

### 153 2.2 Network extraction

154 To extract the approximate soil network structure from the photographs, the photographs were  
155 converted to text files containing the red, green, and blue (RGB) triplet values for each pixel. All non-  
156 soil pixels were then identified as those whose triplet values exceeded the ranges expected for soil  
157 particles, based on the average of the rest of the image. Using the average to determine this  
158 threshold customised it slightly for each image, so that outliers such as roots and rocks specific to  
159 that sample were captured, but samples having an overall more reddish tone were not stripped

160 completely. The identified non-soil particles were removed, and variations in brightness across the  
161 remaining pixels were standardised using the mean pixel intensity.

162 As soil structure and porosity are only loosely related, soils of the same porosity can have different  
163 structural properties. A common assumption made is that soils, unless compressed/compacted,  
164 have up to 50 % pore space. As the pixel resolution of the images here is between 0.3 – 0.5 mm, and  
165 therefore much higher than the smallest pore space possible (sub-micron scale), it follows that the  
166 pore space actually visible is less than this 50 %. An evaluation of the distribution of pixel values  
167 showed that for soil profile images used in this study, the greatest change in the distribution  
168 occurred around a pixel intensity where 30 – 40 % of the pixels were below this value (Fig A2). We  
169 have therefore assumed that 30 % of the soil is 'void' (i.e. dark pixels). Therefore, the darkest 30 % of  
170 the soil pixels were retained as pores, and the image was inverted to convert these darker pixels to  
171 white, and vice versa (Fig A1e, f). The images from the same profile were visually compared after  
172 thresholding and showed a high degree of agreement in the pores identified (e.g. Fig A3). Network  
173 outlines were then drawn through a process known in image morphology as 'skeletonization,' where  
174 lines of white pixels were iteratively stripped down until they were all one pixel in width (Fig. A1g, h).  
175 We then mapped the networks to a list of links, which were series of pixels that were more than one  
176 pixel long, and nodes, defined as junction points between two or more links. Redundant links  
177 between nodes were removed.

178 For simplicity, all links in the final networks were represented with straight lines along the shortest  
179 distance between two nodes. This lost some of the details of the topology, such as pore size and  
180 shape. However, this work intended to create an abstraction of the network taken from the soil,  
181 rather than replicate and analyse the exact soil structure itself. This emphasised overall soil  
182 structural characteristics and heterogeneity, rather than modelling how specific transport processes  
183 and biological activities would occur. Replicating the exact soil network would also have markedly  
184 increased the computational burden, as link lengths would have had to be calculated through pixel-  
185 counting rather than the Euclidean geometry measuring shortest distances. As many of the links as  
186 represented were quite short (see Results), the difference between the true link length and the  
187 shortest distance between nodes was assumed to be negligible. Currently, we assume that the  
188 method requires further validation and improvement to provide a measure of soil structure that can  
189 be used in soil science or pedological characterisation of the soil. We also assume however, that the  
190 method, while not perfect in its current form, provides sufficient quality of network data to allow  
191 simplified networks to be extracted and analysed, and used as the basis for simulations.

192 The process of rendering the network also identified which sections of the network were fully  
193 connected, and which nodes were part of disconnected subnetworks (Fig. A1i, j). An outline of the  
194 image morphology process, and images of each step, are available in Appendix 1.

### 195 2.3 Network analysis

196 Two types of analysis were used to quantify the heterogeneity present in the soil network images.  
197 The first involved applying metrics adapted from network science and transport geography to  
198 measure structural characteristics of the abstracted networks, which allows for easy comparison  
199 among soil types. These were calculated using R v4.0.2 (R Core Team, 2020), including the packages  
200 *igraph*, *qgraph*, and *sp* (Pebesma and Bivand, 2005; Csardi and Nepusz, 2006; Epskamp *et al.*, 2012;  
201 Bivand, Pebesma and Gomez-Rubio, 2013). All additional data analysis and visualisations were also  
202 done in R, using the packages *ARTool* v0.10.7 (Kay and Wobbrock, 2020; Wobbrock *et al.*, 2011),  
203 *emmeans* v1.5.0 (Lenth, 2020), *ImerTest* v3.1.2 (Kuznetsova, Brockhoff, and Christensen, 2017),  
204 *dunn.test* v1.3.5 (Dinno, 2017), *rcompanion* v2.3.25 (Mangiafico, 2020), *dplyr* v1.0.0 and *ggplot2*

205 v3.3.2 packages (Wickham, 2016; Wickham *et al.*, 2019). The scripts for calculating network metrics  
 206 are available at (Davis, 2020).

207 A brief description of each of the metrics chosen is given in Table 1. These were chosen to measure  
 208 the size, connectivity, and structural heterogeneity of the networks from a range of node-centric,  
 209 link-centric, and global perspectives, to obtain a broad picture how the networks may differ. The  
 210 metrics chosen also minimised assumptions about inaccessibility of the soil matrix between pores:  
 211 for example, the convex hull area was chosen over the concave hull area as the former is a more  
 212 generous estimate of the spatial area.

213 **Table 1. The name and description of the metrics used to analyse the soil networks.**

Metric name	Description	Type of measure	Reference
Mean and standard deviation (SD) of link length	Quantifies the typical length and variability of lengths included within the network.	Size	N/A
Beta index	The ratio of links to nodes.	Connectivity	Rodrigue, 2017
Gamma index	Number of observed vs. possible links: $nLinks / (nNodes * (nNodes - 1))$	Connectivity	Rodrigue, 2017
Diameter	The length of the longest geodesic (shortest path between two nodes) in the network – the shortest path between the two most distant nodes.	Size	Rodrigue, 2017
Node count	The number of nodes in the network.	Size	Barabási, 2016
Edge count	The number of edges (links) in the network.	Size	Barabási, 2016
Mean node degree	Mean number of links per node.	Connectivity	Barabási, 2016
Cost	The total length of the network measured in real transport distances.	Size	Rodrigue, 2017
Global reach centrality (GRC)	The difference between the maximum and average local reach centrality (LRC), where the LRC is the nodes that a given node can connect to, weighted by distance (here, spatial distance).	Structure, connectivity	Adapted from Mones, Vicsek and Vicsek (2012)
Mean convex hull area	The area of a polygon that minimally encompasses every node in the network.	Size, connectivity	Rockafellar, 1970
Network density	The ratio of the number of nodes to the convex hull area.	Structure	N/A

214 As introduced, the imaging method and metrics used here are two-dimensional (2D), and we have  
215 been unable to find literature describing characterisations of three-dimensional (3D) soil structure  
216 metrics based on two-dimensional imaging. Aitkenhead *et al.* (1999) derived 3D models of soil pore  
217 systems based on 2D metrics but did not compare the two sets of structural metrics. Future work  
218 would be necessary to determine the extent to which 3D variation in soil structural metrics  
219 correlates to the variation seen in 2D. Here, we are assuming that it does correlate, and that this  
220 allows 2D imaging to provide structural metrics representative of different soil types.

221 We calculated each metric for each of the networks, which contained all nodes and links in the  
222 image, hereon called 'main networks.' We also calculated each metric for each of the disconnected  
223 subnetworks within the main networks, hereon called 'subnetworks.' As the distributions of metrics  
224 in the main soil networks had similar variance across soil types and relatively normal distributions,  
225 these were compared with nested ANOVA, using profile ID as a random effect to account for  
226 replication. The distribution of metrics across the subnetworks did not meet the assumptions for  
227 classical ANOVA, so non-parametric Aligned-Ranks Transformation (ART) ANOVAs were used instead,  
228 also with profile ID as a random effect.

## 229 2.4 Agent-based model overview

230 The second analytical method used a simulated population of consumers to explore each network,  
231 using the resulting heterogeneity in consumer resource stocks to further elucidate the heterogeneity  
232 of the network. This provided a more functional perspective, alongside the structural quantification  
233 of the network metrics. The purpose was to investigate the structure's generalised impact, rather  
234 than test the precision of this model in predicting outcomes for real species. Therefore, rather than  
235 using parameterisations that reflected specific species or groups, five generic model species with  
236 different sets of values for each trait were used, similarly to e.g. Polhill and Gimona (2014).

237 The same approach was taken for resources, with three sets of resource bases of different  
238 combinations of maximum capacity and maximum growth rates. Resources were assumed to be  
239 located at nodes within the network, as identified during the extraction process (see Section 2.2).  
240 Food resources in real soil networks are located throughout the soil matrix, but are often  
241 concentrated in 'hotspots' such as those created by plant roots and decomposition processes  
242 (Ettema and Wardle, 2002), which would be represented in the networks here as nodes. As  
243 exploring the effect of size of the generic species was not in scope for the work here, only the most  
244 accessible areas of the network were treated as potential resources.

245 A brief description of the model purpose, variables, and processes is presented below, following the  
246 Overview, Design concepts, and Details (ODD) protocol (Grimm *et al.*, 2006, 2010). The full ODD  
247 document, including description of design concepts, initialisation, input data, and sub-models, is  
248 available in Appendix 2. The model source code, written in NetLogo 6.1, is available in the Modelling  
249 Commons repository as "Soil network simulation" (Davis and Polhill, 2020).

### 250 **2.4.1 Overview section of Overview, Design Concepts, and Details (ODD)**

- 251 I. Model purpose
- 252 a. The model is designed to be an analytical tool to explore the heterogeneity in  
253 resource supply potential of a network by populating it with idealised energy-  
254 consuming agents, and to quantify the effects of consumer, resource, and network  
255 characteristics on resulting consumer population outcomes.
- 256 II. Entities, state variables, and scales
- 257 a. Consumer entities

258 i. State variables

Property	Description
Location	The resource on which the consumer is located
Target location	The resource to which the consumer will move next
Active?	Whether a consumer is active (or dead)

259 ii. Parameters

Property	Description
Basal metabolism	How much resource an agent needs per day to stay alive
Active metabolism	How much resource an agent uses with each step
Resource stock	How much resource an agent has consumed but not metabolised
Consumption rate	Maximum number of resource units that an agent takes from a resource it visits, per timestep
Spawn energy	How much energy an agent requires to spawn (depletes this quantity from stocks and passed to offspring as starting quota)

260

261 b. Resource entities

262 i. State variables

Property	Description
Current supply	The current quantity of resource at this point

263

264 ii. Parameters

Property	Description
Resource capacity	How much energy is stored in a resource when it is full
Regrow rate	The amount the resource regrows each timestep

265

266 c. Link entities

267 i. State variables

Property	Description
Length	The length of the link - determines energy and time required to traverse it

268

269 d. Scales

Property	Description
Timestep	A single unit of time in the model, defined as that which is required for consumers to move 1 pixel (approximately 0.3 – 0.5 mm), and for which they require <code>basal-metabolism</code> units of energy.
World size	400 x 500, determined by the size of the soil networks used as the environment.

270

271 III. Sequence of events

- 272 a. Consumers start on random nodes around a pre-specified network, where nodes are  
 273 resource patches.  
 274 b. Consumers move around the network randomly following links. If they find a  
 275 resource patch, they consume as much as they can from it, and the patch depletes.  
 276 i. Consumers require `basal-metabolism` units of resource per timestep. If  
 277 they do not consume this resource, they die.  
 278 ii. Consumers can stay put on a resource and consume it (`consumption-`  
 279 `rate` units consumed per timestep), but it depletes, and if there is no more  
 280 resource there then they move on.  
 281 iii. Consumers metabolise `active-metabolism` units of resource per patch  
 282 of link that they cross.  
 283 iv. If there is more than one agent on a resource patch, they each take  
 284 `consumption-rate` units per timestep, or split the remainder if there is  
 285 not enough resource remaining for them to each get `consumption-rate`  
 286 units.  
 287 c. If consumers have twice as much energy as the set `spawn-energy`, they can  
 288 spawn new consumers (who take the same amount of `resource-stock` from  
 289 their parent that the parent started with, so now parent and offspring both have the  
 290 same `resource-stock`).  
 291 d. Resources regrow at a constant rate (`regrow-rate`) per timestep, up to their  
 292 maximum capacity (`resource-capacity`).

#### 293 2.4.2 Sensitivity analysis

294 To determine the sensitivity of the ABM to input parameters, and the robustness of any emergent  
 295 patterns of heterogeneity, we performed an extensive sensitivity analysis following  
 296 recommendations in the agent-based modelling literature. This is detailed in Appendix 3. Table 2  
 297 shows the final parameter values used for the consumer populations, resource populations, and  
 298 general model. In the actual simulation runs, each combination of the five consumer parameter sets,  
 299 and three resource parameter sets, was tested against each network architecture, resulting in 8700  
 300 total runs including replicates.

301 **Table 2. Final values for (a) consumer, (b) resource, and (c) general simulation parameters.**

#### 302 a. Consumer parameters

Parameter	Consumer type				
	High metabolism, high consumption, high spawn energy (HHH)	Low metabolism, low consumption, low spawn energy (LLL)	Low metabolism, moderate consumption, low spawn energy (LML)	Low metabolism, moderate consumption, moderate spawn energy (LMM)	Moderate metabolism, moderate consumption, moderate spawn energy (MMM)
Basal metabolism	3	1	1	1	2
Active metabolism	3	1	1	1	2
Consumption rate	10	5	7	7	7
Spawn energy	100	50	50	75	75
Initial resource stock	30	30	30	30	30

303

#### 304 b. Resource parameters

Parameter	Resource type		
	High capacity, low growth (HL)	Moderate capacity, moderate growth (MM)	Low capacity, high growth (LH)
Maximum resource capacity	50	35	20
Maximum regrow rate	10	15	20

305

306 c. General parameters

Parameter	Value
Initial population size	500 consumers
Length of simulation	2000 timesteps

307

### 308 2.4.3 Analytical method

309 At each time step, the ABM calculated five metrics (Table 3), including measures of centre and  
310 spread of consumer resource stocks, the final population size, and two additional inequality metrics:  
311 the Gini coefficient and a modified form of the Shannon entropy. The latter estimates the  
312 differential entropy of a continuous variable, by discretising the distribution into bins (Appendix 4).  
313 These metrics were chosen to include measures of absolute and relative inequality, and a measure  
314 of evenness common to ecology. As the distributions of each metric across the soil types did not  
315 meet assumptions of most parametric tests, mixed-effects ART ANOVAs with profile ID as a random  
316 effect were again used to quantify how the outcome metrics differed, for each combination of  
317 resource and consumer population parameters and soil type. As the final population size and the  
318 entropy of consumer resource stocks both showed variance not fully explainable by consumer or  
319 resource population parameters, these were also tested with Kruskal-Wallis tests comparing them  
320 across profile IDs and soil types. The significantly different pairs of profiles were identified with Dunn  
321 post-hoc analysis. All data processing, analysis, and visualisation was done in R, using the packages  
322 listed previously, as well as the entropy v1.2.1 (Hausser and Strimmer, 2014) and ineq v0.2.13  
323 packages (Zeileis, 2014).

324

325 **Table 3. The name and description of outcome variables calculated for the agent-based model**  
 326 **(ABM).**

Variable name	Description
Mean consumer resource stock	The mean of the resource stocks held by all active consumers. Units are the same as those of the quantity measured.
Standard deviation consumer resource stock	The square root of the sum of squared absolute differences between each observation and the mean, normalised by the number of observations (minus one, to allow for sample estimation). Units are the same as those of the quantity measured. $s = \sqrt{\frac{\sum  x_i - \bar{x} ^2}{n - 1}}$
Gini coefficient consumer resource stock	Measures the deviation of a population from perfect equality. Mathematically, it can be calculated as half the relative mean absolute difference, or half the average absolute difference between all pairs of the population, divided by the average of the population to normalise. Unitless.
Entropy consumer resource stock (Shannon index)	Measures the amount of information that would be needed to represent the state of the system. Specifically, it is the negative sum of the probability of a consumer's resource stock occurring within a given range, and the log of that probability, normalised by the maximum value ( $\log n$ ). This is the discretised formula for entropy. The units depend on the base of the log: here we use base 2 (units: bits). $H(X) = \frac{-\sum f(x_i) \log_2 \frac{f(x_i)}{w(x_i)}}{H_{max}}$
Final population size	Count of currently active ('alive') consumers.

## 328 3. Results

### 329 3.1 Network metrics

330 The network metrics showed several significant differences between the Cambisols and Arenosols,  
331 with the Cambisols having higher values for most metrics measuring size and structure. These are  
332 summarised in Table 4 and Fig 1.

333 **Table 4. (a) Estimated marginal means, standard errors, and outcomes for mixed-effect nested**  
334 **ANOVAs comparing network metrics between Cambisol and Arenosol main soil networks, and (b)**  
335 **medians and 95 % confidence intervals and results of mixed-effect nested Aligned-Ranks**  
336 **Transformation (ART) ANOVAs comparing Cambisol and Arenosol subnetworks.** Shown in (a) are  
337 the Type II Wald Chi-square statistic and p-values for models comparing each network metric across  
338 soil types. Profile ID was included as a mixed effect; its log-likelihood ratio test (LRT) statistic and p-  
339 value are also shown. Both the Chi-square and LRT used one degree of freedom to compare soil  
340 types. Estimated marginal means and standard errors were calculated from ANOVAs. In (b) ART  
341 ANOVAs were used as the data were non-normal; profile ID was also included as a mixed effect.  
342 Shown are Type III Wald F tests with Kenward-Roger degrees of freedom. The asterisks designate  
343 level of significance: p < 0.1: ·, p < 0.05: \*, p < 0.01: \*\*, p < 0.001: \*\*\*. Descriptions of the metrics are  
344 in Table 1.

345 a. Main networks

346

	Arenosols (n = 25)		Cambisols (n = 25)		Soil type		Profile ID (random effect)	
	Est. marginal mean	SE	Est. marginal mean	SE	$\chi^2$	<i>p</i>	LRT	<i>p</i>
<b>No. of nodes</b>	2670.000	9.570	3326.000	10.070	7.809	0.005 **	21.384	< 0.001 ***
<b>No. of links</b>	4225.000	344.000	5581.000	293.000	9.058	0.003 **	19.280	< 0.001 ***
<b>Mean node degree</b>	3.140	0.055	3.350	0.047	9.098	0.003 **	9.005	0.003 **
<b>Mean link length</b>	3.570	0.023	3.700	0.022	17.667	0.000 ***	0.414	0.520
<b>SD link length</b>	2.110	0.025	2.210	0.025	9.815	0.002 **	0.000	1.000
<b>Gamma index</b>	0.001	0.000	0.001	0.000	4.838	0.028 *	20.231	< 0.001 ***
<b>Beta index</b>	1.570	0.028	1.680	0.024	9.098	0.003 **	9.005	0.003 **
<b>Diameter</b>	154.000	11.100	201.000	10.700	10.017	0.002 **	0.141	0.708
<b>Cost</b>	15180.000	1257.000	20600.000	1072.000	10.840	0.001 ***	16.153	< 0.001 ***
<b>Global reach centrality</b>	0.002	0.000	0.002	0.000	0.049	0.825	10.923	< 0.001 ***
<b>Convex hull area</b>	166940.000	2949.000	167086.000	2514.000	0.001	0.970	11.730	< 0.001 ***
<b>Network density</b>	0.016	0.001	0.020	0.001	8.037	0.005 **	16.547	< 0.001 ***
<b>No. of subnetworks</b>	163.000	3.382	158.000	3.236	0.009	0.924	6.302	0.012 *

347

348

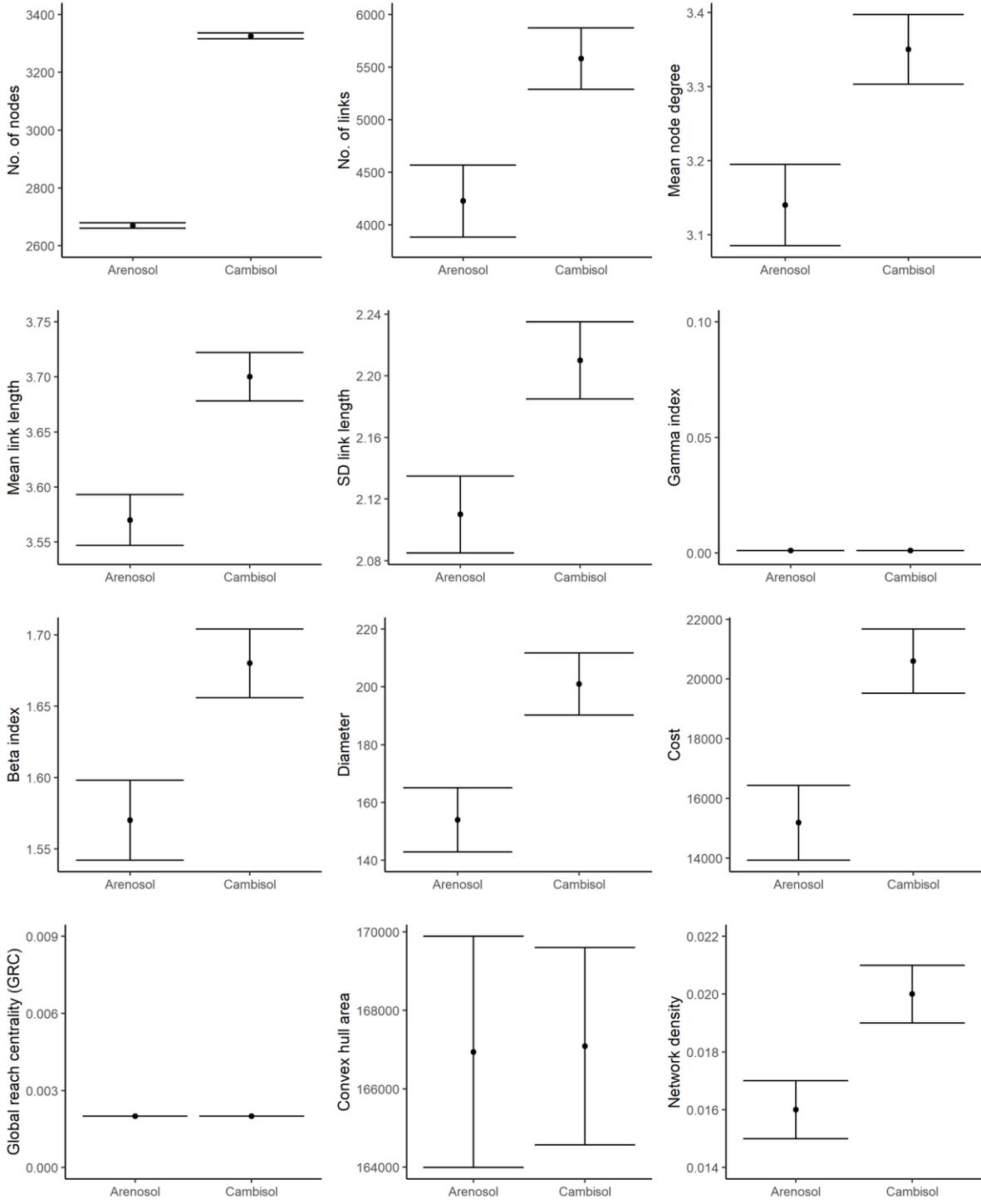
349

## b. Subnetworks

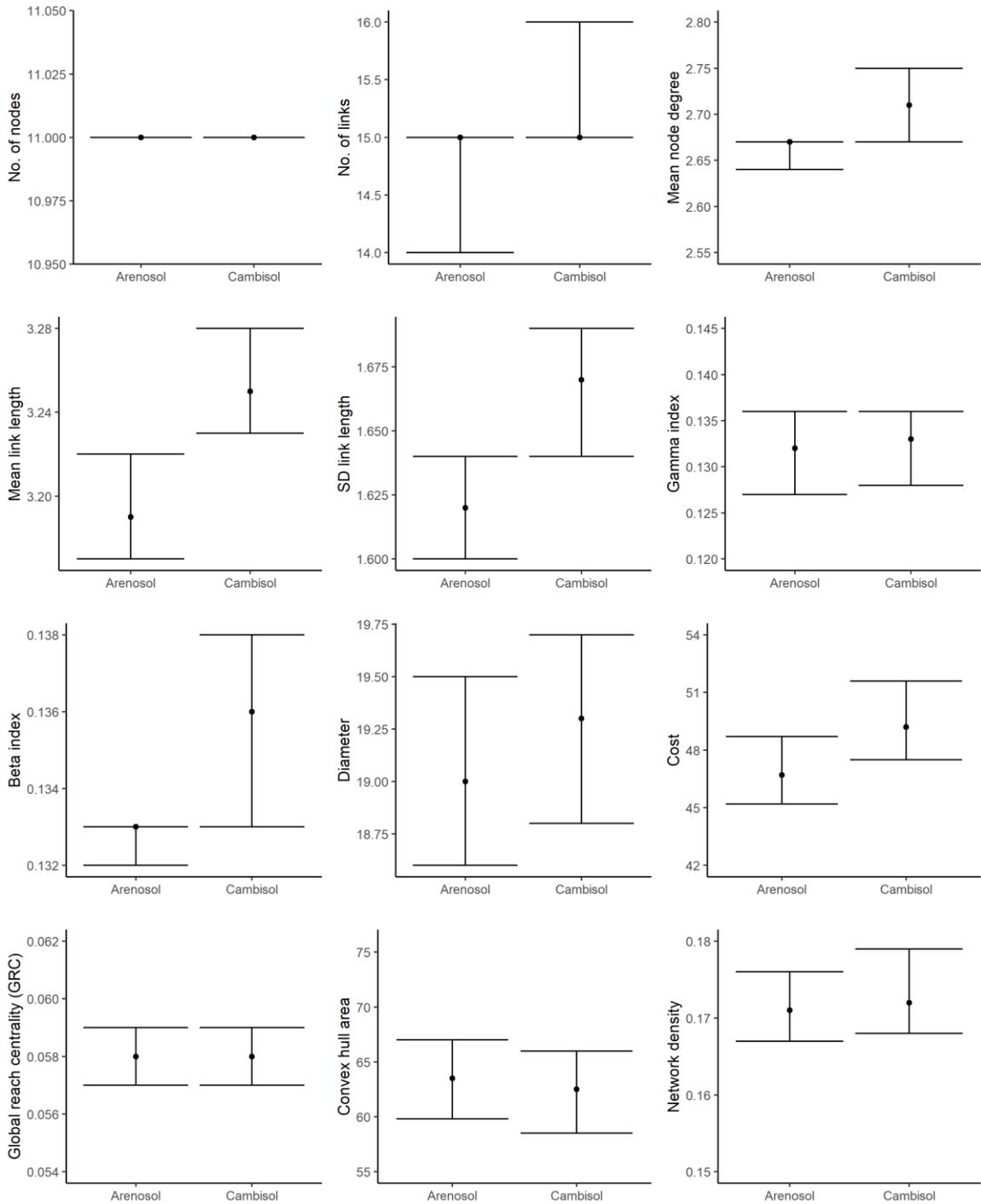
350

	Arenosols (n = 3906)			Cambisols (n = 3834)			ANOVA	
	Median	Lower CI	Upper CI	Median	Lower CI	Upper CI	<i>F</i>	<i>p</i>
<b>Number of nodes</b>	11.000	11.000	11.000	11.000	11.000	11.000	F(1, 8.614) = 1.542	0.247
<b>Number of links</b>	15.000	14.000	15.000	15.000	15.000	16.000	F(1, 8.443) = 3.793	0.085
<b>Mean node degree</b>	2.670	2.640	2.670	2.710	2.670	2.750	F(1, 7.971) = 9.239	0.016 *
<b>Mean link length</b>	3.190	3.170	3.220	3.250	3.230	3.280	F(1, 7.011) = 11.322	0.012 *
<b>SD link length</b>	1.620	1.600	1.640	1.670	1.640	1.690	F(1, 8.334) = 3.648	0.091
<b>Gamma index</b>	0.132	0.127	0.136	0.133	0.128	0.136	F(1, 8.720) = 0.355	0.567
<b>Beta index</b>	0.133	0.132	0.133	0.136	0.133	0.138	F(1, 7.971) = 9.239	0.016 *
<b>Diameter</b>	19.000	18.600	19.500	19.300	18.800	19.700	F(1, 7.957) = 1.664	0.233
<b>Cost</b>	46.700	45.200	48.700	49.200	47.500	51.600	F(1, 8.161) = 4.717	0.061
<b>Global reach centrality</b>	0.058	0.057	0.059	0.058	0.057	0.059	F(1, 7.645) = 0.554	0.479
<b>Convex hull area</b>	63.500	59.800	67.000	62.500	58.500	66.000	F(1, 8.132) = 0.385	0.552
<b>Network density</b>	0.171	0.167	0.176	0.172	0.168	0.179	F(1, 8.117) = 0.013	0.913

351



352 a.  
353



354 **b.**

355 **Figure 1. The distribution of each network metric by soil type, for (a) main soil networks and (b)**  
 356 **subnetworks.** The point and error bars in (a) represent the estimated marginal mean and standard  
 357 error for that network type and soil type, as determined by the ANOVAs (Table 4a), and the point  
 358 and error bars in (b) represent the median and upper and lower 95 % confidence intervals,  
 359 respectively. Descriptions of the network metrics are in Table 1.

360

361

362 At the main network level, the networks extracted from the Cambisols had significantly more nodes  
 363 and links, a larger mean node degree and standard deviation of link length, and longer mean link  
 364 length (Table 4a). These networks also had a higher beta index, higher cost, and higher density. While  
 365 the main networks of the two soil types had significantly different gamma indexes, the absolute  
 366 difference in the estimated marginal means between the two soil types was negligible ( $< 10^{-3}$ ) (Table  
 367 4a, Fig 1a). At the subnetwork level, Cambisol networks had longer mean link length, and higher  
 368 mean node degree and beta index (Table 4b). While not significant, Cambisol subnetworks also had  
 369 noticeably larger number of links and standard deviation of link length, and higher cost (Fig 1b).

370 To control for the effect of replication on the significance, the profile ID was included in the ANOVAs  
 371 as a mixed effect. This was significant for all metrics except mean and standard deviation of link  
 372 length and diameter. Most profiles within each soil type at the main network level showed low  
 373 absolute variation across the networks extracted from each however, and noticeably higher metric  
 374 values for Cambisols than Arenosols (Fig A7a). At the subnetwork level, the distributions were quite  
 375 similar across all profiles, but the Cambisol profiles showed more frequent and higher outliers.

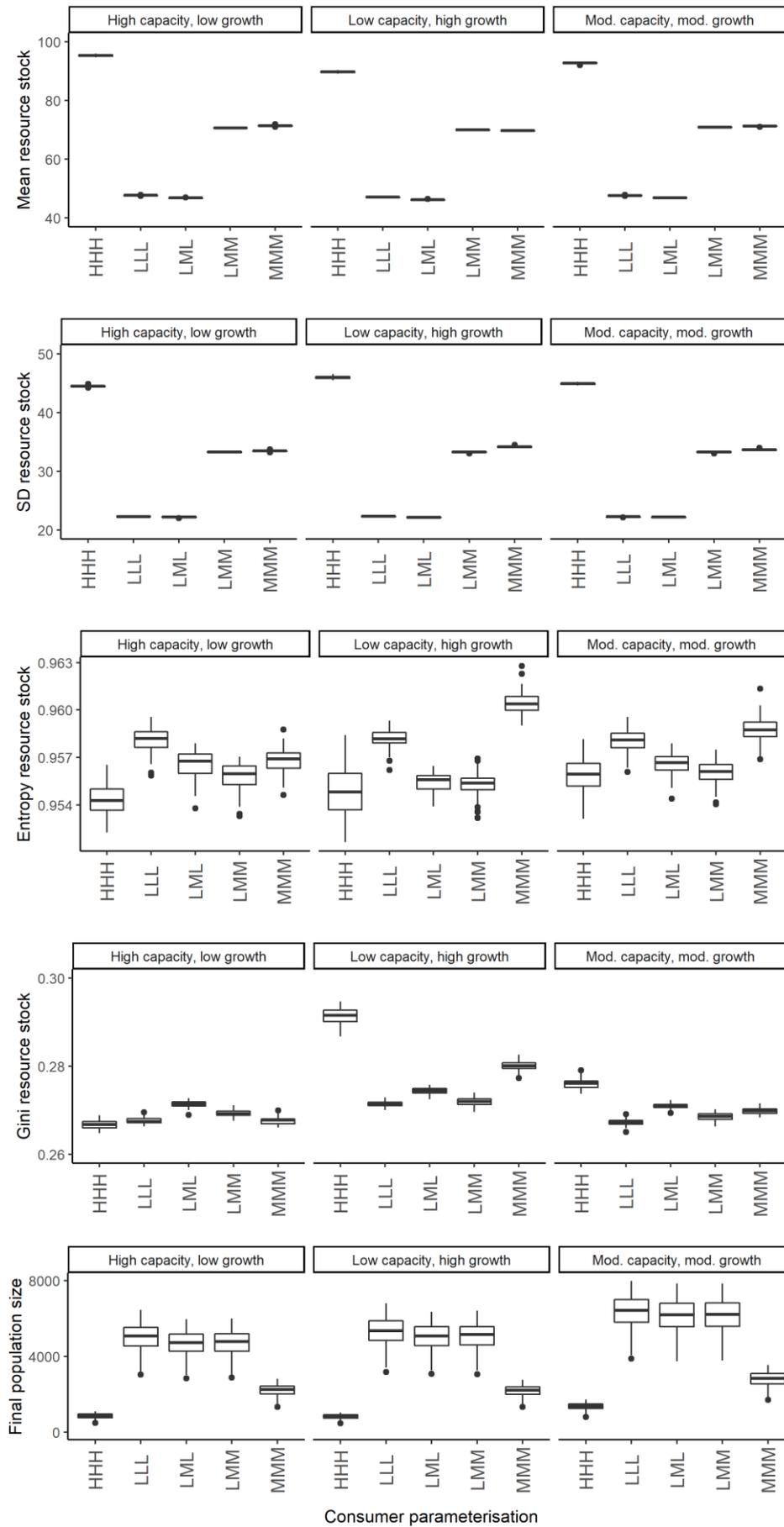
### 376 3.2 Agent-based model

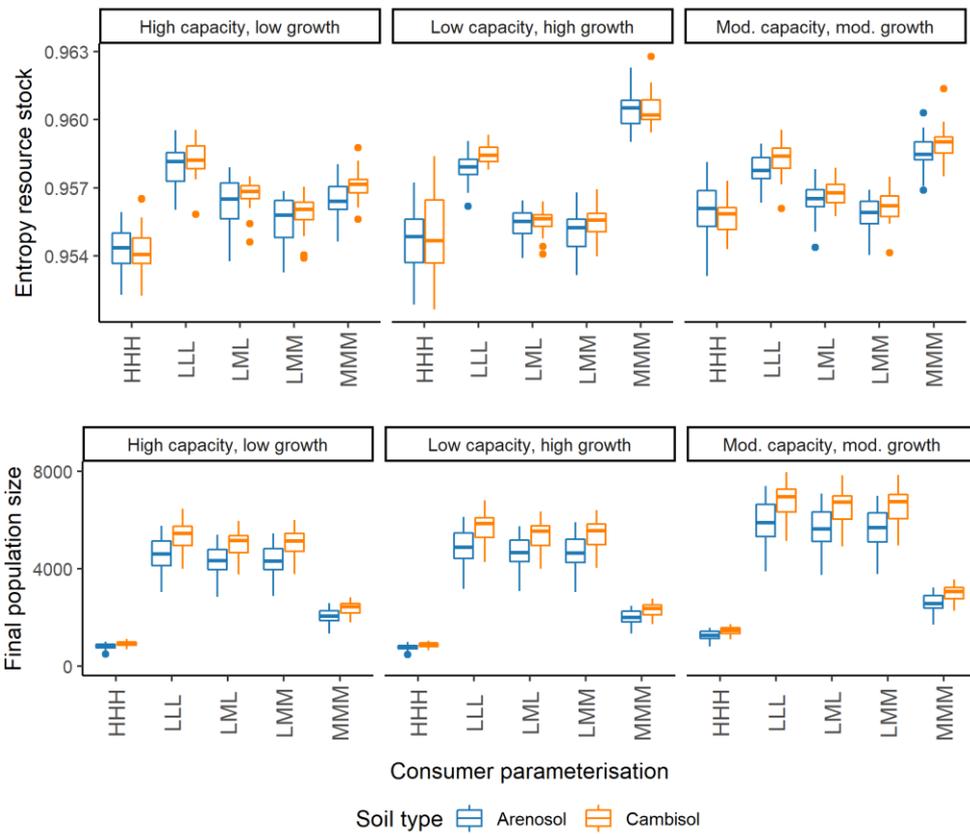
377 The ABM results showed significant differences across the different combinations of  
 378 parameterisations and soil types, summarised in Tables 5 and 6 and Fig 2. The simulations run on the  
 379 Cambisol networks had significantly higher final population sizes (Tables 5 and 6, Fig 2b), and  
 380 interactions between soil type and consumer and resource parameterisation were significant for  
 381 several outcome variables (Table 6).

382

383 **Table 5. The medians, first and third quantiles for agent-based model (ABM) outcome variable**  
 384 **values across the two soil types.** These values represent the overall results across all consumer and  
 385 resource parameterisations. Descriptions of the variables are in Table 3.

	Arenosols (n = 375)			Cambisols (n = 375)		
	Median	1 <sup>st</sup> Quantile	3 <sup>rd</sup> Quantile	Median	1 <sup>st</sup> Quantile	3 <sup>rd</sup> Quantile
<b>Mean resource stock</b>	69.970	47.086	71.391	69.940	47.077	71.397
<b>SD resource stock</b>	33.266	22.262	34.123	33.300	22.276	34.112
<b>Entropy resource stock</b>	0.956	0.955	0.958	0.957	0.956	0.958
<b>Gini resource stock</b>	0.271	0.268	0.273	0.271	0.268	0.274
<b>Final population size</b>	4049.728	1967.921	5102.043	4890.158	2279.1560	5842.834





387

**b.**

Soil type  Arenosol  Cambisol

388 **Figure 2. Distributions of (a) each agent-based model (ABM) outcome variable, grouped by**  
 389 **resource parameterisation (columns, labelled at top) and consumer parameterisation (x axis**  
 390 **within columns), across both soil types, and (b) ABM outcome variables that were significantly**  
 391 **affected by soil type (represented by colour), grouped by resource parameterisation (columns)**  
 392 **and consumer parameterisation (x axis within columns). The three-letter consumer**  
 393 **parameterisation codes refer to the metabolism, consumption rate, and spawning threshold,**  
 394 **respectively, where H is high, M is medium, and L is low. Descriptions of the resource and consumer**  
 395 **parameterisations are in Table 2, and descriptions of the outcome variables are in Table 3.**

396

397 The ART ANOVAs showed that measured outcomes all differed significantly across consumer  
 398 parameterisation, resource parameterisation, and consumer-resource parameterisation interactions.  
 399 Final population size differed significantly by soil type, soil type-resource parameterisation  
 400 interaction, and soil type-consumer parameterisation interaction. Mean resource stock also differed  
 401 significantly by soil type-resource parameterisation interaction.

402

403 **Table 6. Overview of Aligned Ranks Transformation ANOVA models of consumer population**  
 404 **outcomes by consumer and resource parameterisation and soil type.** The tests were Type III Wald F  
 405 tests with Kenward-Roger degrees of freedom. Profile ID was included as a random effect. The  
 406 asterisks designate level of significance:  $p < 0.1$ : ·,  $p < 0.05$ : \*,  $p < 0.01$ : \*\*,  $p < 0.001$ : \*\*\*.  
 407 Descriptions of consumer and resource parameterisations are in Table 2 and descriptions of  
 408 response variables are in Table 3.

Response variable	Predictors	F	Significance
Mean consumer resource stock	Consumer population	F(4, 710.063) = 3585.924	< 0.001 ***
	Resource population	F(2, 710.063) = 2585.400	< 0.001 ***
	Soil type	F(1, 9.692) = 0.328	0.560
	Consumer pop. x resource pop.	F(8, 710.094) = 998.152	< 0.001 ***
	Consumer pop. x soil type	F(4, 710.106) = 1.460	0.213
	Resource pop. x soil type	F(2, 710.103) = 3.513	0.030 *
	Consumer pop. x resource pop. x soil type	F(8, 710.102) = 1.020	0.419
SD consumer resource stock	Consumer population	F(4, 710.185) = 3629.337	< 0.001 ***
	Resource population	F(2, 710.292) = 1137.215	< 0.001 ***
	Soil type	F(1, 9.212) = 0.555	0.475
	Consumer pop. x resource pop.	F(8, 710.233) = 677.837	< 0.001 ***
	Consumer pop. x soil type	F(4, 710.315) = 2.164	0.071 ·
	Resource pop. x soil type	F(2, 710.32) = 0.538	0.584
	Consumer pop. x resource pop. x soil type	F(8, 710.306) = 1.123	0.345
Entropy consumer resource stock	Consumer population	F(4, 710.030) = 586.700	< 0.001 ***
	Resource population	F(2, 710.036) = 59.661	< 0.001 ***
	Soil type	F(1, 9.861) = 3.105	0.109
	Consumer pop. x resource pop.	F(8, 710.025) = 67.989	< 0.001 ***
	Consumer pop. x soil type	F(4, 710.037) = 2.364	0.052 ·
	Resource pop. x soil type	F(2, 710.037) = 0.241	0.786
	Consumer pop. x resource pop. x soil type	F(8, 710.037) = 0.949	0.475
Gini consumer resource stock	Consumer population	F(4, 710.677) = 1296.640	< 0.001 ***
	Resource population	F(2, 711.086) = 2004.095	< 0.001 ***
	Soil type	F(1, 7.791) = 2.445	0.158
	Consumer pop. x resource pop.	F(8, 710.847) = 1005.251	< 0.001 ***
	Consumer pop. x soil type	F(4, 711.364) = 0.470	0.758
	Resource pop. x soil type	F(2, 711.281) = 2.502	0.083 ·
	Consumer pop. x resource pop. x soil type	F(8, 711.287) = 0.614	0.766
Final population size	Consumer population	F(4, 710.001) = 1361.66	< 0.001 ***
	Resource population	F(2, 710.001) = 604.376	< 0.001 ***
	Soil type	F(1, 9.998) = 9.239	0.012 *
	Consumer pop. x resource pop.	F(8, 710.001) = 33.651	< 0.001 ***
	Consumer pop. x soil type	F(4, 710.001) = 41.516	< 0.001 ***
	Resource pop. x soil type	F(2, 710.001) = 5.039	0.007 **
	Consumer pop. x resource pop. x soil type	F(8, 710.001) = 0.282	0.972

410 The entropy of consumer resource stocks and the final population size both showed considerable  
 411 variation in the initial boxplots that was not explained by the consumer and resource  
 412 parameterisation (Fig 2a), and the ANOVA results suggested that soil type was influential on final  
 413 population size. Therefore, these were further explored with Kruskal-Wallis tests, first with profile ID  
 414 as the grouping variable, then soil type (Table 7, also Fig 2b). Significant differences in profile ID were  
 415 explored with Dunn post-hoc analysis. This showed that entropy differed significantly between  
 416 profiles D and H, which were Cambisol and Arenosol, respectively, while final population size  
 417 differed significantly between several pairs of profiles, including both intra- and inter-type profile  
 418 pairings.

419

420 **Table 7. Results of Kruskal-Wallis tests and Dunn post-hoc analysis comparing entropy of**  
 421 **consumer resource stocks and final population size by soil profile ID and soil type.** The degrees of  
 422 freedom for the Chi-square statistics were 11 and 1 for profile ID and soil type, respectively. Profile  
 423 IDs A – G correspond to Cambisols, while profile IDs H – K correspond to Arenosols. Significant pairs  
 424 of profiles were identified at the level of  $\alpha/2$ , where  $\alpha = 0.05$ . Profile pairings in italics denote inter-  
 425 type pairs.

Response variable	Grouping variable	Significance		Significantly different pairs ( $p < 0.025$ )
		$\chi^2$	$p$	
Entropy consumer resource stock	Profile ID	25.824	$p = 0.007^{**}$	<i>D : H</i>
	Soil type	4.965	$p = 0.026^*$	Cambisol : Arenosol
Final population size	Profile ID	65.167	$p < 0.001^{***}$	<i>A : C, A : H, A : I, A : K, A : L, B : H, B : I, B : K, D : H, D : I, D : K, G : H, G : I, G : K, H : J, I : J, J : K</i>
	Soil type	21.974	$p < 0.001^{***}$	Cambisol : Arenosol

426

427

428

## 429 4. Discussion

### 430 4.1 Network analysis

431 Given the known characteristics of the two soil types, the results of the network analysis suggest  
432 that the methodology developed here captures overall trends of soil structural development.  
433 Cambisols typically have more soil structure, higher porosity, higher levels of biotic activity, and  
434 greater stability than Arenosols (FAO, 2015). Correspondingly, the abstracted Cambisol soil networks  
435 analysed here showed higher values for the metrics measuring size, structure, and connectivity than  
436 the abstracted Arenosol soil networks did.

437 Specifically, the Cambisol soil networks had significantly more nodes and links, longer mean and  
438 standard deviation of link lengths, and higher total cost, density, and diameter (Table 4). This  
439 suggests more pore-creating activities modifying the soil, and a soil structural matrix that can  
440 support longer pores. This would also lead to higher water holding capacity, and increased internal  
441 drainage, both of which are commonly associated with Cambisols (FAO, 2015). In contrast, the  
442 smaller and less connected Arenosol networks have a low water-holding capacity, and the weaker  
443 coherence of their matrix material prevents longer pores from being stable, making them prone to  
444 erosion (FAO, 2015). Cambisols are also classified as more structurally developed than Arenosols,  
445 and contain more organic matter (FAO, 2015), both of which further validate the increased structure  
446 seen in the Cambisol networks here.

447 The global reach centrality, gamma index, and convex hull area were not as clearly differentiated  
448 between the Cambisol and Arenosol soil networks, however. The global reach centrality values were  
449 small and functionally identical, with an estimated marginal mean of 0.001 and 0.058 for both soil  
450 types at the main and subnetwork level, respectively (Table 4). Similarly, the estimated marginal  
451 mean gamma index for main networks of both soil types was 0.001. This is likely due to the presence  
452 of a similar number of disconnected subnetworks within each soil network, limiting the total number  
453 of nodes that any given node can reach. The Cambisol main networks also had a slightly smaller  
454 range of convex hull areas, although the opposite trend emerges at the subnetwork level (Fig 1,  
455 Table 4). When this is decomposed by profile, the Cambisols show more variation and outliers across  
456 and within profiles for several metrics, including convex hull area (Fig A7), suggesting that soil type  
457 includes a greater heterogeneity of network sizes and structures. As with the other metrics, further  
458 work is required to establish ranges across different soil types and geographical regions, and to  
459 compare these metrics with those more commonly used in soil analysis. Overall, however, the  
460 differences between the Cambisols and Arenosols as captured in this analysis broadly reflects those  
461 expected, given the known differences in their properties.

462 The improved profile development and heterogeneity of Cambisols highlights their potential for  
463 agriculture and forestry, and in underpinning the diversity of a range of ecosystems. It is vital to  
464 manage them in a way that preserves and enhances their soil structure, however, to maintain their  
465 porosity and biodiversity, and resulting stability, drainage, and aeration. Similarly, Arenosols should  
466 be managed in a way that minimises their propensity for erosion and soil loss. In both cases, this can  
467 be accomplished through limiting or eliminating tillage (e.g. Young and Ritz, 2000; Helgason, Walley  
468 and Germida, 2010; Kravchenko *et al.*, 2011), and increasing cover crops and native species (e.g.  
469 Fernández *et al.*, 2019; Kravchenko *et al.*, 2011). These provide additional organic inputs to the soil  
470 to promote an active and diverse soil biota, and therefore the positive feedback between biota, and  
471 structural development and stability (e.g. Oades, 1993; Young and Ritz, 2009; Crawford *et al.*, 2012).  
472 The feasibility of the measurement and analysis methods presented here could provide a basis for  
473 estimating changes in structure over time and under different management strategies or

474 environmental changes. This would help inform actions taken to preserve or improve the soil  
475 structure. However, further work is required to standardise the approach and demonstrate its  
476 application over multiple soil types.

477 As introduced in the Methods, the networks analysed here represent abstractions of the true soil  
478 structure present in the samples. This simplification is reasonable for analysing overall structural  
479 characteristics and heterogeneity and made the computation of the network metrics feasible.  
480 Although the short link lengths (Table 4) suggest that using Euclidean distance is likely negligibly  
481 different than measuring the path through the pixels, it does limit the interpretation of the findings  
482 we present. Specifically, the absolute values of the metrics cannot be taken to characterise the  
483 precise soil structure, but rather suggest general trends in structural development. As the exact size  
484 and shape of the pores was not preserved, many of the finer distinctions between networks may  
485 also be lost. This could cause the magnitudes of differences found between soil samples here to  
486 appear lower than they are. As discussed above, the relatively rapid, low-cost, and lightweight  
487 approach used here for estimating soil structure should be compared against more established  
488 approaches and metrics to determine its effectiveness. This methodology provides simplified and  
489 potentially inaccurate measurements of soil structure, but with further improvement it could be a  
490 suitable approach for rapid assessment of soil structure in the field. The results presented suggest  
491 that the methodology can still capture general known trends of heterogeneity within soil networks,  
492 meriting further refinements and application.

#### 493 4.2 ABM analysis

494 The ABM evaluated the effects of and interactions between consumer and resource characteristics,  
495 and the structure of the abstracted soil networks, on the measured consumer outcomes. Overall,  
496 the results showed that the size and energetic heterogeneity of the consumer population was  
497 heavily influenced by the parameterisation of the consumer population and resource base, and their  
498 interactions. Moreover, while outcome variables were less directly affected by soil network  
499 structure, they were more influenced by the interactions between this network structure and  
500 consumer or resource parameterisations.

501 Across all simulations, measured outcomes varied most strongly across consumer and resource  
502 characteristics, and their combinations as overall consumer and resource parameterisations or types  
503 (Fig 2a, Table 6). Specifically, the mean, standard deviation, and entropy of consumer resource  
504 stocks, as well as the final population size, were most different across consumer types. These  
505 differences in outcome variables resulted from how each consumer population responded to the  
506 provided resource base. For example, the consumer populations with low metabolisms, low  
507 consumption rate, and a low energy requirement for spawning had a lower mean resource stock,  
508 and a higher final population size, for any given resource base. The consumers with high  
509 metabolisms, high consumption rate, and a high energy requirement for spawning had a lower final  
510 population size, but higher mean resource stock. This is similar to the distinction between r-  
511 strategists and K-strategists. In these simulations, the threshold for spawning and the active and  
512 basal metabolic rates appeared to have the largest impact on the measured outcome variables (Fig  
513 2a). This is likely due to these parameters balancing one another in determining energy allocation  
514 between maintenance and reproduction (e.g. Brown *et al.*, 2004; Kooijman, 2009).

515 In addition to consumer and resource characteristics, the soil type, and therefore soil network  
516 structure, also affected population size and diversity (Table 6). Specifically, the mean consumer  
517 resource stock and final population size showed significant differences across resource and soil type  
518 interactions, and final population size also showed significant differences between soil types (Table

519 5). While the final population size and entropy also differed significantly across profiles (Table 6),  
520 post-hoc analysis revealed that for entropy this was only significant for inter-type profile pairings,  
521 and a slight difference was visible between groups when plotted (Fig 2b). This entropy is also known  
522 as the Shannon Index or Shannon-Wiener Index, and here measures the diversity or 'evenness' of  
523 the distribution of consumer resource stocks (Hill, 1973; Spellerberg and Fedor, 2003). Higher  
524 entropy therefore meant that given quantities of resource stock were represented in equal  
525 proportional abundance across the population. This is typically caused by groups of consumers  
526 emerging, where group members each have the same quantity of resource stock, but these  
527 quantities differ among groups. Over time, adaptations in this context could drive the system toward  
528 speciation. In these simulations, the larger populations supported by the larger Cambisol soil  
529 networks were more likely to have higher entropy, through different quantities of consumer  
530 resource stocks represented with equal proportional abundance.

531 The relatively low Gini coefficients (Table 5, Fig 2a) can also suggest the emergence of distinct  
532 groups of consumers with equal resource stocks, with similar numbers of consumers across the  
533 groups. As the Gini coefficient measures relative inequality, both inequality in resource stocks across  
534 groups, and more groups, cause it to increase. Equal group sizes can somewhat counter this. In both  
535 soil types however, as the consumers in a given simulation had identical characteristics, it is  
536 reasonable that they would have similar outcomes, slightly differing based on the subnetwork in  
537 which they found themselves, and the resource base available to them there. The similarity among  
538 subnetworks of the two soil types (Table 4b) suggests that the heterogeneity between soil types is  
539 more apparent at the main network level. As the consumers in these simulations were unable to  
540 move between subnetworks, they likely did not experience the full range of environmental  
541 heterogeneity between the soil types, which would have limited its effect on the measured  
542 outcomes.

543 Overall, the simulations highlight the differences in population size and diversity across consumer  
544 and resource parameterisations and interactions, soil and resource type interactions, and to a lesser  
545 extent, soil type on its own. Spatial heterogeneity, through both resource and network structural  
546 heterogeneity, can increase the microhabitat diversity (Anderson, 1978; Giller, 1996; Ettema and  
547 Wardle, 2002; Nielson *et al.*, 2010), which was shown here through the increased evenness of  
548 consumer groups with different resource stocks. Similarly, the heterogeneous habitat of soils can  
549 limit competitive exclusion by providing structural and resource niches for different species  
550 (Bardgett, Yeates and Anderson, 2009), such that more structurally heterogeneous Cambisols have  
551 larger and more diverse populations (FAO, 2015). This was reproduced by the larger populations that  
552 emerged in the Cambisol simulations here, although speciation was not explicitly modelled. As with  
553 the findings of the network analysis, this emphasises the importance of preserving soil structure and  
554 providing adequate substrate for maintaining an active soil biota (e.g. Young and Ritz, 2009;  
555 Crawford *et al.*, 2012; Fernández *et al.*, 2019).

556 While the parameterisations presented here were limited, they revealed interesting effects of  
557 consumer and resource characteristics and interactions. The programming of the model itself,  
558 however, may also have had an impact on the outcome of consumer populations. For example,  
559 consumers moved randomly among resources rather than following any sort of search strategy, and  
560 there was no energetic penalty imposed for turning, which are simplifying assumptions based on the  
561 limited sensory and processing capabilities of most soil biota. This eliminated free parameters that  
562 would have to be tuned and analysed or sourced from limited data about specific soil biota  
563 metabolism and cognition. It also eliminated any effect that tortuosity of the network would have on  
564 consumer resource stocks, though. This may not be a correct assumption if turning has a higher

565 burden physically, cognitively, or both. Furthermore, as consumers were not able to extend the  
566 network or move between subnetworks, they were unlikely to experience the full difference  
567 between soil networks, as discussed above. This may have led to a smaller effect of soil type on  
568 measured consumer outcomes.

569 Additionally, the extraction and simplification process used to create the soil networks may have  
570 affected the outcomes of the ABM. As the details of pore size and shape were not maintained, the  
571 consumers' ability to forage or hide in crevices was not intended to mimic the true range of  
572 consumer sizes and behaviours. Since predation was not included in the model, however, we did not  
573 intend to explore the hypothesised effect of physical niches on populations by limiting competitive  
574 exclusion and predation. While this would be an interesting future extension, and these changes  
575 could increase the observed effect of the soil network structure on consumer population outcomes,  
576 it would require refining the network extraction process as discussed above, as well as estimating  
577 ranges of consumer sizes and predation dynamics. The model presented here instead focused on  
578 exploring the overall trends that might emerge in a population of consumers, rather than attempting  
579 to predict how specific populations might evolve. While its design limits the precision of the  
580 implications, it maintains the level of realism and generality assumed within the overall  
581 methodology (Levins, 1966).

582

## 583 5. Conclusion

584 This work has explored how analysing abstracted soil networks using standard network metrics,  
585 combined with simulations, can quantify the underlying structural and functional differences  
586 between soil types. We showed that networks derived from a brown forest soil, or Cambisol, were  
587 significantly larger, more connected, and more spatially heterogeneous than the networks derived  
588 from a less developed sandy beach soil, or Arenosol. These larger and more structured networks  
589 were in turn able to support larger populations of simulated consumers in an agent-based model  
590 (ABM). The ABM also demonstrated how the size and heterogeneity of the simulated population  
591 were significantly different across consumer and resource parameterisations, and interactions  
592 between these parameterisations and soil type.

593 In conclusion, standard network metrics applied to images can be a useful way to quickly assess the  
594 structure of networks within a soil profile, by capturing the broad structural differences between  
595 distinct soil types, in a way that can suggest functional differences as well. These initial estimates can  
596 be used on their own to survey an area more extensively or affordably, or coupled with more  
597 intensive analyses, such as three-dimensional imaging techniques. Agent-based modelling can also  
598 be used, when seeded with networks obtained from images or scans, to evaluate interactions  
599 between consumer and resource characteristics and network structure, and to quantify the impact  
600 these and other environmental factors have on the outcomes of simulated populations. Overall,  
601 combining network analysis and simulation modelling can provide unique insights on the structure,  
602 function, and diversity of an area of soil, and provide avenues for exploring the impact of future  
603 management, structural, or environmental changes.

604

605

606 **Acknowledgements**

607 This work was supported by a joint PhD studentship sponsored by Lancaster University and the  
608 James Hutton Institute, awarded to ND, and by the Rural and Environment Science and Analytical  
609 Services (RESAS) Work Package 1.1 Soils. The authors gratefully acknowledge statistical advice from  
610 Dr Vicki Davis, and constructive comments from Dr Jonathan Ball, Dr Uta Berger, and an anonymous  
611 reviewer.

612

613 **Competing interests**

614 The authors declare no competing interests.

615

## 616 6. References

- 617 Aitkenhead, M. J. *et al.* (1999) 'Modelling water release and absorption in soils using cellular  
618 automata', *Journal of Hydrology* 220(1-2), pp. 104-112. doi: 10.1016/S0022-1694(99)00067-0.
- 619 Aitkenhead, M. *et al.* (2016) 'Automated soil physical parameter assessment using smartphone and  
620 digital camera imagery', *Journal of Imaging*, 2(4). doi: 10.3390/jimaging2040035.
- 621 Anderson, J. M. (1978) 'Inter- and intra-habitat relationships between woodland cryptostigmata  
622 species diversity and the diversity of soil and litter microhabitats', *Oecologia*, 32, pp. 341-348. doi:  
623 <https://doi.org/10.1007/BF00345112>.
- 624 Baer, S. G. *et al.* (2005) 'Soil Heterogeneity Effects on Tallgrass Prairie Community Heterogeneity: An  
625 Application of Ecological Theory to Restoration Ecology', *Restoration Ecology*, 13(2), pp. 413-424.  
626 doi: 10.1111/j.1526-100x.2005.00051.x.
- 627 Barabási, A.-L. (2016) *Network Science*. Cambridge, UK: Cambridge University Press.
- 628 Bardgett, R. D., Yeates, G. W. and Anderson, J. M. (2009) 'Patterns and determinants of soil  
629 biological diversity', in *Biological Diversity and Function in Soils*, pp. 100-119.
- 630 Bivand, R. S., Pebesma, E. and Gomez-Rubio, V. (2013) *Applied spatial data analysis with R*. 2nd edn.  
631 New York, New York, USA: Springer. Available at: <http://www.asdar-book.org>.
- 632 ten Broeke, G., van Voorn, G. and Ligtenberg, A. (2016) 'Which Sensitivity Analysis Method Should I  
633 Use for My Agent-Based Model?', *Journal of Artificial Societies and Social Simulation*, 19(1), pp. 1-  
634 35. doi: 10.18564/jasss.2857.
- 635 Brown, J. H. *et al.* (2004) 'Toward a metabolic theory of ecology', *Ecology*, 85(7), pp. 1771-1789. doi:  
636 10.1890/03-9000.
- 637 Crawford, J. W. *et al.* (2012) 'Microbial diversity affects self-organization of the soil - Microbe system  
638 with consequences for function', *Journal of the Royal Society Interface*, 9(71), pp. 1302-1310. doi:  
639 10.1098/rsif.2011.0679.
- 640 Crawford, J. W., Ritz, K. and Young, I. M. (1993) 'Quantification of fungal morphology, gaseous  
641 transport and microbial dynamics in soil: an integrated framework utilising fractal geometry',  
642 *Geoderma*, 56(1-4), pp. 157-172. doi: 10.1016/0016-7061(93)90107-V.
- 643 Csardi, G. and Nepusz, T. (2006) 'The igraph software package for complex network research',  
644 *InterJournal*, Complex Sy, p. 1695.
- 645 Davis, N. *et al.* (2020). 'Trajectories toward maximum power and inequality in resource distribution  
646 networks', *PLoS ONE*, 15(3), e0229956. doi: <https://doi.org/10.1371/journal.pone.0229956>.
- 647 Davis, N. and Polhill, J. G. (2020). 'Soil network simulation (Version 1.0.0)', *Zenodo*.  
648 <http://doi.org/10.5281/zenodo.4001622>
- 649 Davis, N. (2020) 'Soil network analysis functions (Version 1.0.0).' *Zenodo*.  
650 <http://doi.org/10.5281/zenodo.4001702>
- 651 Dinno, A. (2017) 'dunn.test: Dunn's Test of Multiple Comparisons Using Rank Sums.' R package  
652 version 1.3.5. <https://CRAN.R-project.org/package=dunn.test>
- 653 Epskamp, S. *et al.* (2012) 'qgraph: Network Visualizations of Relationships in Psychometric Data',  
654 *Journal of Statistical Software*, 48(4), pp. 1-18.
- 655 Ettema, C. H. and Wardle, D. A. (2002) 'Spatial soil ecology', *Trends in Ecology and Evolution*, 17(4),  
656 pp. 177-183. doi: 10.1016/S0169-5347(02)02496-5.

657 FAO (2015) *World reference base for soil resources 2014: International soil classification system for*  
658 *naming soils and creating legends for soil maps, World Soil Resources Reports No. 106*. Rome, Italy:  
659 Food and Agriculture Organization of the United Nations. doi: 10.1017/S0014479706394902.

660 Fernández, R. *et al.* (2019) 'Pore morphology reveals interaction of biological and physical processes  
661 for structure formation in soils of the semiarid Argentinean Pampa', *Soil and Tillage Research*.  
662 Elsevier, 191(January), pp. 256–265. doi: 10.1016/j.still.2019.04.011.

663 García-Palacios, P. *et al.* (2012) 'Plant responses to soil heterogeneity and global environmental  
664 change', *Journal of Ecology*, 100(6), pp. 1303–1314. doi: 10.1111/j.1365-2745.2012.02014.x.

665 Gargiulo, L., Mele, G. and Terribile, F. (2013) 'Image analysis and soil micromorphology applied to  
666 study physical mechanisms of soil pore development: An experiment using iron oxides and calcium  
667 carbonate', *Geoderma*. Elsevier B.V., 197–198, pp. 151–160. doi: 10.1016/j.geoderma.2013.01.008.

668 Giller, Paul S. (1996) 'The diversity of soil communities, the 'poor man's tropical rainforest'',  
669 *Biodiversity and Conservation*, 5, pp. 135–168. doi: <https://doi.org/10.1007/BF00055827>.

670 Grimm, V. *et al.* (2006) 'A standard protocol for describing individual-based and agent-based  
671 models', *Ecological Modelling*, 198(1–2), pp. 115–126. doi: 10.1016/j.ecolmodel.2006.04.023.

672 Grimm, V. *et al.* (2010) 'The ODD protocol: A review and first update', *Ecological Modelling*, 221(23),  
673 pp. 2760–2768.

674 Hartemink, A. E. and Minasny, B. (2014) 'Towards digital soil morphometrics', *Geoderma*. Elsevier  
675 B.V., 230–231, pp. 305–317. doi: 10.1016/j.geoderma.2014.03.008.

676 Hausser, J. and Strimmer, K. (2014) 'entropy: Estimation of Entropy, Mutual Information and Related  
677 Quantities.' R package version 1.2.1. <https://CRAN.R-project.org/package=entropy>

678 Helgason, B. L., Walley, F. L. and Germida, J. J. (2010) 'No-till soil management increases microbial  
679 biomass and alters community profiles in soil aggregates', *Applied Soil Ecology*. Elsevier B.V., 46(3),  
680 pp. 390–397. doi: 10.1016/j.apsoil.2010.10.002.

681 Hill, M. O. (1973) 'Diversity and evenness: A unifying notation and its consequences', *Ecology*, 54(2),  
682 pp. 427–432.

683 Kay M., Wobbrock J. (2020) 'ARTool: Aligned Rank Transform for Nonparametric Factorial ANOVAs.'  
684 doi: 10.5281/zenodo.594511

685 Kooijman, S. A. L. M. (2009) *Dynamic Energy Budget Theory for Metabolic Organisation*. 3rd edn.  
686 Cambridge, UK: Cambridge University Press.

687 Kravchenko, A. N. *et al.* (2011) 'Long-term Differences in Tillage and Land Use Affect Intra-aggregate  
688 Pore Heterogeneity', *Soil Science Society of America Journal*, 75(5), pp. 1658–1666. doi:  
689 10.2136/sssaj2011.0096.

690 Kravchenko, A. N. and Guber, A. K. (2017) 'Soil pores and their contributions to soil carbon  
691 processes', *Geoderma*. Elsevier B.V., 287, pp. 31–39. doi: 10.1016/j.geoderma.2016.06.027.

692 Kuznetsova, A., Brockhoff, P. B., Christensen, R. H. B. (2017) 'lmerTest Package: Tests in Linear Mixed  
693 Effects Models', *Journal of Statistical Software*, 82(13), pp. 1–26. doi: 10.18637/jss.v082.i13

694 Lenth, R. (2020). 'emmeans: Estimated Marginal Means, aka Least-Squares Means', R package  
695 version 1.5.0. <https://CRAN.R-project.org/package=emmeans>

696 Levins, R. (1966) 'The strategy of model-building in population biology', *American Scientist*. Sigma Xi,  
697 54(4), pp. 421–431. [www.jstor.org/stable/27836590](http://www.jstor.org/stable/27836590).

698 Lorscheid, I., Heine, B.-O. and Meyer, M. (2012) 'Opening the "black box" of simulations: increased  
699 transparency and effective communication through the systematic design of experiments"',  
700 *Computational and Mathematical Organization Theory*. Springer US, 18(1), pp. 22–62. doi:  
701 10.1007/s10588-011-9097-3.

702 Mangiafico, S. (2020) 'rcompanion: Functions to Support Extension Education Program Evaluation.' R  
703 package version 2.3.25. <https://CRAN.R-project.org/package=rcompanion>

704 Mones, E., Vicsek, L. and Vicsek, T. (2012) 'Hierarchy measure for complex networks', *PLoS ONE*,  
705 7(3), pp. 1–10. doi: 10.1371/journal.pone.0033799.

706 Nielson, U. N. *et al.* (2010) 'The enigma of soil animal species diversity revisited: The role of small-  
707 scale heterogeneity', *PLoS ONE*, 5(7), e11567. doi: 10.1371/journal.pone.0011567.

708 Oades, J. M. (1993) 'The role of biology in the formation, stabilization and degradation of soil  
709 structure', *Geoderma*, 56(1–4), pp. 377–400. doi: 10.1016/0016-7061(93)90123-3.

710 Pebesma, E. J. and Bivand, R. S. (2005) 'Classes and methods for spatial data in R', *R News*, 5(2).  
711 Available at: <https://cran.r-project.org/doc/Rnews/>.

712 Polhill, G. and Gimona, A. (2014) 'Using genetic algorithms to fit species and habitat parameters for  
713 modelling the effect of climate change on species distributions with stochastic patch occupancy  
714 models', *Proceedings - 7th International Congress on Environmental Modelling and Software: Bold  
715 Visions for Environmental Modeling, iEMSs 2014*, 3.

716 R Core Team (2020) 'R: A Language and Environment for Statistical Computing', *R Foundation for  
717 Statistical Computing*. Vienna, Austria, p. <https://www.R-project.org>. Available at: <http://www.r-project.org>.  
718

719 Rockafellar, R. T. (1970) *Convex Analysis*. Princeton, NJ: Princeton University Press.

720 Rodrigue, J. P. (2017) *The Geography of Transport Systems*. Hofstra University, Department of Global  
721 Studies & Geography. Available at: <https://transportgeography.org>.

722 Roshier, D.A., Doerr, V. A. J. and Doerr, E. D. D. (2008) 'Animal movement in dynamic landscapes:  
723 interaction between behavioural strategies and resource distributions', *Oecologia*, 156, pp. 465-477.  
724 doi: 10.1007/s00442-008-0987-0.

725 Spellerberg, I. F. and Fedor, P. J. (2003) 'A tribute to Claude Shannon (1916–2001) and a plea for  
726 more rigorous use of species richness, species diversity and the "Shannon–Wiener" Index', *Global  
727 Ecology and Biogeography*, 12, pp. 177–179. doi: 10.1046/j.1466-822X.2003.00015.x.

728 Stevens, R. D. and Tello, J. S. (2011) 'Diversity begets diversity: relative roles of structural and  
729 resource heterogeneity in determining rodent community structure', *Journal of Mammalogy*, 92(2),  
730 pp. 387–395. doi: 10.1644/10-mamm-a-117.1.

731 Sturges, H. A. (1926) 'The choice of a class interval', *Journal of the American Statistical Association*,  
732 21(153), pp. 65-66. doi: [10.1080/01621459.1926.10502161](https://doi.org/10.1080/01621459.1926.10502161).

733 Tews, J. *et al.* (2004) 'Animal species diversity driven by habitat heterogeneity/diversity: The  
734 importance of keystone structures', *Journal of Biogeography*, 31(1), pp. 79–92. doi: 10.1046/j.0305-  
735 0270.2003.00994.x.

736 Velde, B., Moreau, E. and Terribile, F. (1996) 'Pore networks in an Italian vertisol: Quantitative  
737 characterisation by two dimensional image analysis', *Geoderma*, 72(3–4), pp. 271–285. doi:  
738 10.1016/0016-7061(96)00033-X.

739 Vezzani, F. M. *et al.* (2018) 'The importance of plants to development and maintenance of soil

740 structure, microbial communities and ecosystem functions', *Soil and Tillage Research*. Elsevier,  
741 175(March 2017), pp. 139–149. doi: 10.1016/j.still.2017.09.002.

742 Wickham, H. (2016) *ggplot2: Elegant Graphics for Data Analysis*. New York: Springer-Verlag.  
743 Available at: <http://ggplot2.org>.

744 Wickham, H. *et al.* (2019) 'dplyr: A Grammar of Data Manipulation'.

745 Wijesinghe, D. K., John, E. A. and Hutchings, M. J. (2005) 'Does pattern of soil resource heterogeneity  
746 determine plant community structure? An experimental investigation', *Journal of Ecology*, 93(1), pp.  
747 99–112. doi: 10.1111/j.1365-2745.2004.00934.x.

748 Wobbrock, J., Findlater, L., Gergle, D., Higgins, J. (2011) 'The Aligned Rank Transform for  
749 Nonparametric Factorial Analyses Using Only ANOVA Procedures.' In *Proceedings of the ACM*  
750 *Conference on Human Factors in Computing Systems (CHI '11)*, pp. 143-146.

751 Young, I. M., Crawford, J. W. and Rappoldt, C. (2001) 'New methods and models for characterising  
752 structural heterogeneity of soil', *Soil and Tillage Research*, 61(1–2), pp. 33–45. doi: 10.1016/S0167-  
753 1987(01)00188-X.

754 Young, I. M. and Ritz, K. (2000) 'Tillage, habitat space and function of soil microbes', *Soil and Tillage*  
755 *Research*, 53(3–4), pp. 201–213. doi: 10.1016/S0167-1987(99)00106-3.

756 Young, I. M. and Ritz, K. (2009) 'The habitat of soil microbes', in *Biological Diversity and Function in*  
757 *Soils*, pp. 31–43.

758 Zeileis, A. (2014) 'ineq: Measuring Inequality, Concentration, and Poverty.' R package version 0.2-13.  
759 <https://CRAN.R-project.org/package=ineq>

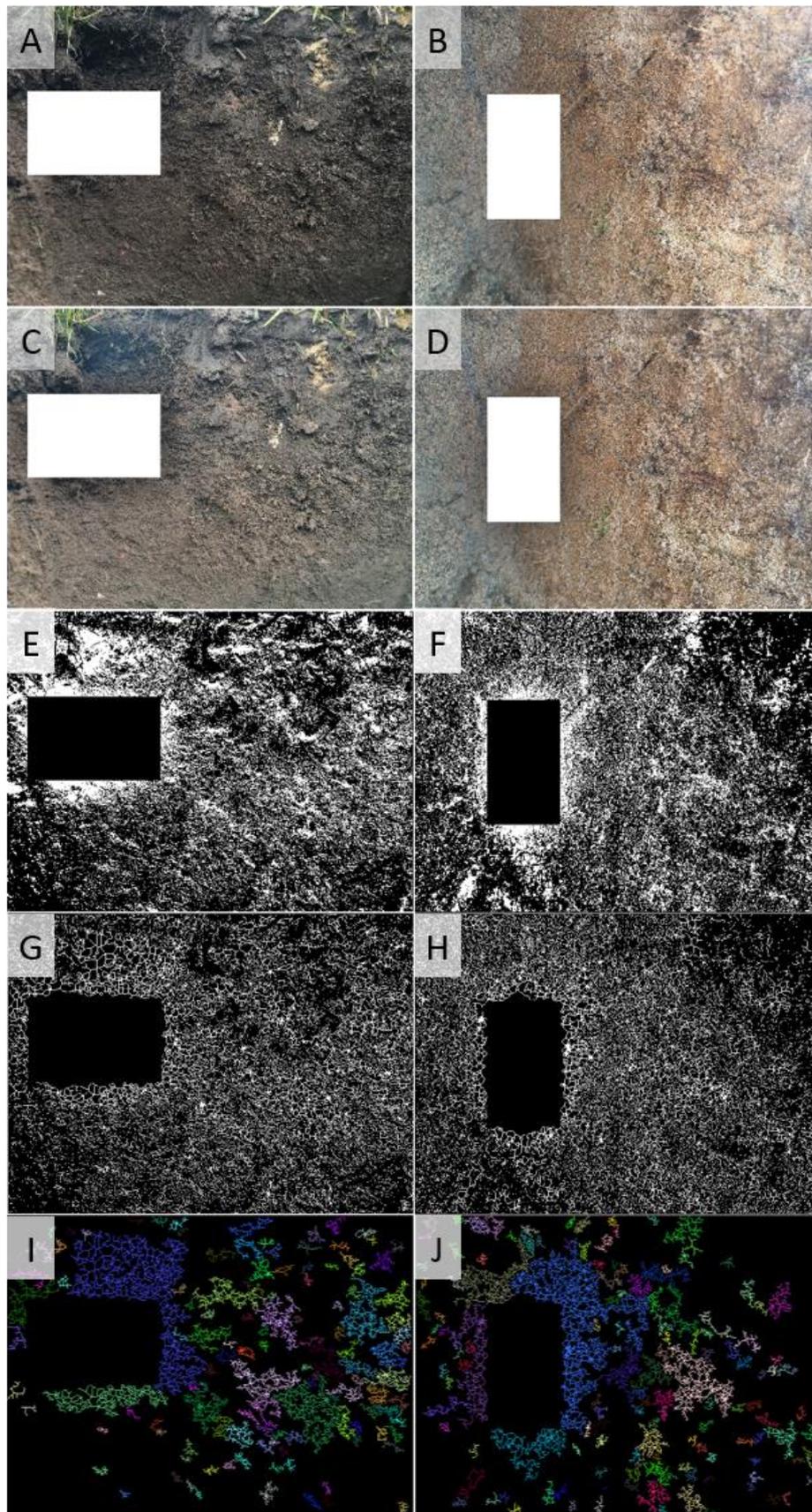
761 **Appendices**

762

763 **Appendix 1. Network extraction process**

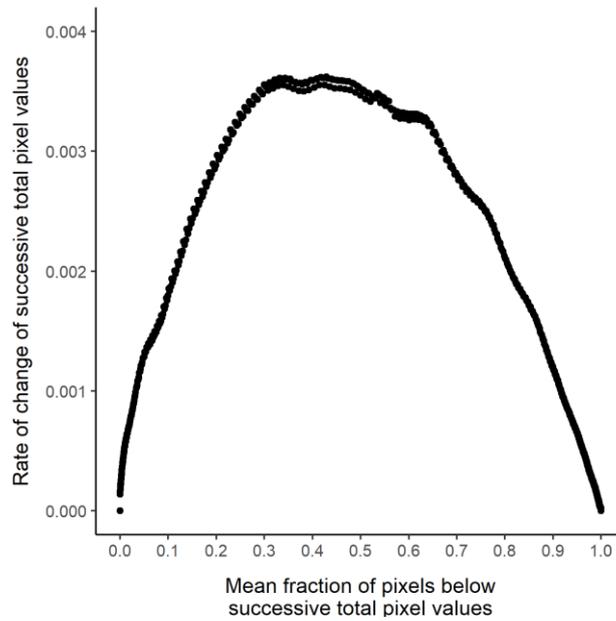
764 For all images:

- 765 1. Convert the image to a text file containing RGB triplets
- 766 2. Identify and eliminate all non-soil pixels (set to -1)
- 767 3. Calculate mean pixel intensity at all points
- 768 4. Adjust pixel intensity to remove variations in brightness across image
- 769 5. Threshold the image to retain the darkest 30 % soil pixels
- 770 6. Carry out erosion and thinning operators
- 771 7. Clean image to produce skeletal pixels
- 772 8. Identify networks
- 773 9. Remove redundant pathways
- 774 10. Calculate distances between nodes
- 775 11. Save the network



776

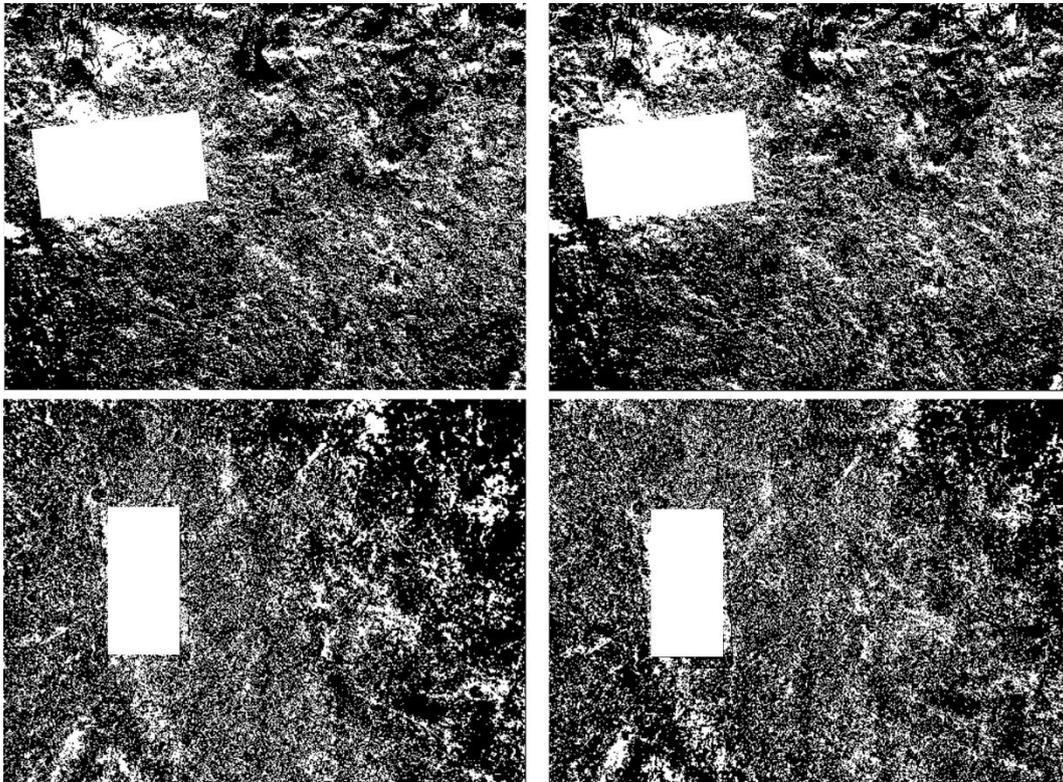
777 **Figure A1. Soil image morphology process for a Cambisol (a) and Arenosol (b) profile image.** Steps  
 778 show include (c-d) colour correction, (e-f) thresholding, (g-h) erosion and thinning operations, and (i-  
 779 j) subnetwork identification. White areas represent colour correction cards, which were excised.



780

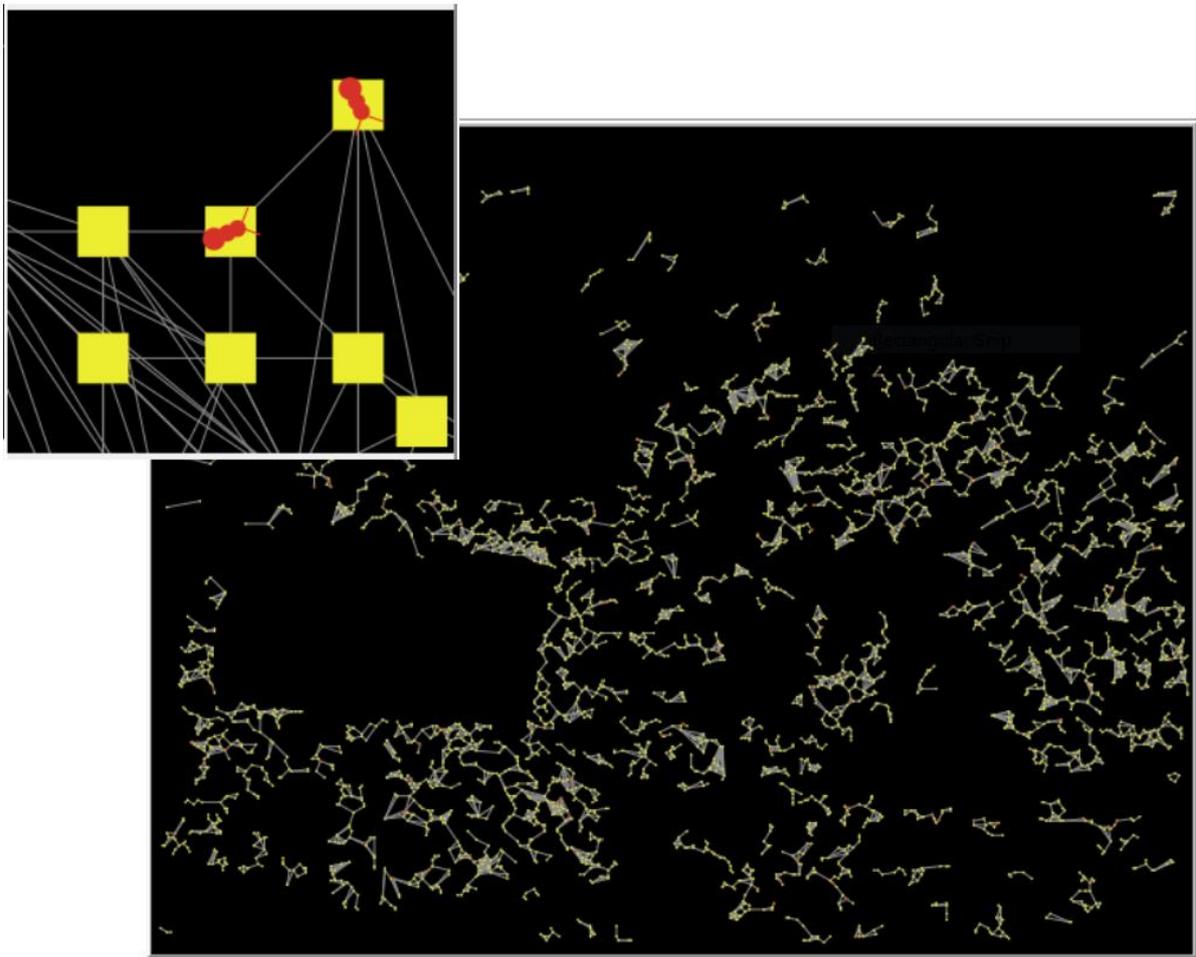
781 **Figure A2. The rate of change of mean fraction of pixels for each mean fraction below a given**  
 782 **threshold value.** The plot starts on the left with pixel values of 0, with no pixels below this value, and  
 783 ends on the right with pixel values of 755 (with correction card removed from image). The y-axis  
 784 shows the rate of change of the mean fraction of pixels below each value.

785



786

787 **Figure A3. Comparison of different images from the same pit after thresholding.** The two pairs of  
 788 images from each pit are arranged horizontally. The white rectangle is the correction card. The  
 789 thresholding process was the same as described in Section 2.2, where the darkest 30 % of pixels  
 790 have been retained as pores, and other pixels removed.



793  
 794 **Figure A4. A screenshot of the model.** The inset at the top left shows an enlarged version of some of  
 795 the resource nodes (yellow squares) and agents (red ants).

- 796 I. Model purpose  
 797 a. The model is designed to be an analytical tool to explore the heterogeneity in  
 798 resource supply potential of a network by populating it with idealised energy-  
 799 consuming agents, and to quantify the effects of consumer, resource, and network  
 800 characteristics on resulting consumer population outcomes.  
 801 II. Entities, state variables, and scales  
 802 a. Consumer entities  
 803 i. State variables

Property	Description
Location	The resource on which the consumer is located
Target location	The resource to which the consumer will move next
Active?	Whether a consumer is active (or dead)

806

ii. Parameters

Property	Description
Basal metabolism	How much resource an agent needs per day to stay alive
Active metabolism	How much resource an agent uses with each step
Resource stock	How much resource an agent has consumed but not metabolised
Consumption rate	Maximum number of resource units that an agent takes from a resource it visits, per timestep
Spawn energy	How much energy an agent requires to spawn (depletes this quantity from stocks and passed to offspring as starting quota)

807

808

b. Resource entities

809

i. State variables

Property	Description
Current supply	The current quantity of resource at this point
Resource capacity	How much energy is stored in a resource when it is full
Regrow rate	The amount the resource regrows each timestep

810

811

ii. Parameters

Property	Description
Resource capacity	How much energy is stored in a resource when it is full
Regrow rate	The amount the resource regrows each timestep

812

813

c. Link entities

814

i. State variables

Property	Description
Length	The length of the link - determines energy and time required to traverse it

815

816

d. Scales

Property	Description
Timestep	A single unit of time in the model, defined as that which is required for consumers to move 1 pixel (approximately 0.3 – 0.5 mm), and for which they require <code>basal-metabolism</code> units of energy.
World size	400 x 500, determined by the size of the soil networks used as the environment.

817

818

III. Sequence of events

819

a. Consumers start on random nodes around a pre-specified network, where nodes are resource patches.

820

821

b. Consumers move around the network randomly following links. If they find a resource patch, they consume as much as they can from it, and the patch depletes.

822

823

i. Consumers require `basal-metabolism` units of resource per timestep. If they do not consume this resource, they die.

824

- 825                   ii. Consumers can stay put on a resource and consume it (`consumption-`  
826                   `rate` units consumed per timestep), but it depletes, and if there is no more  
827                   resource there then they move on.
- 828                   iii. Consumers metabolise `active-metabolism` units of resource per patch  
829                   of link that they cross.
- 830                   iv. If there is more than one agent on a resource patch, they each take  
831                   `consumption-rate` units per timestep, or split the remainder if there is  
832                   not enough resource remaining for them to each get `consumption-rate`  
833                   units.
- 834                   c. If consumers have twice as much energy as the set `spawn-energy`, they can  
835                   spawn new consumers (who take the same amount of `resource-stock` from  
836                   their parent that the parent started with, so now parent and offspring both have the  
837                   same `resource-stock`).
- 838                   d. Resources regrow at a constant rate (`regrow-rate`) per timestep, up to their  
839                   maximum capacity (`resource-capacity`).
- 840                   IV. Design concepts
- 841                   a. Basic principles
- 842                   i. Consumers attempt to consume as much free energy from a resource as  
843                   they are able, to maximise energy reserves for future movement, and  
844                   spawning capability.
- 845                   ii. Conservation equations: energy and matter cannot be created (except at the  
846                   start of the simulation) or destroyed. In spawning, this is represented by  
847                   consumers transferring some of their energy to their offspring. Consumers  
848                   only die when their energy reserves are completely depleted (starvation).
- 849                   iii. Entropy production: some resource energy is consumed in movement and  
850                   cannot be recaptured.
- 851                   b. Emergence
- 852                   i. The distribution of consumers in space around the network and the  
853                   distribution of resource stocks across the consumers both emerge from the  
854                   interactions in the model.
- 855                   c. Objectives
- 856                   i. The consumers' objective is to consume as much resource energy as  
857                   possible, allowing them to stay alive, move, and potentially reproduce.
- 858                   d. Prediction
- 859                   i. Consumers do not 'predict' the results of their course of action per se, they  
860                   are random walkers, but they do 'predict' that they will die if they stay in a  
861                   non-resource patch, or depleted resource patch, so they keep moving.
- 862                   e. Sensing
- 863                   i. Consumers can sense if they are on a resource patch or not, and if it has any  
864                   resource energy in it. They also know the link-neighbours of the resource  
865                   patch that they are currently on.
- 866                   f. Learning
- 867                   i. Consumers are random walkers; they do not learn in any capacity.
- 868                   g. Adaptation
- 869                   i. The population adapts to fill the network in a way that reflects the density of  
870                   resource availability in that area, as consumers will cluster and reproduce  
871                   around resources where they can consume what they need.

- 872 h. Interaction
- 873 i. Consumers interact stigmergically through their consumption of resources.
- 874 While they do not interact directly in any meaningful way, their
- 875 consumption of resources affects the availability of resources for others to
- 876 consume.
- 877 i. Collectives
- 878 i. There are no collectives present.
- 879 j. Stochastic elements
- 880 i. Consumers are initialised in random locations and move randomly.
- 881 Additionally, resources are all initialised with random maximum capacity
- 882 between 1 and `maximum-resource-capacity` and regrow rates
- 883 between 1 and `maximum-regrow-rate`.
- 884 k. Observation
- 885 i. Number of currently active ('alive') consumers at each timestep.
- 886 ii. Mean, standard deviation (SD), Gini coefficient, and entropy of the
- 887 distribution of consumer resource stocks at each timestep.
- 888 iii. The resource capacity and regrow rate of each resource at the start of the
- 889 simulation.
- 890 iv. The resource stock and location of each active consumer at 10, 100, 500,
- 891 1000, and 2000 timesteps.
- 892 V. Initialisation
- 893 a. The network was supplied as two Comma-Separated Values (CSV) files: one of
- 894 resource node locations and another of the connections between the resource
- 895 nodes. The node locations and connections were determined during the process of
- 896 extracting the soil network from a soil profile image, as described in the main text
- 897 (Section 2.2). The resource and consumer types and parameters were specified in an
- 898 Extended Markup Language (XML) file. The models were initialised with 500
- 899 consumers located on random resource nodes throughout the network. The
- 900 consumers each began with 30 resource units in their `resource-stock`, and
- 901 metabolic rates, consumption rate, and spawn energy thresholds as specified in the
- 902 XML file. Resources were all initialised with random maximum capacity between 1
- 903 and `maximum-resource-capacity` and regrow rates between 1 and
- 904 `maximum-regrow-rate` and began the simulation at full capacity.
- 905 VI. Input data
- 906 a. This model has no input data.
- 907 VII. Submodels
- 908 a. Regrowth of resources: at each timestep, all resources that are less than their
- 909 maximum capacity regrow by `regrow-rate` units.
- 910 b. Consuming resources: at each timestep, all consumers currently located on a
- 911 resource node check whether there is any resource available at that node. If there is
- 912 enough for each consumer to take `consumption-rate` units, they do, and these
- 913 are added to their `resource-supply`. If there is not enough, each consumer
- 914 receives what is at the resource, divided by the number of consumers at the
- 915 resource. If there is no resource available at that node, the consumer identifies a
- 916 new `target-node`, selecting randomly from the other resources connected to the
- 917 first, and moves to the `target-node`.

- 918
- 919
- 920
- 921
- 922
- 923
- 924
- 925
- 926
- 927
- 928
- 929
- c. Spawning new consumers: at each timestep, consumers check whether they have twice the amount of energy specified as `spawn-energy` in their `resource-stock`. If so, they spawn a new consumer who is an exact clone of themselves. The new consumer starts with `spawn-energy` units as their initial `resource-supply`, and the parent consumer loses `spawn-energy` units of resource from their `resource-stock`.
  - d. Check consumer resource stocks: at each timestep, all consumers check whether they have more than `resource-requirement` units, or their basal metabolism, of resource in their `resource-supply`. If they do, they consume `resource-requirement` units, removing them from their `resource-supply`, otherwise they die.

930 **Appendix 3. Sensitivity Analysis**

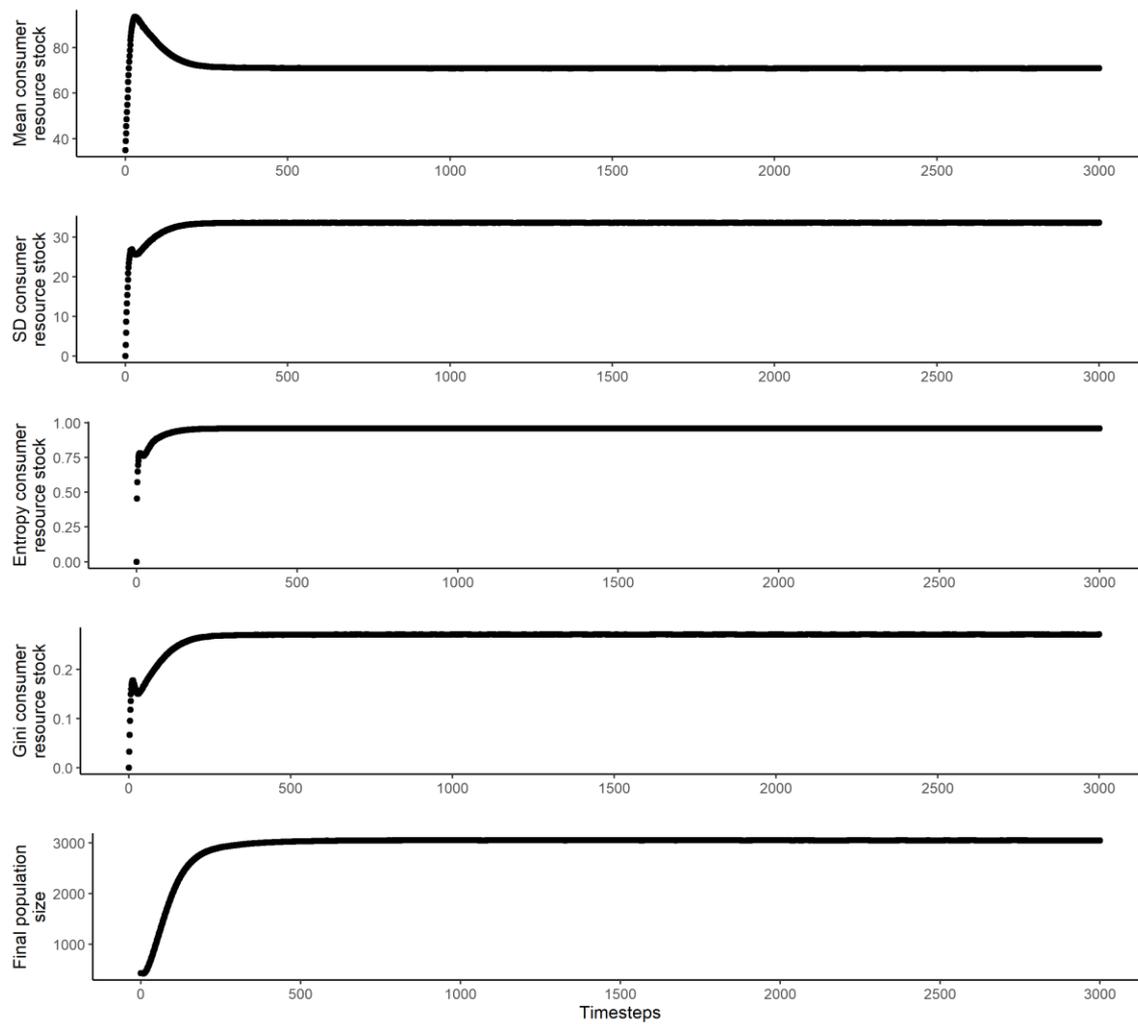
931 First, a pre-test was conducted to determine the number of time steps for which to run the  
932 simulations, and the number of replicates of each parameter set that were necessary for the outputs  
933 to reach equilibrium (ten Broeke, van Voorn and Ligtenberg, 2016). The first set of 500 runs used  
934 varied parameter values and a fixed network architecture, determined by Latin Hypercube Sampling  
935 from the range of values for global analysis (Table A1). One replicate of each parameter set was run  
936 for 3000 timesteps, and the output variables were plotted to determine whether the model reached  
937 a stable state, and if so, when. As all runs showed stability in output parameters after 500 – 1000  
938 timesteps (Fig. A5), apart from small variations due to stochasticity, the final output variable values  
939 for all future runs were calculated as the mean of the values at timesteps 500, 750, and 1000.

940

941 **Table A1. Parameter ranges used for testing to determine length of simulations.** Values shown are  
942 the minimum and maximum for that parameter. Latin Hypercube Sampling was used to generate the  
943 values, which were then multiplied by the range plus the minimum, to get the value for the  
944 parameter for testing.

<b>Parameter</b>	<b>Value</b>
Initial population size	50, 1000
Consumer basal metabolism	1, 3
Consumer active metabolism	1, 3
Initial consumer resource stock	20, 50
Consumer consumption rate	5, 10
Consumer spawn energy	50, 100
Maximum resource capacity	20, 50
Maximum resource regrowth rate	10, 20

945



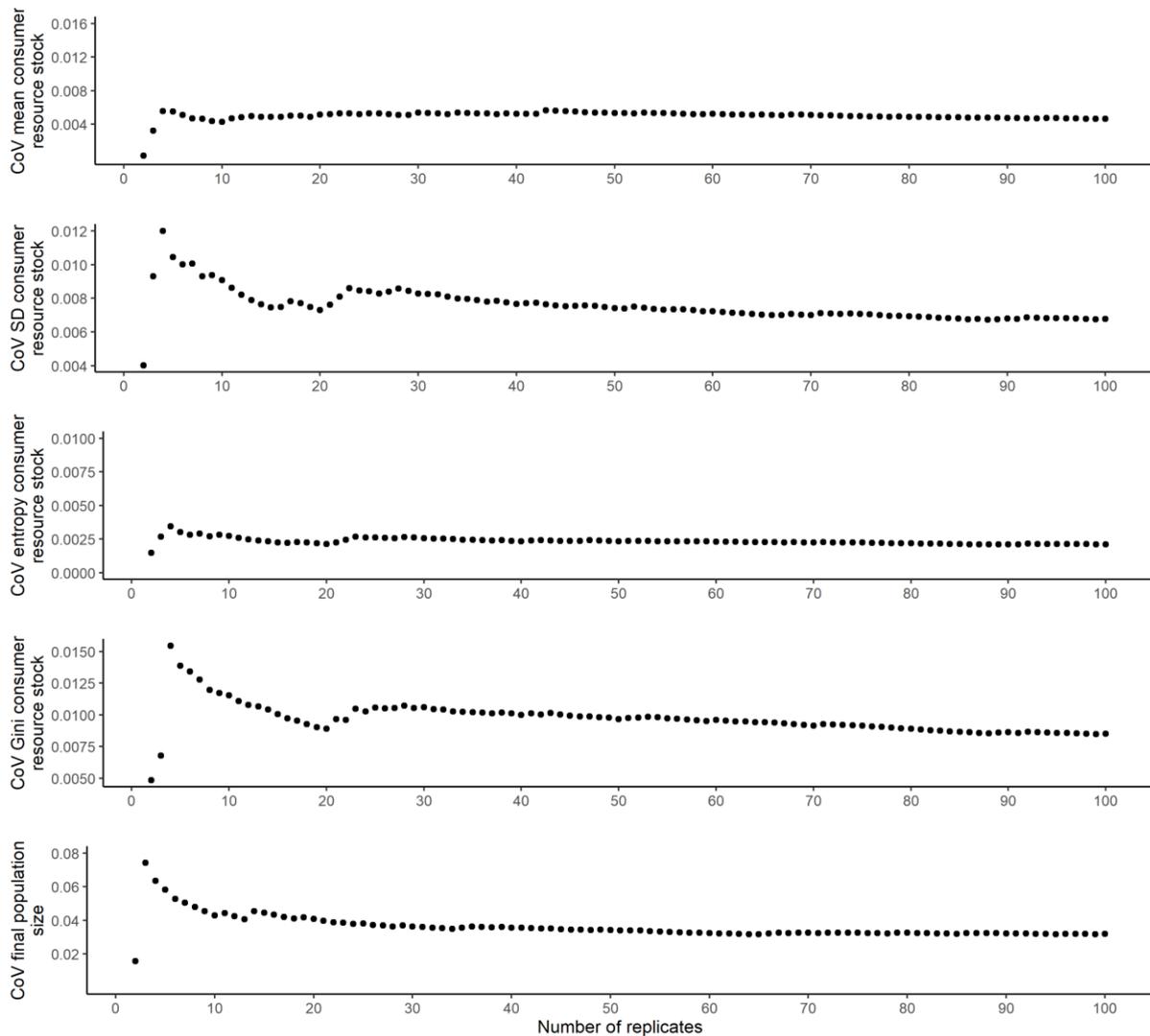
946

947 **Figure A5. Stability plots from testing to determine the length of simulations.** Shown are values  
 948 averaged for each timestep over 500 runs.

949

950 The second set of pre-test runs used the baseline parameter values for all parameters, and a fixed  
 951 network, which we repeated 100 times. We then calculated a rolling coefficient of variation for the  
 952 output variables, including progressively more replicates (Fig. A6). The coefficients of variation for all  
 953 output variables stabilised around 10 runs. Plotting the distribution of the output variables at that  
 954 point show approximate normality, such that the mean value across runs is a reasonable measure of  
 955 centre. Therefore, for all future simulations, the mean of the outcome variables across 10 replicates  
 956 was used to reduce the effects of stochasticity on the output. As the mean value across replicates  
 957 was used, there was no effect from replication on the experimental results.

958



959

960 **Figure A6. Plots of rolling Coefficient of Variation (CoV) for each outcome variable against the**  
 961 **number of replicates included in its calculation.** This was used to determine number of replicates  
 962 needed to average across to minimise stochasticity in output variables.

963

964 After the pre-test, we used the One-Factor-at-a-Time methodology to identify which of the control  
 965 variables significantly affected the output variables, and which could be held constant. For this test,  
 966 the four control variables (initial consumer population, initial consumer resource stock, maximum  
 967 resource regrowth rate, and maximum resource capacity) were varied across four levels each,  
 968 changing one variable at a time, while holding all other variables constant at middle values for each.  
 969 Both the maximum resource regrowth rate and maximum resource capacity significantly affected the  
 970 output variables, while initial consumer resource stock did not (Table A2). The initial consumer  
 971 population size significantly affected all but the standard deviation of consumer resource stock  
 972 (Table A2e), but as the magnitude of the effect was quite small, both the initial population size and  
 973 initial consumer resource stock were held constant at middle values for the rest of the simulations.

974

975 **Table A2. Regression results from One-Factor-at-a-Time analysis.** This was used to identify which  
 976 control parameters could be fixed, and which significantly affected the outcome variables and  
 977 needed to be explored. The asterisks designate level of significance:  $p < 0.1$ : ·,  $p < 0.05$ : \*,  $p < 0.01$ :  
 978 \*\*,  $p < 0.001$ : \*\*\*.

**a. Mean consumer resource stock**

	Estimate	Standard error	t value	p
Intercept	54.360	0.100	543.566	0.000 ***
Initial population size	0.000	0.000	-3.728	0.002 ***
Initial consumer resource stock	0.000	0.001	0.100	0.920
Maximum resource regrow rate	0.054	0.004	12.874	0.000 ***
Maximum resource capacity	0.060	0.001	41.088	0.000 ***
F(4, 251) = 467 ( $p < 0.001$ )		R <sup>2</sup> = 0.88		

**b. SD consumer resource stock**

	Estimate	Standard error	t value	p
Intercept	27.110	0.038	711.283	0.000 ***
Initial population size	0.000	0.000	0.948	0.344
Initial consumer resource stock	0.000	0.000	0.333	0.739
Maximum resource regrow rate	0.016	0.002	10.073	0.000 ***
Maximum resource capacity	-0.010	0.000	-17.718	0.000 ***
F(4, 251) = 104.1 ( $p < 0.001$ )		R <sup>2</sup> = 0.62		

979

**c. Entropy consumer resource stock**

	Estimate	Standard error	t value	p
Intercept	0.953	0.000	2288.704	0.000 ***
Initial population size	0.000	0.000	6.500	0.000 ***
Initial consumer resource stock	0.000	0.000	0.446	0.656
Maximum resource regrow rate	0.000	0.000	19.891	0.000 ***
Maximum resource capacity	0.000	0.000	2.891	0.004 **
F(4, 251) = 111.6 ( $p < 0.001$ )		R <sup>2</sup> = 0.63		

980

**d. Gini consumer resource stock**

	Estimate	Standard error	t value	p
Intercept	0.283	0.000	431.434	0.000 ***
Initial population size	0.000	0.000	3.472	0.001 ***
Initial consumer resource stock	0.000	0.000	0.154	0.876
Maximum resource regrow rate	0.000	0.000	-2.267	0.024*
Maximum resource capacity	0.000	0.000	-38.517	0.000 ***
F(4, 251) = 375.2 ( $p < 0.001$ )		R <sup>2</sup> = 0.85		

**e. Final population size**

	<b>Estimate</b>	<b>Standard error</b>	<b>t value</b>	<b>p</b>
Intercept	-	188.701	-14.773	0.000
Initial population size	2787.749	0.118	21.110	0.000
Initial consumer resource stock	2.496	2.738	0.215	0.830
Maximum resource regrow rate	0.589	8.038	23.805	0.000
Maximum resource capacity	191.341	2.738	22.634	0.000
F(4, 251) = 381.2 ( $p < 0.001$ )				R <sup>2</sup> = 0.86

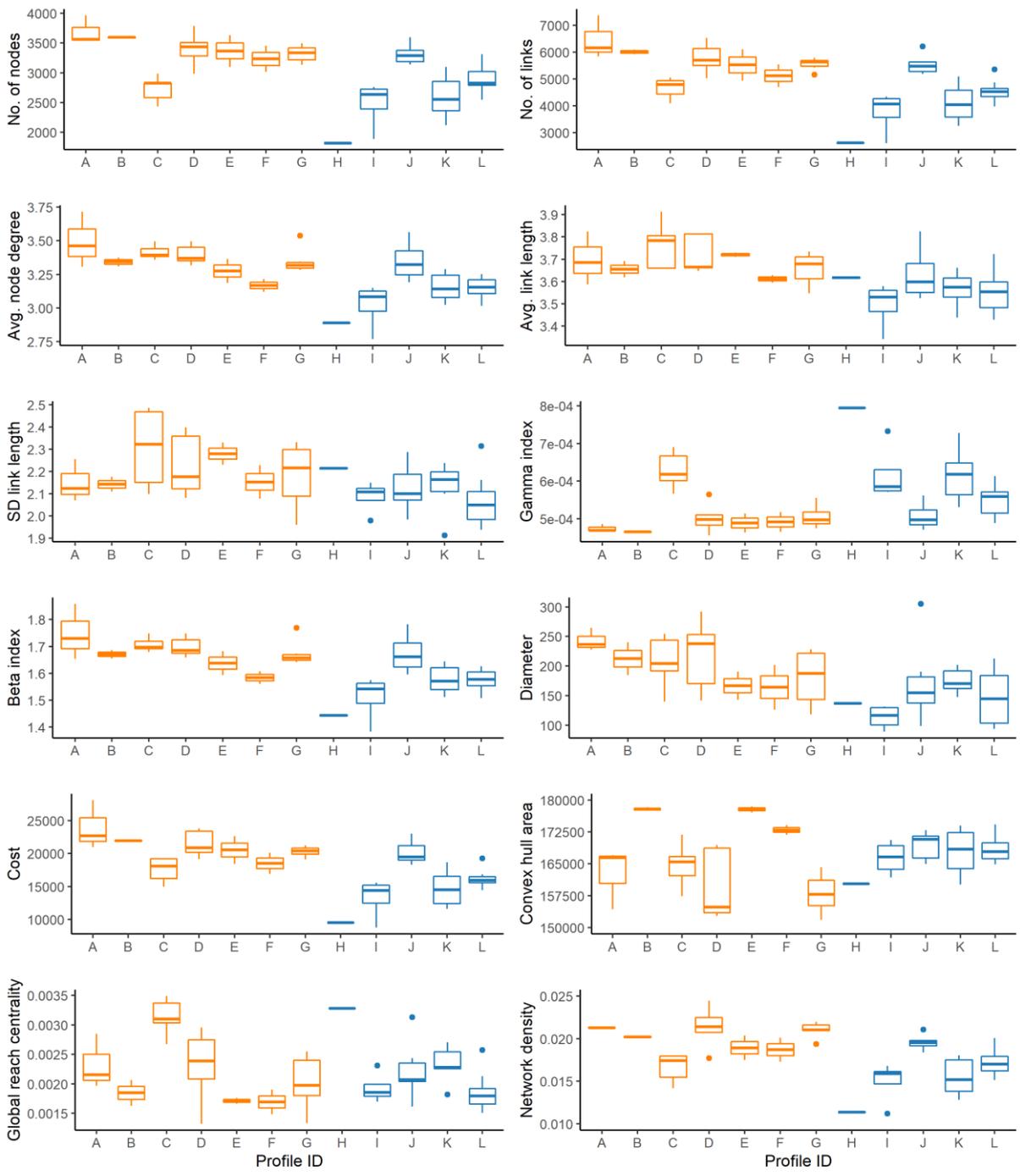
991 **Appendix 4. Calculation of Entropy**

992 The entropy of the consumer resource stocks was calculated as the Shannon index, or Shannon  
 993 entropy, of the resource stocks held by consumers. As the Shannon entropy is meant to be applied  
 994 to discrete data, the consumer resource stocks were discretised into a fixed number of ‘bins’ using  
 995 Sturges’ formula (Sturges, 1926), and the Shannon entropy was calculated for the bins.

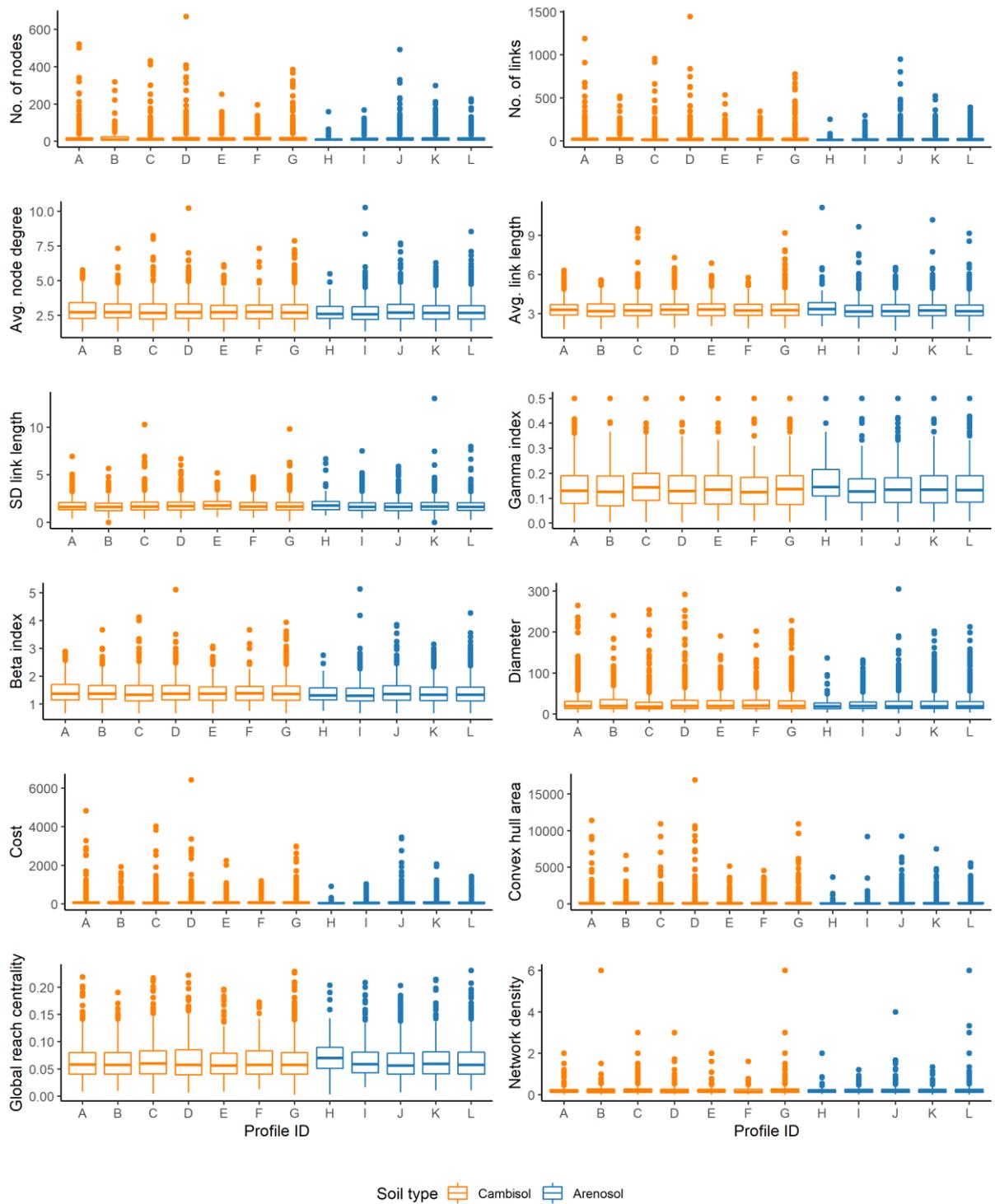
996 Sturges’ formula for the number of bins  $k$  for a population of size  $n$  is

997  $k = \lceil \log_2 n \rceil + 1.$  Eq. A1

998 Using a sample of 100 runs from the stability test for run length (Section A3), the normality of the  
 999 consumer resource stocks at the sampling timesteps  $T = 500$ ,  $T = 750$ , and  $T = 1000$  was tested.  
 1000 Additionally, the entropy was calculated for 5, 10, 15, 20, 25, 50, 75, and 100 bins and compared  
 1001 with the entropy binned using Sturges’ formula. By normalising the calculated entropy by the  
 1002 maximum possible entropy for that number of bins,  $\log(N)$ , the differences in entropy between  
 1003 different numbers of bins were  $< 0.001$ . As the data were found to be approximately normally  
 1004 distributed at the sampling timesteps, the assumptions for Sturges’ formula was met, and it was  
 1005 chosen to determine the final bin width.



Soil type ▭ Cambisol ▭ Arenosol



1008

**b.**

1009

**Figure A7. Boxplots showing distributions of network metrics across soil profiles for (a) main**

1010

**networks and (b) subnetworks.** Profiles A – G correspond to Cambisol soil profiles, and profiles H – L

1011

are Arenosol soil profiles. Descriptions of network metrics are in Table 1.